# Package 'GJRM'

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Title Generalised Joint Regression Modelling
<b>Description</b> Routines for fitting various joint (and univariate) regression models, with several types of covariate effects, in the presence of equations' errors association, endogeneity, nonrandom sample selection or partial observability.
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## **Description**

This package provides a function for fitting various generalised joint regression models with several types of covariate effects and distributions. Many modelling options are supported and all parameters of the joint distribution can be specified as flexible functions of covariates.

The original name of this package was SemiParBIVProbit which was designed to fit flexible bivariate binary response models. However, since then the package has expanded so much that its original name no longer gave a clue about all modelling options available. The new name should more closely reflect past, current and future developments.

The main fitting functions are listed below.

gjrm() which fits bivariate regression models with binary responses (useful for fitting bivariate binary models in the presence of (i) non-random sample selection or (ii) associated responses/endogeneity or (iii) partial observability), bivariate models with binary/discrete/continuous/survival margins in the presence of associated responses/endogeneity, bivariate sample selection models with continuous/discrete response, trivariate binary models (with and without double sample selection). This function essentially merges all previously available fitting functions, namely SemiParBIV(), SemiParTRIV(), copulaReg() and copulaSampleSel().

gamlss() fits flexible univariate regression models where the response can be binary (only the extreme value distribution is allowed for), continuous, discrete and survival. The purpose of this function was only to provide, in some cases, starting values for the above functions, but it has now been made available in the form of a proper function should the user wish to fit univariate models using the general estimation approach of this package.

We are currently working on several multivariate extensions.

#### **Details**

GJRM provides functions for fitting general joint models in various situations. The estimation approach is based on a very generic penalized maximum likelihood based framework, where any (parametric) distribution can in principle be employed, and the smoothers (representing several types of covariate effects) are set up using penalised regression splines. Several marginal and copula distributions are available and the numerical routine carries out function minimization using a trust region algorithm in combination with an adaptation of an automatic multiple smoothing parameter estimation procedure for GAMs (see mgcv for more details on this last point). The smoothers supported by this package are those available in mgcv.

Confidence intervals for smooth components and nonlinear functions of the model parameters are derived using a Bayesian approach. P-values for testing individual smooth terms for equality to the zero function are also provided and based on the approach implemented in mgcv. The usual plotting and summary functions are also available. Model/variable selection is also possible via the use of shrinakge smoothers and/or information criteria.

#### Author(s)

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with contributions from Panagiota Filippou (specifically on the trivariate binary models), Francesco Donat (on the bivariate models with ordinal and continuous margins, and that with ordinal margins), Matteo Fasiolo (for the implementation of the pdf and cdf, and related derivatives, of the Tweedie

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distribution), Alessia Eletti (on survival models with mixed censoring and excess hazards), Kiron Das (for help with the implementation of the Galambos copula), Eva Cantoni and William Aeberhard (for help with the implementation of the robust gamlss), Alessia Eletti and Danilo Petti (for help with the implementation of the copula survival model with general censoring scheme).

Thanks to Bear Braumoeller for suggesting the implementation of bivariate models with partial observability, and Carmen Cadarso for suggesting the inclusion of various modelling extensions.

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#### References

Submitted papers (developments available from release 0.2-5):

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## See Also

gjrm, gamlss

adjCov

Adjustment for the covariance matrix from a fitted gjrm model

## Description

adjCov can be used to adjust the covariance matrix of a fitted gjrm object.

#### Usage

adjCov(x, id)

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## Arguments

A fitted gjrm object as produced by the respective fitting function.

id Cluster identifier.

#### **Details**

This adjustment can be made when dealing with clustered data and the cluster structure is neglected when fitting the model. The basic idea is that the model is fitted as though observations were independent, and subsequently adjust the covariance matrix of the parameter estimates. Using the terminology of Liang and Zeger (1986), this would correspond to using an independence structure within the context of generalized estimating equations. The parameter estimators are still consistent but are inefficient as compared to a model which accounts for the correct cluster dependence structure. The covariance matrix of the independence estimators can be adjusted as described in Liang and Zeger (1986, Section 2).

#### Value

This function returns a fitted object which is identical to that supplied in adjCov but with adjusted covariance matrix.

#### WARNINGS

This correction may not be appropriate for models fitted using penalties.

## Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

#### References

Liang K.-Y. and Zeger S. (1986), Longitudinal Data Analysis Using Generalized Linear Models. *Biometrika*, 73(1), 13-22.

#### See Also

GJRM-package, gjrm

adjCovSD	Adjustment for the covariance matrix from a gjrm model fitted to com-
	plex survey data.

## **Description**

adjCovSD can be used to adjust the covariance matrix of a fitted gjrm object.

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#### Usage

```
adjCovSD(x, design)
```

## **Arguments**

x A fitted gjrm object as produced by the respective fitting function.

design A svydesign object as produced by svydesign() from the survey package.

#### **Details**

This function has been extracted from the survey package and adapted to the class of this package's models. It computes the sandwich variance estimator for a copula model fitted to data from a complex sample survey (Lumley, 2004).

#### Value

This function returns a fitted object which is identical to that supplied in adjCovSD but with adjusted covariance matrix.

## WARNINGS

This correction may not be appropriate for models fitted using penalties.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### References

Lumley T. (2004), Analysis of Complex Survey Samples. *Journal of Statistical Software*, 9(8), 1-19

## See Also

GJRM-package, gjrm

AT Average treatment effect of a binary/continuous/discrete endogenous variable

## **Description**

AT can be used to calculate the treatment effect of a binary/continuous/discrete endogenous predictor/treatment, with corresponding interval obtained using posterior simulation.

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## Usage

```
AT(x, nm.end, eq = NULL, E = TRUE, treat = TRUE, type = "joint", ind = NULL, percentage = FALSE, n.sim = 100, prob.lev = 0.05, length.out = NULL, hd.plot = FALSE, te.plot = FALSE, main = "Histogram and Kernel Density of Simulated Average Effects", xlab = "Simulated Average Effects", ...)
```

## **Arguments**

guments	
x	A fitted gjrm object as produced by the respective fitting function.
nm.end	Name of the endogenous variable.
eq	Number of equation containing the endogenous variable. This is only used for trivariate models.
Е	If TRUE then AT calculates the sample ATE. If FALSE then it calculates the sample AT for the treated individuals only.
treat	If TRUE then AT calculates the AT using the treated only. If FALSE then it calculates the effect on the control group. This only makes sense if E = FALSE.
type	This argument can take three values: "naive" (the effect is calculated ignoring the presence of observed and unobserved confounders), "univariate" (the effect is obtained from the univariate model which neglects the presence of unobserved confounders) and "joint" (the effect is obtained from the simultaneous model which accounts for observed and unobserved confounders).
ind	Binary logical variable. It can be used to calculate the AT for a subset of the data. Note that it does not make sense to use ind when some observations are excluded from the AT calculation (e.g., when using E = FALSE).
percentage	Only for the Roy model, when TRUE it provides results in terms of percentage.
n.sim	Number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used when delta = FALSE. It may be increased if more precision is required.
prob.lev	Overall probability of the left and right tails of the AT distribution used for interval calculations.
length.out	Ddesired length of the sequence to be used when calculating the effect that a continuous/discrete treatment has on a binary outcome.
hd.plot	If TRUE then a plot of the histogram and kernel density estimate of the simulated average effects is produced. This can only be produced when when binary responses are used.
te.plot	For the case of continuous/discrete endogenous variable and binary outcome, if TRUE then a plot showing the treatment effects that the binary outcome is equal to 1 for each incremental value of the endogenous variable and respective intervals is produced.
main	Title for the plot.
xlab	Title for the x axis.
• • •	Other graphics parameters to pass on to plotting commands. These are used only when hd.plot = TRUE.

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#### **Details**

AT measures the average difference in outcomes under treatment (the binary predictor or treatment assumes value 1) and under control (the binary treatment assumes value 0). Posterior simulation is used to obtain a confidence/credible interval. See the references below for details.

AT can also calculate the effect that a continuous/discrete endogenous variable has on a binary outcome. In this case the effect will depend on the unit increment chosen (as shown by the plot produced).

#### Value

res It returns three values: lower confidence interval limit, estimated AT and upper

interval limit.

prob.lev Probability level used.

sim.AT It returns a vector containing simulated values of the average treatment effect.

This is used to calculate intervals.

Effects For the case of continuous/discrete endogenous variable and binary outcome,

it returns a matrix made up of three columns containing the effects for each

incremental value in the endogenous variable and respective intervals.

#### Author(s)

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#### References

Marra G. and Radice R. (2011), Estimation of a Semiparametric Recursive Bivariate Probit in the Presence of Endogeneity. *Canadian Journal of Statistics*, 39(2), 259-279.

#### See Also

GJRM-package, gjrm

BCDF Internal Function		
	BCDF	Internal Function

#### **Description**

It evaluates the cdf of several copulae.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

bcont 11

bcont Internal Function

## Description

This and other similar internal functions provide the log-likelihood, gradient and observed information matrix for penalized/unpenalized maximum likelihood optimization when copula models with continuous margins are employed.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

bdiscrcont

Internal Function

## Description

This and other similar internal functions provide the log-likelihood, gradient and observed information matrix for penalized/unpenalized maximum likelihood optimization when copula models with discrete and continuous margins are employed.

## Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

bdiscrdiscr

Internal Function

## Description

This and other similar internal functions provide the log-likelihood, gradient and observed information matrix for penalized/unpenalized maximum likelihood optimization when copula models with discrete margins are employed.

#### Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

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## **Description**

It provides the log-likelihood, gradient and observed/Fisher information matrix for penalized/unpenalized maximum likelihood optimization when copula models with binary outcomes are employed.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

gHsCont Internal Function	

## Description

It provides the log-likelihood, gradient and observed information matrix for penalized/unpenalized maximum likelihood optimization when copula models with binary and continuous margins are employed.

#### Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

bprobgHsContSS	Internal Function		
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## **Description**

It provides the log-likelihood, gradient and observed information matrix for penalized/unpenalized maximum likelihood optimization when copula sample selection models with continuous margins are employed.

## Author(s)

Maintainer: Giampiero Marra <giampiero.marra@ucl.ac.uk>

bprobgHsContUniv 13

bprobgHsContUniv	Internal Function
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## **Description**

It provides the log-likelihood, gradient and observed information matrix for penalized/unpenalized maximum likelihood optimization when fitting univariate models with discrete/continuous response.

#### Author(s)

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bprobgHsDiscr1	Internal Function	

## Description

It provides the log-likelihood, gradient and observed information matrix for penalized/unpenalized maximum likelihood optimization when copula models with binary and discrete margins are employed.

## Author(s)

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bprobgHsDiscr1SS	Internal Function	

## **Description**

It provides the log-likelihood, gradient and observed information matrix for penalized/unpenalized maximum likelihood optimization when copula sample selection models with discrete margins are employed.

## Author(s)

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bprobgHsP0

Internal Function

## Description

It provides the log-likelihood, gradient and observed or expected information matrix for penalized/unpenalized maximum likelihood optimization when bivariate probit models with partial observability are employed.

## Author(s)

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bprobgHsSS

Internal Function

## **Description**

It provides the log-likelihood, gradient and observed/Fisher information matrix for penalized/unpenalized maximum likelihood optimization when copula sample selection models with binary outcomes are employed.

## Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

conv.check

Some convergence diagnostics

## **Description**

It takes a fitted model object and produces some diagnostic information about the fitting procedure.

## Usage

```
conv.check(x)
```

#### **Arguments**

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gjrm object.

copgHs 15

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

## See Also

gamlss, gjrm

copgHs

Internal Function

## Description

This and other similar internal functions evaluate the first and second derivatives with respect to the margins and association parameter of several copulae.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

CopulaCLM

Internal fitting function

## Description

Internal fitting and set up function.

## Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

copulaSampleSel

Internal fitting function

## Description

Internal fitting and set up function.

## Author(s)

Maintainer: Giampiero Marra <giampiero.marra@ucl.ac.uk>

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cv.inform

Cross validation for informative censoring univariate survival models

#### **Description**

cv. inform carries out cross validation to help choosing the set of informative covariates.

## Usage

```
cv.inform(x, K = 5, data, informative = "yes")
```

## **Arguments**

x A fitted gamlss object as produced by the respective fitting function.

K No. of folds.

data Data.

informative If no then cv is carried out for the case of no informative censoring. This is

useful for comparison purposes.

#### **Details**

cv. inform carries out cross validation to help choosing the set of informative covariates.

## Value

Sl Overall sum of predicted likelihood contributions.

## Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

#### See Also

GJRM-package, gamlss

distrHs

Internal Function

## **Description**

This and other similar internal functions evaluate the margins' derivatives needed in the likelihood function for the binary, discrete and continuous cases.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

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Dpens

Differentiable penalties

## Description

work in progress, temp function

## Usage

## **Arguments**

params coefficients.

type lasso, alasso or scad.lambda smoothing parameter.

w.alasso for alasso.gamma default 1.a for scad.eps tolerance.

## **Details**

work in progress.

## Value

The function returns a penalty.

eta.tr

Internal Function

## **Description**

This and other similar internal functions map certain key quantities into a feasible parameter space. Some functions carry out some general consistency checks.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

g.tri

Internal Function

#### **Description**

This and other similar internal functions calculate the score for trivariate binary models.

#### Author(s)

Author: Panagiota Filippou

Maintainer: Giampiero Marra < giampiero . marra@ucl.ac.uk>

gamlss

Generalised Additive Models for Location, Scale and Shape

#### **Description**

gamlss fits flexible univariate regression models with several continuous and discrete distributions, and types of covariate effects. The purpose of this function was only to provide, in some cases, starting values for the simultaneous models in the package, but it has now been made available in the form of a proper function should the user wish to fit univariate models using the general estimation approach of this package. The distributions implemented here have been parametrised according to Rigby and Stasinopoulos (2005).

#### Usage

```
gamlss(formula, data = list(), weights = NULL, subset = NULL,
    margin = "N", surv = FALSE, cens = NULL, type.cens = "R", upperB = NULL,
    robust = FALSE, rc = 3, lB = NULL, uB = NULL, infl.fac = 1,
    rinit = 1, rmax = 100, iterlimsp = 50, tolsp = 1e-07,
    gc.l = FALSE, parscale, extra.regI = "t", gev.par = -0.25,
    chunk.size = 10000, k.tvc = 0, knots = NULL,
    informative = "no", inform.cov = NULL, margin2 = "PH",
    fp = FALSE, sp = NULL,
    drop.unused.levels = TRUE, siginit = NULL, shinit = NULL,
    sp.method = "perf", hrate = NULL, d.lchrate = NULL, d.rchrate = NULL,
    d.lchrate.td = NULL, d.rchrate.td = NULL, truncation.time = NULL,
    min.dn = 1e-40, min.pr = 1e-16, max.pr = 0.9999999, ygrid.tol = 1e-08)
```

#### **Arguments**

formula

List of equations. This should contain one or more equations.

data

An optional data frame, list or environment containing the variables in the model. If not found in data, the variables are taken from environment(formula), typically the environment from which gamlss is called.

weights Optional vector of prior weights to be used in fitting.

subset Optional vector specifying a subset of observations to be used in the fitting pro-

cess.

margin Possible distributions are normal ("N"), log-normal ("LN"), Gumbel ("GU"), re-

verse Gumbel ("rGU"), generelised Pareto ("GP"), generelised Pareto II ("GPII") where the shape parameter is forced to be > -0.5, generelised Pareto (with orthogonal parametrisation) ("GPo") where the shape parameter is forced to be > -0.5, discrete generelised Pareto ("DGP"), discrete generelised Pareto II ("DG-PII") where the shape parameter is forced to be positive, discrete generelised Pareto derived under the scenario in which shape = 0 ("DGPO"), logistic ("LO"), Weibull ("WEI"), inverse Gaussian ("iG"), gamma ("GA"), Dagum ("DAGUM"), Singh-Maddala ("SM"), beta ("BE"), Fisk ("FISK", also known as log-logistic distribution), Poisson ("PO"), zero truncated Poisson ("ZTP"), negative binomial - type I ("NBI"), negative binomial - type II ("NBII"), Poisson inverse Gaussian ("PIG"), generalised extreme value link function ("GEVlink", this is used for binary responses and is more stable and faster than the R package

bgeva).

surv If TRUE then a survival model is fitted. Here margin can be "PH" (generalised

proportional hazards), "PO" (generalised proportional odds), "probit" (gener-

alised probit).

cens This is required when surv = TRUE. When type.cens is different from mixed,

this variable can be equal to 1 if the event occurred and 0 otherwise. If type.cens = "mixed" then cens is a mixed factor variable (made up of four possible cate-

gories: I for interval, L for left, R for right, and U for uncensored.

type.cens Type of censoring mechanism. This can be "R", "L", "I" or "mixed".

upperB Variable name of right/upper bound when type.cens = "I" or type.cens =

"mixed" and interval censoring is present.

robust If TRUE then the robust version of the model is fitted.

rc Robust constant.

1B, uB Bounds for integral in robust case.

infl. fac Inflation factor for the model degrees of freedom in the approximate AIC. Smoother

models can be obtained setting this parameter to a value greater than 1.

rinit Starting trust region radius. The trust region radius is adjusted as the algorithm

proceeds.

rmax Maximum allowed trust region radius. This may be set very large. If set small,

the algorithm traces a steepest descent path.

iterlimsp A positive integer specifying the maximum number of loops to be performed

before the smoothing parameter estimation step is terminated.

tolsp Tolerance to use in judging convergence of the algorithm when automatic smooth-

ing parameter estimation is used.

gc.1 This is relevant when working with big datasets. If TRUE then the garbage collec-

tor is called more often than it is usually done. This keeps the memory footprint

down but it will slow down the routine.

parscale The algorithm will operate as if optimizing objfun(x / parscale, ...) where

parscale is a scalar. If missing then no rescaling is done. See the documen-

tation of trust for more details.

extra.regI If "t" then regularization as from trust is applied to the information matrix if

needed. If different from "t" then extra regularization is applied via the options "pC" (pivoted Choleski - this will only work when the information matrix is semi-positive or positive definite) and "sED" (symmetric eigen-decomposition).

gev.par GEV link parameter.

chunk.size This is used for discrete robust models.

k.tvc Experimental. Only used for tvc ps smoothers when using survival models.

knots Optional list containing user specified knot values to be used for basis construc-

tion.

informative If "yes" then informative censoring is assumed when using a survival model.

inform.cov If above is "yes" then a set of informative covariates must be provided.

margin2 In the informative survival case, the margin for the censored equation can be

different from that of the survival equation.

fp If TRUE then a fully parametric model with unpenalised regression splines if

fitted.

sp A vector of smoothing parameters can be provided here. Smoothing parame-

ters must be supplied in the order that the smooth terms appear in the model

equation(s).

drop.unused.levels

By default unused levels are dropped from factors before fitting. For some smooths involving factor variables this may have to be turned off (only use if

you know what you are doing).

siginit, shinit

For the GP and DGP distributions, initial values for sigma and shape may be

provided.

sp. method Multiple smoothing automatic parameter selection is perf. efs is an alternative

and only sensible option for robust models.

hrate Vector of population hazard rates computed at time of death of each uncensored

patient. The length of hrate should be equal to the number of uncensored observations in the dataset. Needed in the context of excess hazard modelling when uncensored observations are present. Note that this includes left truncated

uncensored observations as well.

d.lchrate Vector of differences of population cumulative excess hazards computed at the

age of the patient when the left censoring occurred and at the initial age of the patient. The length of d.lchrate should be equal to the number of left and/or interval censored observations in the dataset. Needed in the context of excess hazard modelling if left censored and/or interval censored observations

are present. In the latter case, d.rchrate also need be provided.

d.rchrate Vector of differences of population cumulative excess hazards computed at the

age of the patient when the at the right interval censoring time and at the initial age of the patient. The length of d.rchrate should be equal to the number of

right censored and/or interval censored observations in the dataset. Needed in the context of excess hazard modelling if right censored and/or interval censored observations are present. In the latter case, d.lchrate also need be provided.

d.lchrate.td

Vector of differences of population cumulative excess hazards computed at the age of the patient when the left censoring occurred and at the age of the patient when the truncation occurred. The length of d.lchrate.td should be equal to the number of left truncated left censored and/or left truncated interval censored observations in the dataset. Needed in the context of excess hazard modelling if left truncated left censored and/or left truncated interval censored observations are present. In the latter case, d.rchrate.td also need be provided.

d.rchrate.td

Vector of differences of population cumulative excess hazards computed at the age of the patient when the right censoring occurred and at the age of the patient when the truncation occurred. The length of d.rchrate.td should be equal to the number of left truncated right censored and/or left truncated interval censored observations in the dataset. Needed in the context of excess hazard modelling if left truncated right censored and/or left truncated interval censored observations are present. In the latter case, d.lchrate.td also need be provided.

truncation.time

Variable name of truncation time.

min.dn, min.pr, max.pr

These values are used to set, depending on the model used for modelling, the minimum and maximum allowed for the densities and probabilities. These parameters are employed to avoid potential overflows/underflows in the calculations and the default values seem to offer a good compromise. Function conv.check() provides some relevant diagnostic information which can be used, for example, to check whether the lower bounds of min.dn and min.pr have been reached. So based on this or if the user wishes to do some sensitivity analysis then this can be easily carried out using these three arguments. However, the user has to be cautious. For instance, it would not make much sense to choose for min.dn and min.pr values bigger than the default ones. Bear in mind that the bounds can be reached for ill-defined models. For certain distributions/models, if convergence failure occurs and the bounds have been reached then the user can try a sensitivity analysis as mentioned above.

ygrid.tol

Tolerance used to choose grid of response values for robust discrete models. Values smaller than 1e-160 are not allowed for.

## Details

The underlying algorithm is described in ?gjrm.

There are many continuous/discrete/survival distributions to choose from and we plan to include more options. Get in touch if you are interested in a particular distribution.

The "GEVlink" option is used for binary response additive models and is more stable and faster than the R package bgeva. This model has been incorporated into this package to take advantage of the richer set of smoother choices, and of the estimation approach. Details on the model can be found in Calabrese, Marra and Osmetti (2016).

#### Value

The function returns an object of class gamlss as described in gamlssObject.

#### WARNINGS

Convergence can be checked using conv. check which provides some information about the score and information matrix associated with the fitted model. The former should be close to 0 and the latter positive definite. gamlss() will produce some warnings if there is a convergence issue.

Convergence failure may sometimes occur. This is not necessarily a bad thing as it may indicate specific problems with a fitted model. In such a situation, the user may use some extra regularisation (see extra.regI) and/or rescaling (see parscale). However, the user should especially consider re-specifying/simplifying the model, and/or checking that the chosen distribution fits the response well. In our experience, we found that convergence failure typically occurs when the model has been misspecified and/or the sample size is low compared to the complexity of the model. It is also worth bearing in mind that the use of three parameter distributions requires the data to be more informative than a situation in which two parameter distributions are used instead.

#### Author(s)

Maintainer: Giampiero Marra < giampiero . marra@ucl . ac . uk>

#### References

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Rigby R.A., Stasinopoulos D.M. (2005). Generalized additive models for location, scale and shape (with discussion). *Journal of the Royal Statistical Society, Series C*, 54(3), 507-554.

Calabrese R., Marra G., Osmetti SA (2016), Bankruptcy Prediction of Small and Medium Enterprises Using a Flexible Binary Generalized Extreme Value Model. *Journal of the Operational Research Society*, 67(4), 604-615.

Marincioni V., Marra G., Altamirano-Medina H. (2018), Development of Predictive Models for the Probabilistic Moisture Risk Assessment of Internal Wall Insulation. *Building and Environment*, 137, 5257-267.

#### See Also

```
GJRM-package, gamlssObject, conv.check, summary.gamlss
```

## **Examples**

```
## Not run:
library(GJRM)
```

```
set.seed(0)
n <- 400
x1 <- round(runif(n))</pre>
x2 <- runif(n)</pre>
x3 <- runif(n)
f1 <- function(x) cos(pi*2*x) + sin(pi*x)
y1 < -1.55 + 2*x1 + f1(x2) + rnorm(n)
dataSim <- data.frame(y1, x1, x2, x3)</pre>
resp.check(y1, "N")
eq.mu <- y1 \sim x1 + s(x2) + s(x3)
eq.s \leftarrow \sim s(x3)
fl <- list(eq.mu, eq.s)
out <- gamlss(fl, data = dataSim)</pre>
conv.check(out)
post.check(out)
plot(out, eq = 1, scale = 0, pages = 1, seWithMean = TRUE)
plot(out, eq = 2, seWithMean = TRUE)
summary(out)
AIC(out)
BIC(out)
#################
# Robust example
################
eq.mu <- y1 \sim x1 + x2 + x3
fl <- list(eq.mu)</pre>
out <- gamlss(fl, data = dataSim, margin = "N", robust = TRUE,</pre>
                   rc = 3, lB = -Inf, uB = Inf)
conv.check(out)
summary(out)
rob.const(out, 100)
##
eq.s <- ~ x3
fl <- list(eq.mu, eq.s)</pre>
out <- gamlss(fl, data = dataSim, margin = "N", robust = TRUE)</pre>
conv.check(out)
summary(out)
```

```
eq.mu <- y1 \sim x1 + s(x2) + s(x3)
eq.s \leftarrow \sim s(x3)
fl <- list(eq.mu, eq.s)</pre>
out1 <- gamlss(fl, data = dataSim, margin = "N", robust = TRUE,
               sp.method = "efs")
conv.check(out1)
summary(out1)
AIC(out, out1)
plot(out1, eq = 1, all.terms = TRUE, pages = 1, seWithMean = TRUE)
plot(out1, eq = 2, seWithMean = TRUE)
############################
## GEV link binary example
############################
# this incorporates the bgeva
# model implemented in the bgeva package
# however this implementation is more general
# stable and efficient
set.seed(0)
n <- 400
x1 <- round(runif(n)); x2 <- runif(n); x3 <- runif(n)</pre>
f1 \leftarrow function(x) cos(pi*2*x) + sin(pi*x)
f2 <- function(x) x + exp(-30*(x-0.5)^2)
y \leftarrow ifelse(-3.55 + 2*x1 + f1(x2) + rnorm(n) > 0, 1, 0)
dataSim <- data.frame(y, x1, x2, x3)</pre>
out1 <- gamlss(list(y ~ x1 + x2 + x3), margin = "GEVlink", data = dataSim)
out2 <- gamlss(list(y \sim x1 + s(x2) + s(x3)), margin = "GEVlink", data = dataSim)
conv.check(out1)
conv.check(out2)
summary(out1)
summary(out2)
AIC(out1, out2)
BIC(out1, out2)
plot(out2, eq = 1, all.terms = TRUE, pages = 1, seWithMean = TRUE)
###################
# prediction of Pr
##################
```

```
# Calculate eta (that is, X*model.coef)
# For a new data set the argument newdata should be used
eta <- predict(out2, eq = 1, type = "link")</pre>
# extract gev tail parameter
gev.par <- out2$gev.par</pre>
# multiply gev tail parameter by eta
gevpeta <- gev.par*eta
# establish for which values the model is defined
gevpetaIND <- ifelse(gevpeta < -1, FALSE, TRUE)</pre>
gevpeta <- gevpeta[gevpetaIND]</pre>
# estimate probabilities
pr \leftarrow exp(-(1 + gevpeta)^{-1/gev.par))
## Flexible survival model examples
library(GJRM)
## Simulate proportional hazards data ##
set.seed(0)
n <- 2000
c <- runif(n, 3, 8)</pre>
u <- runif(n, 0, 1)
z1 <- rbinom(n, 1, 0.5)
z2 <- runif(n, 0, 1)
t <- rep(NA, n)
beta_0 <- -0.2357
beta_1 <- 1
f <- function(t, beta_0, beta_1, u, z1, z2){</pre>
 S_0 < 0.7 * exp(-0.03*t^1.9) + 0.3*exp(-0.3*t^2.5)
 \exp(-\exp(\log(-\log(S_0))+beta_0*z1 + beta_1*z2))-u
for (i in 1:n){
  t[i] \leftarrow uniroot(f, c(0, 8), tol = .Machine$double.eps^0.5,
                 beta_0 = beta_0, beta_1 = beta_1, u = u[i],
```

```
z1 = z1[i], z2 = z2[i], extendInt = "yes" )$root
}
delta \leftarrow ifelse(t < c, 1, 0)
       <- apply(cbind(t, c), 1, min)
dataSim <- data.frame(u, delta, z1, z2)</pre>
1-mean(delta) # average censoring rate
# log(u) helps obtaining smoother hazards
out <- gamlss(list(u \sim s(log(u), bs = "mpi") + z1 + s(z2) ), data = dataSim,
              surv = TRUE, margin = "PH", cens = delta)
post.check(out)
summary(out)
AIC(out)
BIC(out)
plot(out, eq = 1, scale = 0, pages = 1)
hazsurv.plot(out, newdata = data.frame(z1 = 0, z2 = 0), shade = TRUE,
             n.sim = 1000, baseline = TRUE)
hazsurv.plot(out, type = "hazard", newdata = data.frame(z1 = 0, z2 = 0),
             shade = TRUE, n.sim = 1000, baseline = TRUE)
out1 <- gam(u \sim z1 + s(z2), family = cox.ph(),
            data = dataSim, weights = delta)
summary(out1)
\# estimates of z1 and s(z2) are
# nearly identical between out and out1
# note that the Weibull is implemented as AFT
# as using the PH parametrisation makes
# computation unstable
out2 <- gamlss(list(u \sim z1 + s(z2) ), data = dataSim, surv = TRUE,
              margin = "WEI", cens = delta)
## Simulate proportional odds data ##
set.seed(0)
n <- 2000
c <- runif(n, 4, 8)</pre>
u <- runif(n, 0, 1)
z < - rbinom(n, 1, 0.5)
beta_0 <- -1.05
t \leftarrow rep(NA, n)
f <- function(t, beta_0, u, z){</pre>
  S_0 < 0.7 * exp(-0.03*t^1.9) + 0.3*exp(-0.3*t^2.5)
  1/(1 + \exp(\log((1-S_0)/S_0) + beta_0 \times z)) - u
}
```

```
for (i in 1:n){
    t[i] \leftarrow uniroot(f, c(0, 8), tol = .Machine$double.eps^0.5,
                    beta_0 = beta_0, u = u[i], z = z[i],
                    extendInt="yes" )$root
}
delta <- ifelse(t < c,1, 0)
        <- apply(cbind(t, c), 1, min)
dataSim <- data.frame(u, delta, z)</pre>
1-mean(delta) # average censoring rate
out <- gamlss(list(u \sim s(log(u), bs = "mpi") + z), data = dataSim, surv = TRUE,
              margin = "PO", cens = delta)
post.check(out)
summary(out)
AIC(out)
BIC(out)
plot(out, eq = 1, scale = 0)
hazsurv.plot(out, newdata = data.frame(z = 0), shade = TRUE, n.sim = 1000,
             baseline = TRUE)
hazsurv.plot(out, type = "hazard", newdata = data.frame(z = 0),
             shade = TRUE, n.sim = 1000)
## Mixed censoring example ##
######################################
f1 <- function(t, u, z1, z2, z3, z4, s1, s2){
   S_0 \leftarrow 0.7 * exp(-0.03*t^1.8) + 0.3*exp(-0.3*t^2.5)
   \exp(-\exp(\log(-\log(S_0)) + 1.3*z1 + 0.5*z2 + s1(z3) + s2(z4))) - u
  }
datagen <- function(n, z1, z2, z3, z4, s1, s2, f1){
  u <- runif(n, 0, 1)
  t \leftarrow rep(NA, n)
  for (i in 1:n) t[i] \leftarrow uniroot(f1, c(0, 100), tol = .Machine$double.eps^0.5,
                                 u = u[i], s1 = s1, s2 = s2, z1 = z1[i], z2 = z2[i],
                                 z3 = z3[i], z4 = z4[i], extendInt = "yes")$root
  c1 <-
             runif(n, 0, 2)
  c2 <- c1 + runif(n, 0, 6)
  df <- data.frame(u1 = t, u2 = t, cens = character(n), stringsAsFactors = FALSE)</pre>
for (i in 1:n){
```

```
if(t[i] \le c1[i]) \{
        df[i, 1] <- c1[i]
        df[i, 2] <- NA
        df[i, 3] <- "L"
  else if(c1[i] < t[i] & t[i] <= c2[i]){
        df[i, 1] \leftarrow c1[i]
        df[i, 2] <- c2[i]
        df[i, 3] <- "I"
  else if(t[i] > c2[i]){
        df[i, 1] \leftarrow c2[i]
        df[i, 2] \leftarrow NA
        df[i, 3] \leftarrow "R"
}
uncens <- (df[, 3] \%in\% c("L", "I")) + (rbinom(n, 1, 0.2) == 1) == 2
df[uncens, 1] <- t[uncens]</pre>
df[uncens, 2] <- NA
df[uncens, 3] <- "U"
dataSim \leftarrow data.frame(u1 = df$u1, u2 = df$u2, cens = as.factor(df$cens), z1, z2, z3, z4, t)
dataSim
}
set.seed(0)
       <- 1000
SigmaC <- matrix(0.5, 4, 4); diag(SigmaC) <- 1</pre>
      <- rMVN(n, rep(0,4), SigmaC)
       <- pnorm(cov)
COV
z1
       <- round(cov[, 1])
       <- round(cov[, 2])
z2
       <- cov[, 3]
z3
       <- cov[, 4]
z4
s1
       <- function(x) -0.075*exp(3.2 * x)
       <- function(x) sin(2*pi*x)
eq1
       -u1 \sim s(\log(u1), bs = "mpi") + z1 + z2 + s(z3) + s(z4)
dataSim <- datagen(n, z1, z2, z3, z4, s1, s2, f1)
out <- gamlss(list(eq1), data = dataSim, surv = TRUE, margin = "PH",</pre>
              cens = cens, type.cen = "mixed", upperB = "u2")
conv.check(out)
summary(out)
plot(out, eq = 1, scale = 0, pages = 1)
```

gamlssObject 29

```
ndf <- data.frame(z1 = 1, z2 = 0, z3 = 0.2, z4 = 0.5)
hazsurv.plot(out, eq = 1, newdata = ndf, type = "surv")
hazsurv.plot(out, eq = 1, newdata = ndf, type = "hazard", n.sim = 1000)
## End(Not run)</pre>
```

gamlssObject

Fitted gamlssObject object

#### Description

wor.c

A fitted gamlss object returned by function gamlss and of class "gamlss" and "SemiParBIV".

#### Value

```
fit
                  List of values and diagnostics extracted from the output of the algorithm.
gam1, gam2, gam3
                  Univariate starting values' fits.
coefficients
                  The coefficients of the fitted model.
                  Prior weights used during model fitting.
weights
                  Estimated smoothing parameters of the smooth components.
sp
iter.sp
                  Number of iterations performed for the smoothing parameter estimation step.
iter.if
                  Number of iterations performed in the initial step of the algorithm.
iter.inner
                  Number of iterations performed within the smoothing parameter estimation step.
                  Sample size.
X1, X2, X3, ...
                  Design matrices associated with the linear predictors.
X1.d2, X2.d2, X3.d2, ...
                  Number of columns of X1, X2, X3, etc.
1.sp1, 1.sp2, 1.sp3, ...
                  Number of smooth components in the equations.
He
                  Penalized -hessian/Fisher. This is the same as HeSh for unpenalized models.
                   Unpenalized -hessian/Fisher.
HeSh
٧b
                  Inverse of He. This corresponds to the Bayesian variance-covariance matrix used
                  for confidence/credible interval calculations.
F
                  This is obtained multiplying Vb by HeSh.
                  Total degrees of freedom of the estimated bivariate model. It is calculated as
t.edf
                  sum(diag(F)).
edf1, edf2, edf3, ...
                  Degrees of freedom for the model's equations.
```

Working model quantities.

30 ggmtrust

```
eta1, eta2, eta3, ...
```

Estimated linear predictors.

y1 Response.

logLik Value of the (unpenalized) log-likelihood evaluated at the (penalized or unpe-

nalized) parameter estimates.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

## See Also

```
gamlss, summary.gamlss
```

ggmtrust

ggmtrust for penalised network

## Description

penalised network, work in progress.

## Usage

## **Arguments**

s Sample covariance matrix.

n Sample size.

data Data.

lambda Regularisation parameter. pen Either "lasso" or "ridge".

params If different from null then these are taken as the starting values.

method Either "H" or "BHHH".

w. alasso weight for alasso.
gamma alasso param.
a scad param.

#### **Details**

penalised network, work in progress.

#### Value

The function returns an object of class ggmtrust.

gjrm Generalised Joint Regression Models with Binary/Continuous/Discrete/Survival Margins

#### **Description**

gjrm fits flexible joint models with binary/continuous/discrete/survival margins, with several types of covariate effects, copula and marginal distributions.

### Usage

```
gjrm(formula, data = list(), weights = NULL, subset = NULL,
    BivD = "N", BivD2 = "N", margins, Model, dof = 3, dof2 = 3, ordinal = FALSE,
    surv = FALSE, cens1 = NULL, cens2 = NULL, cens3 = NULL, dep.cens = FALSE,
    upperBt1 = NULL, upperBt2 = NULL,
    gamlssfit = FALSE, fp = FALSE, infl.fac = 1,
    rinit = 1, rmax = 100,
    iterlimsp = 50, tolsp = 1e-07,
    gc.l = FALSE, parscale, extra.regI = "t",
    k1.tvc = 0, k2.tvc = 0, knots = NULL,
    penCor = "unpen", sp.penCor = 3,
    Chol = FALSE, gamma = 1, w.alasso = NULL,
    drop.unused.levels = TRUE,
    min.dn = 1e-40, min.pr = 1e-16, max.pr = 0.999999)
```

#### **Arguments**

formula

In the basic setup this will be a list of two (or three) formulas, one for equation 1, the other for equation 2 and another one for equation 3 if a trivariate model is fitted to the data. Otherwise, more equations can be used depending on the number of distributional parameters. s terms are used to specify smooth functions of predictors; see the documentation of mgcv for further details on formula specifications. Note that if a selection model is employed (that is, Model = "BSS" or Model = "TSS") then the first formula (and the second as well for trivariate models) MUST refer to the selection equation(s). When one outcome is binary and the other continuous/discrete then the first equation should refer to the binary outcome whereas the second to the continuous/discrete one. When one outcome is discrete and the other continuous then the first equation has to be the discrete one.

data

An optional data frame, list or environment containing the variables in the model. If not found in data, the variables are taken from environment(formula), typically the environment from which girm is called.

weights

Optional vector of prior weights to be used in fitting.

subset

Optional vector specifying a subset of observations to be used in the fitting process.

BivD

Type of bivariate error distribution employed. Possible choices are "N", "C0", "C90", "C180", "C270", "GAL0", "GAL90", "GAL180", "GAL270", "J0", "J90", "J180", "J270", "G0", "G90", "G180", "G270", "F", "AMH", "FGM", "T", "PL", "HO" which stand for bivariate normal, Clayton, rotated Clayton (90 degrees), survival Clayton, rotated Clayton (270 degrees), Galambos, rotated Galambos (90 degrees), survival Galambos, rotated Galambos (270 degrees), Joe, rotated Joe (90 degrees), survival Joe, rotated Joe (270 degrees), Gumbel, rotated Gumbel (90 degrees), survival Gumbel, rotated Gumbel (270 degrees), Frank, Ali-Mikhail-Haq, Farlie-Gumbel-Morgenstern, Student-t with dof, Plackett, Hougaard. Each of the Clayton, Galambos, Joe and Gumbel copulae is allowed to be mixed with a rotated version of the same family. The options are: "C0C90", "C0C270", "C180C90", "C180C270", "GAL0GAL90", "GAL0GAL270", "GAL180GAL270", "GAL180GAL270", "G180G90", "G180G270", "J0J90", "J0J270", "J180J90" and "J180J270". This allows the user to model negative and positive tail dependencies.

BivD2

As above but used only for Roy models.

margins

It indicates the distributions used for the two or three margins. Possible distributions are normal ("N"), log-normal ("LN"), Gumbel ("GU"), reverse Gumbel ("rGU"), generelised Pareto ("GP"), generelised Pareto II ("GPII") where the shape parameter is forced to be > -0.5, generelised Pareto (with orthogonal parametrisation) ("GPo") where the shape parameter is forced to be > -0.5, discrete generelised Pareto ("DGP"), discrete generelised Pareto II ("DG-PII") where the shape parameter is forced to be positive, discrete generelised Pareto derived under the scenario in which shape = 0 ("DGPO"), logistic ("LO"), Weibull ("WEI"), inverse Gaussian ("iG"), gamma ("GA"), Dagum ("DAGUM"), Singh-Maddala ("SM"), beta ("BE"), Fisk ("FISK", also known as log-logistic distribution), Poisson ("PO"), zero truncated Poisson ("ZTP"), negative binomial - type I ("NBI"), negative binomial - type II ("NBI"), Poisson inverse Gaussian ("PIG"). If the responses are binary then possible link functions are "probit", "logit", "cloglog". For survival models, the margins can be proportional hazars ("PH"), odds ("PO") or "probit".

Model

Possible values are "B" (bivariate model), "T" (trivariate model) "BSS" (bivariate model with non-random sample selection), "TSS" (trivariate model with double non-random sample selection), "TESS" (trivariate model with endogeneity and non-random sample selection), "BPO" (bivariate model with partial observability) and "BPO0" (bivariate model with partial observability) and zero correlation). Options "T", "TESS" and "TSS" are currently for trivariate binary models only. "BPO" and "BPO0" are for bivariate binary models only. "ROY" is for the Roy switching regression model.

dof

If BivD = "T" then the degrees of freedom can be set to a value greater than 2 and smaller than 249. Only for continuous margins, this will be taken as a starting

	value and the dof estimated from the data.
dof2	As above but used only for Roy models.
ordinal	If TRUE then the ordinal model is employed.
surv	If TRUE then a bivariate survival model is fitted.
cens1	Censoring indicator for the first equation. This is required when surv = TRUE. For the case of right censored data only, this variable can be equal to 1 if the event occurred and 0 otherwise. However, if there are several censoring mechanisms then cens will have to be specified as a factor variable made up of four possible categories: I for interval, L for left, R for right, and U for uncensored.
cens2	Same as above but for the second equation.
cens3	Binary censoring indicator employed only when surv = TRUE, dep.cens = TRUE and administrative censoring is present.
dep.cens	If TRUE then the dependence censored model is employed.
upperBt1, upperBt2	
	Variable names of right/upper bounds when interval censoring is present.
gamlssfit	If gamlssfit = TRUE then gamlss univariate models are also fitted. This is useful for obtaining starting values, for instance.
fp	If TRUE then a fully parametric model with unpenalised regression splines if fitted. See the Example 2 below.
infl.fac	Inflation factor for the model degrees of freedom in the approximate AIC. Smoother models can be obtained setting this parameter to a value greater than 1.
rinit	Starting trust region radius. The trust region radius is adjusted as the algorithm proceeds. See the documentation of trust for further details.
rmax	Maximum allowed trust region radius. This may be set very large. If set small, the algorithm traces a steepest descent path.
iterlimsp	A positive integer specifying the maximum number of loops to be performed before the smoothing parameter estimation step is terminated.
tolsp	Tolerance to use in judging convergence of the algorithm when automatic smoothing parameter estimation is used.
gc.l	This is relevant when working with big datasets. If TRUE then the garbage collector is called more often than it is usually done. This keeps the memory footprint down but it will slow down the routine.
parscale	The algorithm will operate as if optimizing objfun( $x$ / parscale,) where parscale is a scalar. If missing then no rescaling is done. See the documentation of trust for more details.
extra.regI	If "t" then regularization as from trust is applied to the information matrix if needed. If different from "t" then extra regularization is applied via the options "pC" (pivoted Choleski - this will only work when the information matrix is semi-positive or positive definite) and "sED" (symmetric eigen-decomposition).
k1.tvc, k2.tvc	Experimental. Only used for tvc ps smoothers when using survival models.

Optional list containing user specified knot values to be used for basis construc-

knots

tion.

penCor This and the arguments below are only for trivariate binary models. Type of

penalty for correlation coefficients. Possible values are "unpen", "lasso", "ridge",

"alasso".

sp.penCor Starting value for smoothing parameter of penCor.

Chol If TRUE then the Cholesky method instead of the eigenvalue method is employed

for the correlation matrix.

gamma Inflation factor used only for the alasso penalty.

w. alasso When using the alasso penalty a weight vector made up of three values must be

provided.

drop.unused.levels

By default unused levels are dropped from factors before fitting. For some smooths involving factor variables this may have to be turned off (only use if

you know what you are doing).

min.dn, min.pr, max.pr

These values are used to set, depending on the model used for modelling, the minimum and maximum allowed for the densities and probabilities; recall that the margins of copula models have to be in the range (0,1). These parameters are employed to avoid potential overflows/underflows in the calculations and the default values seem to offer a good compromise. Function conv.check() provides some relevant diagnostic information which can be used, for example, to check whether the lower bounds of min.dn and min.pr have been reached. So based on this or if the user wishes to do some sensitivity analysis then this can be easily carried out using these three arguments. However, the user has to be cautious. For instance, it would not make much sense to choose for min.dn and min.pr values bigger than the default ones. Bear in mind that the bounds can be reached for ill-defined models. For certain distributions/models, if convergence failure occurs and the bounds have been reached then the user can try a sensitivity analysis as mentioned above.

#### Details

The joint models considered by this function consist of two or three model equations which depend on flexible linear predictors and whose dependence between the responses is modelled through one or more parameters of a chosen multivariate distribution. The additive predictors of the equations are flexibly specified using parametric components and smooth functions of covariates. The same can be done for the dependence parameter(s) if it makes sense. Estimation is achieved within a penalized likelihood framework with integrated automatic multiple smoothing parameter selection. The use of penalty matrices allows for the suppression of that part of smooth term complexity which has no support from the data. The trade-off between smoothness and fitness is controlled by smoothing parameters associated with the penalty matrices. Smoothing parameters are chosen to minimise an approximate AIC.

For sample selection models, if there are factors in the model then before fitting the user has to ensure that the numbers of factor variables' levels in the selected sample are the same as those in the complete dataset. Even if a model could be fitted in such a situation, the model may produce fits which are not coherent with the nature of the correction sought. As an example consider the situation in which the complete dataset contains a factor variable with five levels and that only three of them appear in the selected sample. For the outcome equation (which is the one of interest)

only three levels of such variable exist in the population, but their effects will be corrected for nonrandom selection using a selection equation in which five levels exist instead. Having differing numbers of factors' levels between complete and selected samples will also make prediction not feasible (an aspect which may be particularly important for selection models); clearly it is not possible to predict the response of interest for the missing entries using a dataset that contains all levels of a factor variable but using an outcome model estimated using a subset of these levels.

There are many continuous/discrete/survival distributions and copula functions to choose from and we plan to include more options. Get in touch if you are interested in a particular distribution.

#### Value

The function returns an object of class gjrm as described in gjrmObject.

#### WARNINGS

Convergence can be checked using conv. check which provides some information about the score and information matrix associated with the fitted model. The former should be close to 0 and the latter positive definite. gjrm() will produce some warnings if there is a convergence issue.

Convergence failure may sometimes occur. This is not necessarily a bad thing as it may indicate specific problems with a fitted model. In such a situation, the user may use some extra regularisation (see extra.regI) and/or rescaling (see parscale). Using gamlssfit = TRUE is typically more effective than the first two options as this will provide better calibrated starting values as compared to those obtained from the default starting value procedure. The default option is, however, gamlssfit = FALSE only because it tends to be computationally cheaper and because the default procedure has typically been found to do a satisfactory job in most cases. (The results obtained when using gamlssfit = FALSE and gamlssfit = TRUE could also be compared to check if starting values make any difference.)

The above suggestions may help, especially the latter option. However, the user should also consider re-specifying/simplifying the model, and/or using a diferrent dependence structure and/or checking that the chosen marginal distributions fit the responses well. In our experience, we found that convergence failure typically occurs when the model has been misspecified and/or the sample size is low compared to the complexity of the model. Examples of misspecification include using a Clayton copula rotated by 90 degrees when a positive association between the margins is present instead, using marginal distributions that do not fit the responses, and employing a copula which does not accommodate the type and/or strength of the dependence between the margins (e.g., using AMH when the association between the margins is strong). When using smooth functions, if the covariate's values are too sparse then convergence may be affected by this. It is also worth bearing in mind that the use of three parameter marginal distributions requires the data to be more informative than a situation in which two parameter distributions are used instead.

In the contexts of endogeneity and non-random sample selection, extra attention is required when specifying the dependence parameter as a function of covariates. This is because in these situations the dependence parameter mainly models the association between the unobserved confounders in the two equations. Therefore, this option would make sense when it is believed that the strength of the association between the unobservables in the two equations varies based on some grouping factor or across geographical areas, for instance. In any case, a clear rationale is typically needed in such cases.

#### Author(s)

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#### References

See help("GJRM-package").

#### See Also

adjCov, VuongClarke, GJRM-package, gjrmObject, conv.check, summary.gjrm

## **Examples**

```
library(GJRM)
# JOINT MODELS WITH BINARY MARGINS #
############
## Example 1
############
set.seed(0)
n <- 400
Sigma \leftarrow matrix(0.5, 2, 2); diag(Sigma) \leftarrow 1
  <- rMVN(n, rep(0,2), Sigma)
x1 <- round(runif(n)); x2 <- runif(n); x3 <- runif(n)</pre>
   <- function(x) cos(pi*2*x) + sin(pi*x)
f2 <- function(x) x+exp(-30*(x-0.5)^2)
y1 \leftarrow ifelse(-1.55 + 2*x1 + f1(x2) + u[,1] > 0, 1, 0)
y2 \leftarrow ifelse(-0.25 - 1.25*x1 + f2(x2) + u[,2] > 0, 1, 0)
dataSim <- data.frame(y1, y2, x1, x2, x3)</pre>
## CLASSIC BIVARIATE PROBIT
out \leftarrow gjrm(list(y1 ~ x1 + x2 + x3,
                  y2 \sim x1 + x2 + x3),
                  data = dataSim,
                  margins = c("probit", "probit"),
                  Model = "B")
```

```
conv.check(out)
summary(out)
AIC(out)
BIC(out)
## Not run:
## BIVARIATE PROBIT with Splines
out <- gjrm(list(y1 \sim x1 + s(x2) + s(x3),
                  y2 \sim x1 + s(x2) + s(x3)),
                   data = dataSim,
                   margins = c("probit", "probit"),
                   Model = "B")
conv.check(out)
summary(out)
AIC(out)
## estimated smooth function plots - red lines are true curves
x2 <- sort(x2)
f1.x2 \leftarrow f1(x2)[order(x2)] - mean(f1(x2))
f2.x2 \leftarrow f2(x2)[order(x2)] - mean(f2(x2))
f3.x3 \leftarrow rep(0, length(x3))
par(mfrow=c(2,2), mar=c(4.5,4.5,2,2))
plot(out, eq = 1, select = 1, seWithMean = TRUE, scale = 0)
lines(x2, f1.x2, col = "red")
plot(out, eq = 1, select = 2, seWithMean = TRUE, scale = 0)
lines(x3, f3.x3, col = "red")
plot(out, eq = 2, select = 1, seWithMean = TRUE, scale = 0)
lines(x2, f2.x2, col = "red")
plot(out, eq = 2, select = 2, seWithMean = TRUE, scale = 0)
lines(x3, f3.x3, col = "red")
## BIVARIATE PROBIT with Splines and
## varying dependence parameter
eq.mu.1 <- y1 \sim x1 + s(x2)
eq.mu.2 <- y2 \sim x1 + s(x2)
eq.theta \leftarrow \sim x1 + s(x2)
fl <- list(eq.mu.1, eq.mu.2, eq.theta)</pre>
outD <- gjrm(fl, data = dataSim,</pre>
             margins = c("probit", "probit"),
             Model = "B")
conv.check(outD)
summary(outD)
```

```
outD$theta
plot(outD, eq = 1, seWithMean = TRUE)
plot(outD, eq = 2, seWithMean = TRUE)
plot(outD, eq = 3, seWithMean = TRUE)
graphics.off()
############
## Example 2
###########
## Generate data with one endogenous variable
## and exclusion restriction
set.seed(0)
n <- 400
Sigma \leftarrow matrix(0.5, 2, 2); diag(Sigma) \leftarrow 1
   <- rMVN(n, rep(0,2), Sigma)
     <- rMVN(n, rep(0,2), Sigma)
cov <- pnorm(cov)</pre>
x1 \leftarrow round(cov[,1]); x2 \leftarrow cov[,2]
f1 <- function(x) cos(pi*2*x) + sin(pi*x)
f2 <- function(x) x+exp(-30*(x-0.5)^2)
y1 \leftarrow ifelse(-1.55 + 2*x1 + f1(x2) + u[,1] > 0, 1, 0)
y2 \leftarrow ifelse(-0.25 - 1.25*y1 + f2(x2) + u[,2] > 0, 1, 0)
dataSim <- data.frame(y1, y2, x1, x2)</pre>
## Testing the hypothesis of absence of endogeneity...
LM.bpm(list(y1 \sim x1 + s(x2), y2 \sim y1 + s(x2)), dataSim, Model = "B")
## CLASSIC RECURSIVE BIVARIATE PROBIT
out <- gjrm(list(y1 \sim x1 + x2,
                  y2 \sim y1 + x2),
                  data = dataSim,
                  margins = c("probit", "probit"),
                  Model = "B")
conv.check(out)
summary(out)
AIC(out); BIC(out)
## FLEXIBLE RECURSIVE BIVARIATE PROBIT
out <- gjrm(list(y1 \sim x1 + s(x2),
```

```
y2 \sim y1 + s(x2)),
                 data = dataSim,
                 margins = c("probit", "probit"),
                 Model = "B")
conv.check(out)
summary(out)
AIC(out); BIC(out)
## Testing the hypothesis of absence of endogeneity post estimation...
gt.bpm(out)
## treatment effect, risk ratio and odds ratio with CIs
mb(y1, y2, Model = "B")
AT(out, nm.end = "y1", hd.plot = TRUE)
RR(out, nm.end = "y1")
OR(out, nm.end = "y1")
AT(out, nm.end = "y1", type = "univariate")
re.imp <- imputeCounter(out, m = 10, "y1")</pre>
re.imp$AT
## try a Clayton copula model...
outC <- gjrm(list(y1 \sim x1 + s(x2),
                  y2 \sim y1 + s(x2)),
                   data = dataSim, BivD = "C0",
                  margins = c("probit", "probit"),
                  Model = "B")
conv.check(outC)
summary(outC)
AT(outC, nm.end = "y1")
re.imp <- imputeCounter(outC, m = 10, "y1")</pre>
re.imp$AT
## try a Joe copula model...
outJ <- gjrm(list(y1 \sim x1 + s(x2),
                  y2 \sim y1 + s(x2)),
                   data = dataSim, BivD = "J0",
                  margins = c("probit", "probit"),
                  Model = "B")
conv.check(outJ)
summary(outJ)
AT(outJ, "y1")
re.imp <- imputeCounter(outJ, m = 10, "y1")</pre>
re.imp$AT
VuongClarke(out, outJ)
#
```

```
## recursive bivariate probit modelling with unpenalized splines
## can be achieved as follows
outFP <- gjrm(list(y1 \sim x1 + s(x2, bs = "cr", k = 5),
                   y2 \sim y1 + s(x2, bs = "cr", k = 6)),
                   fp = TRUE, data = dataSim,
                   margins = c("probit", "probit"),
                   Model = "B")
conv.check(outFP)
summary(outFP)
# in the above examples a third equation could be introduced
# as illustrated in Example 1
##################
## See also ?meps
###################
###########
## Example 3
## Generate data with a non-random sample selection mechanism
## and exclusion restriction
set.seed(0)
n <- 2000
Sigma \leftarrow matrix(0.5, 2, 2); diag(Sigma) \leftarrow 1
   <- rMVN(n, rep(0,2), Sigma)
SigmaC <- matrix(0.5, 3, 3); diag(SigmaC) <- 1</pre>
       <- rMVN(n, rep(0,3), SigmaC)
cov
       <- pnorm(cov)
bi <- round(cov[,1]); x1 <- cov[,2]; x2 <- cov[,3]
f11 <- function(x) -0.7*(4*x + 2.5*x^2 + 0.7*sin(5*x) + cos(7.5*x))
f12 <- function(x) -0.4*(-0.3 - 1.6*x + \sin(5*x))
f21 \leftarrow function(x) \ 0.6*(exp(x) + sin(2.9*x))
ys <- 0.58 + 2.5*bi + f11(x1) + f12(x2) + u[, 1] > 0
y < -0.68 - 1.5*bi + f21(x1) + u[, 2] > 0
yo <- y*(ys > 0)
dataSim <- data.frame(y, ys, yo, bi, x1, x2)</pre>
## Testing the hypothesis of absence of non-random sample selection...
LM.bpm(list(ys \sim bi + s(x1) + s(x2), yo \sim bi + s(x1)), dataSim, Model = "BSS")
# p-value suggests presence of sample selection, hence fit a bivariate model
```

```
#
## SEMIPARAMETRIC SAMPLE SELECTION BIVARIATE PROBIT
## the first equation MUST be the selection equation
out <- gjrm(list(ys \sim bi + s(x1) + s(x2),
                 yo \sim bi + s(x1)),
                 data = dataSim, Model = "BSS",
                 margins = c("probit", "probit"))
conv.check(out)
gt.bpm(out)
## compare the two summary outputs
## the second output produces a summary of the results obtained when
## selection bias is not accounted for
summary(out)
summary(out$gam2)
## corrected predicted probability that 'yo' is equal to 1
mb(ys, yo, Model = "BSS")
prev(out, hd.plot = TRUE)
prev(out, type = "univariate", hd.plot = TRUE)
## estimated smooth function plots
## the red line is the true curve
## the blue line is the univariate model curve not accounting for selection bias
x1.s <- sort(x1[dataSim$ys>0])
f21.x1 \leftarrow f21(x1.s)[order(x1.s)]-mean(f21(x1.s))
plot(out, eq = 2, ylim = c(-1.65, 0.95)); lines(x1.s, f21.x1, col="red")
par(new = TRUE)
plot(out\$gam2, se = FALSE, col = "blue", ylim = c(-1.65,0.95),
     ylab = "", rug = FALSE)
#
## try a Clayton copula model...
outC <- gjrm(list(ys \sim bi + s(x1) + s(x2),
                  yo \sim bi + s(x1)),
                  data = dataSim, Model = "BSS", BivD = "C0",
                  margins = c("probit", "probit"))
conv.check(outC)
summary(outC)
prev(outC)
##############################
# Impute using Mice
##############################
```

```
library(mice)
ys <- dataSim$ys
dataSim$yo[dataSim$ys == FALSE] <- NA
dataSim <- dataSim[, -c(1:2)]</pre>
imp <- mice(dataSim, m = 1, method = c("copulaSS", "", "", ""),</pre>
            mice.formula = outC$mice.formula, margins = outC$margins,
            BivD = outC$BivD, maxit = 1)
comp.yo <- dataSim$yo</pre>
comp.yo[ys == 0] <- impsyo[[1]]
mean(comp.yo)
#
#################
## See also ?hiv
#################
###########
## Example 4
###########
## Generate data with partial observability
set.seed(0)
n <- 10000
Sigma \leftarrow matrix(0.5, 2, 2); diag(Sigma) \leftarrow 1
     <- rMVN(n, rep(0,2), Sigma)
x1 <- round(runif(n)); x2 <- runif(n); x3 <- runif(n)</pre>
y1 \leftarrow ifelse(-1.55 + 2*x1 + x2 + u[,1] > 0, 1, 0)
y2 \leftarrow ifelse(0.45 - x3 + u[,2] > 0, 1, 0)
y <- y1*y2
dataSim <- data.frame(y, x1, x2, x3)</pre>
## BIVARIATE PROBIT with Partial Observability
out <- gjrm(list(y \sim x1 + x2,
                  y \sim x3),
                   data = dataSim, Model = "BPO",
                   margins = c("probit", "probit"))
conv.check(out)
summary(out)
# first ten estimated probabilities for the four events from object out
```

```
cbind(out$p11, out$p10, out$p00, out$p01)[1:10,]
# case with smooth function
# (more computationally intensive)
f1 <- function(x) cos(pi*2*x) + sin(pi*x)
y1 \leftarrow ifelse(-1.55 + 2*x1 + f1(x2) + u[,1] > 0, 1, 0)
y2 <- ifelse( 0.45 - x3
                     + u[,2] > 0, 1, 0)
y <- y1*y2
dataSim <- data.frame(y, x1, x2, x3)</pre>
out <- gjrm(list(y \sim x1 + s(x2),
             y \sim x3),
             data = dataSim, Model = "BPO",
             margins = c("probit", "probit"))
conv.check(out)
summary(out)
# plot estimated and true functions
x2 \leftarrow sort(x2); f1.x2 \leftarrow f1(x2)[order(x2)] - mean(f1(x2))
plot(out, eq = 1, scale = 0); lines(x2, f1.x2, col = "red")
##################
## See also ?war
##################
## End(Not run)
## Not run:
# JOINT MODELS WITH BINARY AND/OR CONTINUOUS MARGINS #
library(GJRM)
############
## Example 5
## Generate data
## Correlation between the two equations 0.5 - Sample size 400
set.seed(0)
```

```
n <- 400
Sigma <- matrix(0.5, 2, 2); diag(Sigma) <- 1
     <- rMVN(n, rep(0,2), Sigma)
x1 <- round(runif(n)); x2 <- runif(n); x3 <- runif(n)</pre>
f1 <- function(x) cos(pi*2*x) + sin(pi*x)
f2 <- function(x) x+exp(-30*(x-0.5)^2)
y1 < -1.55 + 2*x1 + f1(x2) + u[,1]
y2 < -0.25 - 1.25*x1 + f2(x2) + u[,2]
dataSim <- data.frame(y1, y2, x1, x2, x3)</pre>
resp.check(y1, "N")
resp.check(y2, "N")
eq.mu.1
           <-y1 \sim x1 + s(x2) + s(x3)
eq.mu.2
           <- y2 \sim x1 + s(x2) + s(x3)
eq.sigma1 <- ~ 1
eq.sigma2 <- ~ 1
          <- ~ x1
eq.theta
f1 <- list(eq.mu.1, eq.mu.2, eq.sigma1, eq.sigma2, eq.theta)</pre>
# the order above is the one to follow when
# using more than two equations
out <- gjrm(fl, data = dataSim, margins = c("N", "N"),
             Model = "B")
conv.check(out)
post.check(out)
summary(out)
AIC(out)
BIC(out)
jc.probs(out, 1.4, 2.3, intervals = TRUE)[1:4,]
###########
## Example 6
###########
## Generate data with one endogenous binary variable
## and continuous outcome
set.seed(0)
n <- 1000
Sigma \leftarrow matrix(0.5, 2, 2); diag(Sigma) \leftarrow 1
u <- rMVN(n, rep(0,2), Sigma)
```

```
<- rMVN(n, rep(0,2), Sigma)
cov
     <- pnorm(cov)
COV
x1 <- round(cov[,1]); x2 <- cov[,2]</pre>
f1 <- function(x) cos(pi*2*x) + sin(pi*x)
f2 <- function(x) x+exp(-30*(x-0.5)^2)
y1 \leftarrow ifelse(-1.55 + 2*x1 + f1(x2) + u[,1] > 0, 1, 0)
             -0.25 - 1.25*y1 + f2(x2) + u[,2]
y2 <-
dataSim <- data.frame(y1, y2, x1, x2)</pre>
## RECURSIVE Model
rc <- resp.check(y2, margin = "N", print.par = TRUE, loglik = TRUE)</pre>
AIC(rc); BIC(rc)
out <- gjrm(list(y1 \sim x1 + x2,
                 y2 \sim y1 + x2),
                 data = dataSim, margins = c("probit","N"),
                 Model = "B")
conv.check(out)
summary(out)
post.check(out)
## SEMIPARAMETRIC RECURSIVE Model
eq.mu.1 <-y1 \sim x1 + s(x2)
eq.mu.2 <- y2 \sim y1 + s(x2)
eq.sigma <- ~ 1
eq.theta <- ~ 1
fl <- list(eq.mu.1, eq.mu.2, eq.sigma, eq.theta)</pre>
out <- gjrm(fl, data = dataSim,</pre>
            margins = c("probit","N"), gamlssfit = TRUE,
            Model = "B")
conv.check(out)
summary(out)
post.check(out)
jc.probs(out, 1, 1.5, intervals = TRUE)[1:4,]
AT(out, nm.end = "y1")
AT(out, nm.end = "y1", type = "univariate")
#
############
## Example 7
###########
## Generate data with one endogenous continuous exposure
## and binary outcome
```

```
set.seed(0)
n <- 1000
Sigma <- matrix(0.5, 2, 2); diag(Sigma) <- 1
     <- rMVN(n, rep(0,2), Sigma)
cov <- rMVN(n, rep(0,2), Sigma)</pre>
cov <- pnorm(cov)</pre>
x1 <- round(cov[,1]); x2 <- cov[,2]</pre>
f1 <- function(x) cos(pi*2*x) + sin(pi*x)
f2 <- function(x) x+exp(-30*(x-0.5)^2)
y1 <-
             -0.25 - 2*x1 + f2(x2) + u[,2]
y2 \leftarrow ifelse(-0.25 - 0.25*y1 + f1(x2) + u[,1] > 0, 1, 0)
dataSim <- data.frame(y1, y2, x1, x2)</pre>
eq.mu.1 <- y2 \sim y1 + s(x2)
eq.mu.2 <-y1 \sim x1 + s(x2)
eq.sigma <- ~ 1
eq.theta <- ~ 1
fl <- list(eq.mu.1, eq.mu.2, eq.sigma, eq.theta)</pre>
out <- gjrm(fl, data = dataSim,</pre>
            margins = c("probit","N"),
            Model = "B")
conv.check(out)
summary(out)
post.check(out)
AT(out, nm.end = "y1")
AT(out, nm.end = "y1", type = "univariate")
RR(out, nm.end = "y1", rr.plot = TRUE)
RR(out, nm.end = "y1", type = "univariate")
OR(out, nm.end = "y1", or.plot = TRUE)
OR(out, nm.end = "y1", type = "univariate")
#
###########
## Example 8
###################
## Survival models
####################
set.seed(0)
n <- 2000
c <- runif(n, 3, 8)</pre>
```

```
u <- runif(n, 0, 1)
z1 <- rbinom(n, 1, 0.5)
z2 <- runif(n, 0, 1)
t < -rep(NA, n)
beta_0 <- -0.2357
beta_1 <- 1
f <- function(t, beta_0, beta_1, u, z1, z2){</pre>
  S_0 \leftarrow 0.7 * exp(-0.03*t^1.9) + 0.3*exp(-0.3*t^2.5)
  \exp(-\exp(\log(-\log(S_0))+beta_0*z1 + beta_1*z2))-u
}
for (i in 1:n){
   t[i] \leftarrow uniroot(f, c(0, 8), tol = .Machine$double.eps^0.5,
                    beta_0 = beta_0, beta_1 = beta_1, u = u[i],
                    z1 = z1[i], z2 = z2[i], extendInt = "yes" )$root
}
delta1 <- ifelse(t < c, 1, \theta)
        <- apply(cbind(t, c), 1, min)
dataSim <- data.frame(u1, delta1, z1, z2)</pre>
c <- runif(n, 4, 8)</pre>
u <- runif(n, 0, 1)
z < - rbinom(n, 1, 0.5)
beta_0 <- -1.05
      <- rep(NA, n)
f <- function(t, beta_0, u, z){</pre>
  S_0 \leftarrow 0.7 * exp(-0.03*t^1.9) + 0.3*exp(-0.3*t^2.5)
  1/(1 + \exp(\log((1-S_0)/S_0) + beta_0*z)) - u
}
for (i in 1:n){
    t[i] \leftarrow uniroot(f, c(0, 8), tol = .Machine$double.eps^0.5,
                     beta_0 = beta_0, u = u[i], z = z[i],
                     extendInt="yes" )$root
}
delta2 \leftarrow ifelse(t < c,1, 0)
       <- apply(cbind(t, c), 1, min)
dataSim$delta2 <- delta2</pre>
dataSim$u2
               <- u2
dataSim$z
                <- z
eq1 <- u1 \sim s(\log(u1), bs = "mpi") + z1 + s(z2)
```

```
eq2 <- u2 \sim s(log(u2), bs = "mpi") + z
eq3 <- \sim s(z2)
out <- gjrm(list(eq1, eq2), data = dataSim, surv = TRUE,
            margins = c("PH", "PO"),
            cens1 = delta1, cens2 = delta2, Model = "B")
# PH margin fit can also be compared with cox.ph from mgcv
conv.check(out)
res <- post.check(out)</pre>
## martingale residuals
mr1 <- out$cens1 - res$qr1</pre>
mr2 <- out$cens2 - res$qr2</pre>
# can be plotted against covariates
# obs index, survival time, rank order of
# surv times
# to determine func form, one may use
# res from null model against covariate
# to test for PH, use:
# library(survival)
# fit <- coxph(Surv(u1, delta1) ~ z1 + z2, data = dataSim)</pre>
# temp <- cox.zph(fit)</pre>
# print(temp)
# plot(temp, resid = FALSE)
summary(out)
AIC(out); BIC(out)
plot(out, eq = 1, scale = 0, pages = 1)
plot(out, eq = 2, scale = 0, pages = 1)
hazsurv.plot(out, eq = 1, newdata = data.frame(z1 = 0, z2 = 0),
             shade = TRUE, n.sim = 100, baseline = TRUE)
hazsurv.plot(out, eq = 1, newdata = data.frame(z1 = 0, z2 = 0),
             shade = TRUE, n.sim = 100, type = "hazard", baseline = TRUE,
             intervals = FALSE)
hazsurv.plot(out, eq = 2, newdata = data.frame(z = 0),
             shade = FALSE, n.sim = 100, baseline = TRUE)
hazsurv.plot(out, eq = 2, newdata = data.frame(z = 0),
             shade = TRUE, n.sim = 100, type = "hazard", baseline = TRUE)
jc.probs(out, type = "joint", intervals = TRUE)[1:5,]
newd0 <- newd1 <- data.frame(z = 0, z1 = mean(dataSim$z1),
                             z2 = mean(dataSim$z2),
                              u1 = mean(dataSim$u1) + 1,
                              u2 = mean(dataSim$u2) + 1)
newd1$z <- 1
```

```
jc.probs(out, type = "joint", newdata = newd0, intervals = TRUE)
jc.probs(out, type = "joint", newdata = newd1, intervals = TRUE)
out1 <- gjrm(list(eq1, eq2, eq3), data = dataSim, surv = TRUE,
                 margins = c("PH", "PO"),
                 cens1 = delta1, cens2 = delta2, gamlssfit = TRUE,
                 Model = "B")
\# eq1 <- u1 \sim z1 + s(z2)
# eq2 <- u2 ~ z
\# eq3 < - \sim s(z2)
# note that Weibull is implemented as AFT model (test case)
\# out2 <- gjrm(list(eq1, eq2, ~ 1, ~ 1, eq3), data = dataSim, surv = TRUE,
                   margins = c("WEI", "WEI"),
#
                   cens1 = delta1, cens2 = delta2,
#
                   Model = "B")
## Joint continuous and survival outcomes
# work in progress
# eq1 <- z2 ~ z1
\# eq2 <- u2 \sim s(u2, bs = "mpi") + z
# eq3 <- ~ s(z2)
# eq4 <- ~ s(z2)
# f.1 <- list(eq1, eq2, eq3, eq4)
# out3 <- gjrm(f.1, data = dataSim, surv = TRUE,</pre>
                   margins = c("N", "PO"),
                   cens1 = NULL, cens2 = delta2,
                   gamlssfit = TRUE, Model = "B")
# conv.check(out3)
# post.check(out3)
# summary(out3)
# AIC(out3); BIC(out3)
\# plot(out3, eq = 2, scale = 0, pages = 1)
# plot(out3, eq = 3, scale = 0, pages = 1)
# plot(out3, eq = 4, scale = 0, pages = 1)
# newd <- newd1 <- data.frame(z = 0, z1 = mean(dataSim$z1),
                              z2 = mean(dataSim$z2),
#
                             u2 = mean(dataSim$u2) + 1)
# jc.probs(out3, y1 = 0.6, type = "joint", newdata = newd, intervals = TRUE)
## End(Not run)
## Not run:
```

```
# JOINT MODELS WITH THREE BINARY MARGINS #
library(GJRM)
############
## Example 9
############
## Generate data
## Correlation between the two equations 0.5 - Sample size 400
set.seed(0)
n <- 400
Sigma <- matrix(0.5, 3, 3); diag(Sigma) <- 1
  <- rMVN(n, rep(0,3), Sigma)
x1 <- round(runif(n)); x2 <- runif(n); x3 <- runif(n)</pre>
f1 <- function(x) cos(pi*2*x) + sin(pi*x)
f2 <- function(x) x+exp(-30*(x-0.5)^2)
y1 \leftarrow ifelse(-1.55 + 2*x1 - f1(x2) + u[,1] > 0, 1, 0)
y2 \leftarrow ifelse(-0.25 - 1.25*x1 + f2(x2) + u[,2] > 0, 1, 0)
y3 <- ifelse(-0.75 + 0.25*x1
                              + u[,3] > 0, 1, 0)
dataSim <- data.frame(y1, y2, y3, x1, x2)</pre>
f.1 <- list(y1 \sim x1 + s(x2),
         y2 \sim x1 + s(x2),
          y3 \sim x1)
out <- gjrm(f.l, data = dataSim, Model = "T",</pre>
          margins = c("probit", "probit", "probit"))
out1 <- gjrm(f.l, data = dataSim, Chol = TRUE, Model = "T",
          margins = c("probit", "probit", "probit"))
conv.check(out)
summary(out)
plot(out, eq = 1)
plot(out, eq = 2)
AIC(out)
BIC(out)
out <- gjrm(f.1, data = dataSim, Model = "T",
          margins = c("probit","logit","cloglog"))
```

```
out1 <- gjrm(f.l, data = dataSim, Chol = TRUE, Model = "T",
             margins = c("probit","logit","cloglog"))
conv.check(out)
summary(out)
plot(out, eq = 1)
plot(out, eq = 2)
AIC(out)
BIC(out)
f.1 <- list(y1 ~ x1 + s(x2),
             y2 \sim x1 + s(x2),
             y3 \sim x1,
               ~ 1, ~ 1, ~ 1)
out1 <- gjrm(f.l, data = dataSim, Chol = TRUE, Model = "T",
              margins = c("probit", "probit", "probit"))
f.1 <- list(y1 \sim x1 + s(x2),
             y2 \sim x1 + s(x2),
             y3 \sim x1,
               ~ 1, ~ s(x2), ~ 1)
out2 <- gjrm(f.1, data = dataSim, Chol = TRUE, Model = "T",</pre>
              margins = c("probit", "probit", "probit"))
f.1 \leftarrow list(y1 \sim x1 + s(x2),
            y2 \sim x1 + s(x2),
             y3 \sim x1,
               \sim x1, \sim s(x2), \sim x1 + s(x2)
out2 <- gjrm(f.1, data = dataSim, Chol = TRUE, Model = "T",</pre>
              margins = c("probit", "probit", "probit"))
f.1 <- list(y1 ~ x1 + s(x2),
             y2 \sim x1 + s(x2),
             y3 \sim x1,
                ~ x1, ~ x1, ~ s(x2))
out2 <- gjrm(f.1, data = dataSim, Chol = TRUE, Model = "T",
              margins = c("probit", "probit", "probit"))
f.1 <- list(y1 ~ x1 + s(x2),
             y2 \sim x1 + s(x2),
             y3 \sim x1,
               \sim x1, \sim x1 + x2, \sim s(x2)
out2 <- gjrm(f.1, data = dataSim, Chol = TRUE, Model = "T",
              margins = c("probit", "probit", "probit"))
f.1 <- list(y1 ~ x1 + s(x2),
            y2 \sim x1 + s(x2),
             y3 \sim x1,
                \sim x1 + x2, \sim x1 + x2, \sim x1 + x2)
```

```
out2 <- gjrm(f.1, data = dataSim, Chol = TRUE, Model = "T",
             margins = c("probit", "probit", "probit"))
jcres1 <- jc.probs(out2, 1, 1, 1, type = "joint", cond = 0,</pre>
                    intervals = TRUE, n.sim = 100)
nw \leftarrow data.frame(x1 = 0, x2 = seq(0, 1, length.out = 100))
jcres2 <- jc.probs(out2, 1, 1, 1, newdata = nw, type = "joint",</pre>
                    cond = 0, intervals = TRUE, n.sim = 100)
############
## Example 10
############
## Generate data
## with double sample selection
set.seed(0)
n <- 5000
Sigma <- matrix(c(1, 0.5, 0.4,
                  0.5, 1, 0.6,
                   0.4, 0.6, 1), 3, 3)
u \leftarrow rMVN(n, rep(0,3), Sigma)
f1 <- function(x) cos(pi*2*x) + sin(pi*x)
f2 <- function(x) x+exp(-30*(x-0.5)^2)
x1 <- runif(n)</pre>
x2 <- runif(n)</pre>
x3 <- runif(n)
x4 <- runif(n)</pre>
y1 \leftarrow 1 + 1.5*x1 - x2 + 0.8*x3 - f1(x4) + u[, 1] > 0
y2 \leftarrow 1 - 2.5*x1 + 1.2*x2 + x3 + u[, 2] > 0
                                                 + u[, 3] > 0
y3 \leftarrow 1.58 + 1.5*x1 - f2(x2)
dataSim <- data.frame(y1, y2, y3, x1, x2, x3, x4)</pre>
f.1 \leftarrow list(y1 \sim x1 + x2 + x3 + s(x4),
            y2 \sim x1 + x2 + x3,
            y3 \sim x1 + s(x2)
out <- gjrm(f.1, data = dataSim, Model = "TSS",</pre>
            margins = c("probit", "probit", "probit"))
conv.check(out)
summary(out)
plot(out, eq = 1)
plot(out, eq = 3)
```

```
prev(out)
prev(out, type = "univariate")
prev(out, type = "naive")
## End(Not run)
## Not run:
# JOINT MODELS WITH BINARY AND CONTINUOUS MARGINS #
# WITH SAMPLE SELECTION
                                   #
library(GJRM)
## Generate data
## Correlation between the two equations and covariate correlation 0.5
## Sample size 2000
############
## Example 11
#############
set.seed(0)
n <- 2000
rh <- 0.5
sigmau \leftarrow matrix(c(1, rh, rh, 1), 2, 2)
    <- rMVN(n, rep(0,2), sigmau)
sigmac <- matrix(rh, 3, 3); diag(sigmac) <- 1</pre>
cov <- rMVN(n, rep(0,3), sigmac)</pre>
    <- pnorm(cov)
cov
bi <- round(cov[,1]); x1 <- cov[,2]; x2 <- cov[,3]
f11 <- function(x) -0.7*(4*x + 2.5*x^2 + 0.7*sin(5*x) + cos(7.5*x))
f12 <- function(x) -0.4*(-0.3 - 1.6*x + \sin(5*x))
f21 \leftarrow function(x) \ 0.6*(exp(x) + sin(2.9*x))
ys \leftarrow 0.58 + 2.5*bi + f11(x1) + f12(x2) + u[, 1] > 0
y < -0.68 - 1.5*bi + f21(x1) +
yo <- y*(ys > 0)
dataSim <- data.frame(ys, yo, bi, x1, x2)</pre>
## CLASSIC SAMPLE SELECTION MODEL
```

```
## the first equation MUST be the selection equation
resp.check(yo[ys > 0], "N")
out <- gjrm(list(ys \sim bi + x1 + x2,
                 yo \sim bi + x1),
                 data = dataSim, Model = "BSS",
                 margins = c("probit", "N"))
conv.check(out)
post.check(out)
summary(out)
AIC(out)
BIC(out)
## SEMIPARAMETRIC SAMPLE SELECTION MODEL
out <- gjrm(list(ys \sim bi + s(x1) + s(x2),
                 yo \sim bi + s(x1)),
                 data = dataSim, Model = "BSS",
                 margins = c("probit", "N"))
conv.check(out)
post.check(out)
AIC(out)
## compare the two summary outputs
## the second output produces a summary of the results obtained when only
## the outcome equation is fitted, i.e. selection bias is not accounted for
summary(out)
summary(out$gam2)
## estimated smooth function plots
## the red line is the true curve
## the blue line is the naive curve not accounting for selection bias
x1.s <- sort(x1[dataSim$ys>0])
f21.x1 \leftarrow f21(x1.s)[order(x1.s)] - mean(f21(x1.s))
plot(out, eq = 2, ylim = c(-1, 0.8)); lines(x1.s, f21.x1, col = "red")
par(new = TRUE)
plot(out$gam2, se = FALSE, lty = 3, lwd = 2, ylim = c(-1, 0.8),
     ylab = "", rug = FALSE)
## IMPUTE MISSING VALUES
n.m <- 10
res <- imputeSS(out, n.m)</pre>
bet <- NA
for(i in 1:n.m){
```

```
dataSim$yo[dataSim$ys == 0] <- res[[i]]</pre>
outg <- gamlss(list(yo \sim bi + s(x1)), data = dataSim)
bet[i] <- coef(outg)["bi"]</pre>
print(i)
}
mean(bet)
##
## SEMIPARAMETRIC SAMPLE SELECTION MODEL with association
## and dispersion parameters
## depending on covariates as well
eq.mu.1 <- ys \sim bi + s(x1) + s(x2)
eq.mu.2 \leftarrow yo \sim bi + s(x1)
eq.sigma <- ~ bi
eq.theta \leftarrow \sim bi + x1
f1 <- list(eq.mu.1, eq.mu.2, eq.sigma, eq.theta)</pre>
out <- gjrm(fl, data = dataSim, Model = "BSS",</pre>
                  margins = c("probit", "N"))
conv.check(out)
post.check(out)
summary(out)
out$sigma
out$theta
jc.probs(out, 0, 0.3, intervals = TRUE)[1:4,]
outC0 <- gjrm(fl, data = dataSim, BivD = "C0", Model = "BSS",
              margins = c("probit", "N"))
conv.check(outC0)
post.check(outC0)
AIC(out, outC0)
BIC(out, outC0)
## IMPUTE MISSING VALUES
n.m <- 10
res <- imputeSS(outC0, n.m)</pre>
################
# or using MICE
################
library(mice)
ys <- dataSim$ys
```

```
dataSim$yo[dataSim$ys == FALSE] <- NA</pre>
dataSim <- dataSim[, -1]</pre>
imp <- mice(dataSim, m = 1, method = c("copulaSS", "", ""),</pre>
           mice.formula = outC0$mice.formula, margins = outC0$margins,
           BivD = outC0$BivD, maxit = 1)
comp.yo <- dataSim$yo</pre>
comp.yo[ys == 0] <- impsyo[[1]]
mean(comp.yo)
#
#
## example using Gumbel copula and normal-gamma margins
############
## Example 12
#############
set.seed(1)
y \leftarrow \exp(-0.68 - 1.5*bi + f21(x1) + u[, 2])
yo <- y*(ys > 0)
dataSim <- data.frame(ys, yo, bi, x1, x2)</pre>
out <- gjrm(list(ys \sim bi + s(x1) + s(x2),
                yo \sim bi + s(x1)),
                data = dataSim, BivD = "G0",
                margins = c("probit", "GA"),
                Model = "BSS")
conv.check(out)
post.check(out)
summary(out)
ATE <- NA
n.m <- 10
res <- imputeSS(out, n.m)</pre>
for(i in 1:n.m){
dataSim$yo[dataSim$ys == 0] <- res[[i]]</pre>
outg <- gamlss(list(yo ~ bi + s(x1)), margin = "GA", data = dataSim)
out$gamlss <- outg
ATE[i] <- AT(out, nm.end = "bi", type = "univariate")$res[2]
print(i)
```

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```
}
AT(out, nm.end = "bi")
mean(ATE)
## End(Not run)
```

gjrmObject

Fitted gjrm object

# Description

A fitted joint model returned by function gjrm and of class "gjrm", "SemiParBIV", "SemiParTRIV", etc.

# Value

fit	List of values and diagnostics extracted from the output of the algorithm.
gam1	Univariate fit for equation 1. See the documentation of mgcv for full details.
gam2, gam3,	Univariate fit for equation 2, equation 3, etc.
coefficients	The coefficients of the fitted model.
weights	Prior weights used during model fitting.
sp	Estimated smoothing parameters of the smooth components.
iter.sp	Number of iterations performed for the smoothing parameter estimation step.
iter.if	Number of iterations performed in the initial step of the algorithm.
iter.inner	Number of iterations performed within the smoothing parameter estimation step.
theta	Estimated dependence parameter linking the two equations.
n	Sample size.
V1 V2 V3	Design matrices associated with the linear predictors.
ΛΙ, ΛΖ, ΛϽ,	Design matrices associated with the inlear predictors.
X1, X2, X3, X1.d2, X2.d2, X3	
	Number of columns of X1, X2, X3, etc.
X1.d2, X2.d2, X3	Number of columns of X1, X2, X3, etc.
X1.d2, X2.d2, X3	Number of columns of X1, X2, X3, etc.
X1.d2, X2.d2, X3 1.sp1, 1.sp2, 1	Number of columns of X1, X2, X3, etc. sp3, Number of smooth components in the equations.
X1.d2, X2.d2, X3  1.sp1, 1.sp2, 1  He	Number of columns of X1, X2, X3, etc.  sp3,  Number of smooth components in the equations.  Penalized -hessian/Fisher. This is the same as HeSh for unpenalized models.
X1.d2, X2.d2, X3 1.sp1, 1.sp2, 1 He HeSh	Number of columns of X1, X2, X3, etcsp3,  Number of smooth components in the equations.  Penalized -hessian/Fisher. This is the same as HeSh for unpenalized models.  Unpenalized -hessian/Fisher.  Inverse of He. This corresponds to the Bayesian variance-covariance matrix used

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edf1, edf2, edf3, ...

Degrees of freedom for the two equations of the fitted bivariate model (and for the third and fourth equations if present. They are calculated when splines are

used.

bs.mgfit List of values and diagnostics extracted from magic in mgcv.

conv. sp If TRUE then the smoothing parameter selection algorithm stopped before reach-

ing the maximum number of iterations allowed.

wor.c Working model quantities.

eta1, eta2, eta3, ...

Estimated linear predictors for the two equations (as well as the third and fourth

equations if present).

y1, y2 Responses of the two equations.

logLik Value of the (unpenalized) log-likelihood evaluated at the (penalized or unpe-

nalized) parameter estimates.

respvec List containing response vectors.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### See Also

gjrm, summary.gjrm

gt.bpm

Gradient test

### Description

gt.bpm can be used to test the hypothesis of absence of endogeneity, correlated model equations/errors or non-random sample selection in binary bivariate probit models.

#### Usage

gt.bpm(x)

#### **Arguments**

Х

A fitted gjrm object.

### Details

The gradient test was first proposed by Terrell (2002) and it is based on classic likelihood theory. See Marra et al. (in press) for full details.

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## Value

It returns a numeric p-value corresponding to the null hypothesis that the correlation,  $\theta$ , is equal to 0.

#### WARNINGS

This test's implementation is only valid for bivariate binary probit models with normal errors.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

### References

Marra G., Radice R. and Filippou P. (2017), Regression Spline Bivariate Probit Models: A Practical Approach to Testing for Exogeneity. *Communications in Statistics - Simulation and Computation*, 46(3), 2283-2298.

Terrell G. (2002), The Gradient Statistic. Computing Science and Statistics, 34, 206-215.

# **Examples**

## see examples for gjrm

H.tri Internal Function

# Description

This and other similar internal functions calculate the Hessian for trivariate binary models.

## Author(s)

Author: Panagiota Filippou

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

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Hazard and survival plots

## **Description**

This function produces either a survival or hazard plot.

# Usage

```
hazsurv.plot(x, eq, newdata, type = "surv", t.range = NULL, t.vec = NULL,
    intervals = TRUE, n.sim = 100, prob.lev = 0.05, shade = FALSE,
    bars = FALSE, ylim, ylab, xlab, pch, ls = 100, baseline = FALSE,
    min.dn = 1e-200, min.pr = 1e-200, max.pr = 1, plot.out = TRUE,
    print.progress = TRUE, ...)
```

### **Arguments**

n.sim

X	A fitted gamlss/gjrm object.
eq	Equation number. This can be ignored for univariate models.
newdata	A data frame or list containing the values of the model covariates at which predictions are required. This must always be provided.
	For the individual survival/hazard/cumulative hazard function, the data frame must have one row containing the values of the model covariates corresponding to the individual of interest.
	For the (sub-)population survival/hazard/cumulative hazard function, the data frame must have as many rows as there are individuals in the (sub-)population of interest. Each row must contain the values of the model covariates of the corresponding individual.
type	The plot to produce, either "surv", "hazard" or "cumhaz". In the excess hazard setting these are, respectively, the net survival, the excess hazard and the cumulative excess hazard.
t.range	Time variable range to be considered for plotting. This must be a vector with only two elements: the minimum and maximum of the time range. If NULL then it is determined automatically based on the observed data.
t.vec	Vector of time values to be considered for plotting. This could also be a single time. Note you cannot provide both t.range and t.vec as they are two mutually exclusive ways of defining the time variable. If NULL then it is determined automatically based on the observed data.
intervals	If TRUE then intervals are also produced.

Number of simulated coefficient vectors from the posterior distribution of the

estimated model parameters. This is used for interval calculations.

prob.lev Overall probability of the left and right tails of the probabilities' distributions

used for interval calculations.

shade If TRUE then it produces shaded regions as confidence bands.

bars If TRUE then the confidence intervals are plotted as bars rather than continuous

curves. If t.vec is used and only one time value is provided, this is the only possible plotting option for the confidence intervals. Note shade and bars are

mutually exclusive.

ylim, ylab, xlab, pch

Usual plot arguments.

1s Length of sequence to use for time variable.

baseline If baseline is desired; this will set all covariate/smooth effects to zero.

min.dn, min.pr, max.pr

Allowed minimum and maximum for estimated probabities and densities for

survival, hazard and cumulative hazard calculations.

plot.out If FALSE then the function does not produce a plot. The default is TRUE.

print.progress If FALSE then the function does not print progress made. The default is TRUE.

... Other arguments to pass to plot.

#### Value

It produces a plot or set of plots.

#### Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

hiv HIV Zambian data

## Description

HIV Zambian data by region, together with polygons describing the regions' shapes.

#### Usage

```
data(hiv)
data(hiv.polys)
```

### Format

hiv is a 6416 row data frame with the following columns

**hivconsent** binary variable indicating consent to test for HIV.

hiv binary variable indicating whether an individual is HIV positive.

age age in years.

**education** years of education.

**region** code identifying region, and matching names(hiv.polys). It can take nine possible values: 1 central, 2 copperbelt, 3 eastern, 4 luapula, 5 lusaka, 6 northwestern, 7 northern, 8 southern, 9 western.

marital never married, currently married, formerly married.

std had a sexually transmitted disease.

highhiv had high risk sex.

condom used condom during last intercourse.

aidscare equal to 1 if would care for an HIV-infected relative.

knowsdiedofaids equal to 1 if know someone who died of HIV.

evertestedHIV equal to 1 if previously tested for HIV.

smoke smoker.

**ethnicity** bemba, lunda (luapula), lala, ushi, lamba, tonga, luvale, lunda (northwestern), mbunda, kaonde, lozi, chewa, nsenga, ngoni, mambwe, namwanga, tumbuka, other.

language English, Bemba, Lozi, Nyanja, Tonga, other.

interviewerID interviewer identifier.

sw survey weights.

hiv.polys contains the polygons defining the areas in the format described below.

#### **Details**

The data frame hiv relates to the regions whose boundaries are coded in hiv.polys. hiv.polys[[i]] is a 2 column matrix, containing the vertices of the polygons defining the boundary of the ith region. names(hiv.polys) matches hiv\$region (order unimportant).

#### Source

The data have been produced as described in:

McGovern M.E., Barnighausen T., Marra G. and Radice R. (2015), On the Assumption of Joint Normality in Selection Models: A Copula Approach Applied to Estimating HIV Prevalence. *Epidemiology*, 26(2), 229-237.

#### References

Marra G., Radice R., Barnighausen T., Wood S.N. and McGovern M.E. (2017), A Simultaneous Equation Approach to Estimating HIV Prevalence with Non-Ignorable Missing Responses. *Journal of the American Statistical Association*, 112(518), 484-496.

#### **Examples**

## Not run:

#:	##:	##:	##	##	##	##	##	##	###	##	##	##	#	##	##	#:	##	#:	##	##	##	##	#:	##	#:	##	#	##	##	##	##	#:	##	#	##	‡#
#:	##:	##:	##	##	##	##	##	##	###	##	##	##	#:	##	‡#	#:	##	#:	##	‡#	##	##	#:	##	#:	##	#	##	‡#	##	##	#:	##	#:	##	‡#

```
library("GJRM")
data("hiv", package = "GJRM")
data("hiv.polys", package = "GJRM")
## The stuff below is useful if the user wishes to employ
## a Markov Random Field (MRF) smoother. It provides
## the instructions to set up polygons automatically
## and the dataset variable needed to fit a model with
## MRF.
# ## hiv.polys was already created and
# ## made available via the call
# ## data("hiv.polys", package = "GJRM")
# ## hiv.polys was created using the code below
# obj <- readRDS("ZMB_adm1.rds")</pre>
# ## RDS Zambian Level 1 file obtained from
# ## http://www.gadm.org.
# pol <- polys.setup(obj)</pre>
# hiv.polys <- pol$polys</pre>
# name <- cbind(names(hiv.polys), pol$names1)</pre>
# name
## last step was to create a factor variable with range
## range(name[,1]) where the numerical values were linked
## to the regions in name[, 2]. This is what was done in
## the hiv dataset; see hiv$region. Specifically,
## the procedure used was
##
# reg <- NULL
# for(i in 1:dim(hiv)[1]){
# if(hiv$region[i] == "Central")
                                   reg[i] <- 1
# if(hiv$region[i] == "Copperbelt")
                                   reg[i] \leftarrow 2
# if(hiv$region[i] == "Eastern")
                                   reg[i] \leftarrow 3
# if(hiv$region[i] == "Luapula")
                                   reg[i] \leftarrow 4
# if(hiv$region[i] == "Lusaka")
                                   reg[i] <- 5
# if(hiv$region[i] == "North-Western") reg[i] <- 6</pre>
# if(hiv$region[i] == "Northern")
                                   reg[i] <- 7
# if(hiv$region[i] == "Southern")
                                   reg[i] <- 8
# if(hiv$region[i] == "Western")
                                   reg[i] <- 9
#
# }
#
```

```
# hiv$region <- as.factor(reg)</pre>
xt <- list(polys = hiv.polys)</pre>
# neighbourhood structure info for MRF
# to use in model specification
# Bivariate brobit model with non-random sample selection
sel.eq \leftarrow hivconsent \sim s(age) + s(education) + s(wealth) +
                    s(region, bs = "mrf", xt = xt, k = 7) +
                    marital + std + age1sex_cat + highhiv +
                    partner + condom + aidscare +
                    knowsdiedofaids + evertestedHIV +
                    smoke + religion + ethnicity +
                    language + s(interviewerID, bs = "re")
out.eq <- hiv
                  \sim s(age) + s(education) + s(wealth) +
                    s(region, bs = "mrf", xt = xt, k = 7) +
                    marital + std + age1sex_cat + highhiv +
                    partner + condom + aidscare +
                    knowsdiedofaids + evertestedHIV +
                    smoke + religion + ethnicity +
                    language
                  \sim s(region, bs = "mrf", xt = xt, k = 7)
theta.eq <-
fl <- list(sel.eq, out.eq, theta.eq)</pre>
# the above model specification is fairly
# complex and it serves to illustrate the
# flexibility of the modelling approach
bss <- gjrm(fl, data = hiv, BivD = "J90", Model = "BSS",
          margins = c("probit", "probit"))
conv.check(bss)
set.seed(1)
sb <- summary(bss)</pre>
sb
plot(bss, eq = 1, seWithMean = TRUE, scheme = 1,
    scale = 0, pages = 1, jit = TRUE)
plot(bss, eq = 2, seWithMean = TRUE, scheme = 1,
    scale = 0, pages = 1, jit = TRUE)
```

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```
prev(bss, sw = hiv$sw, type = "naive")
set.seed(1)
prev(bss, sw = hiv$sw, type = "univariate")
prev(bss, sw = hiv$sw)
lr <- length(hiv.polys)</pre>
prevBYreg <- matrix(NA, lr, 2)</pre>
thetaBYreg <- NA
for(i in 1:lr) {
prevBYreg[i,1] <- prev(bss, sw = hiv$sw, ind = hiv$region==i,</pre>
                        type = "univariate")$res[2]
prevBYreg[i,2] <- prev(bss, sw = hiv$sw, ind = hiv$region==i)$res[2]</pre>
thetaBYreg[i] <- bss$theta[hiv$region==i][1]</pre>
}
zlim <- range(prevBYreg) # to establish a common prevalence range</pre>
par(mfrow = c(1, 3), cex.axis = 1.3)
polys.map(hiv.polys, prevBYreg[,1], zlim = zlim, lab = "",
          cex.lab = 1.5, cex.main = 1.5,
          main = "HIV - Imputation Model")
polys.map(hiv.polys, prevBYreg[,2], zlim = zlim, cex.main = 1.5,
          main = "HIV - Selection Model")
polys.map(hiv.polys, thetaBYreg, rev.col = FALSE, cex.main = 1.7,
          main = expression(paste("Copula parameter (",hat(theta),")")))
sb$CItheta[1,]
## End(Not run)
```

imputeCounter

Imputation of Counterfactual

## **Description**

imputeCounter imputes counterfactual missing values for a gjrm model object.

imputeCounter

### Usage

```
imputeCounter(x, m = 10, nm.end)
```

## **Arguments**

x A fitted gjrm object.

m Number of imputed response vectors.

nm. end Name endogenous variable.

## **Details**

This function generates m sets of imputed values for the outcome of interest under a fitted joint causal model. The algorithm draws parameters from the posterior distribution of the model which are then used to obtain simulated responses (from the posterior predictive distribution of the missing values) via a rejection algorithm. The bound for acceptance/rejection is obtained via a trust region optimisation.

The imputed values are used to create m complete imputed datasets and perform complete data analysis and inference about the parameters of interest using any summary statistics.

#### Value

It returns a list containing m imputed response vectors.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

## References

Robert C. and Casella G. (2004). Monte Carlo Statistical Methods. New York: Springer-Verlag.

Ripley B. D. (1987) Stochastic Simulation. New York: John Wiley & Sons, Inc.

#### See Also

gjrm

#### **Examples**

```
## see examples for gjrm
```

imputeSS 67

imputeSS

Missing values' imputation

# Description

imputeSS imputes missing values for a gjrm model object.

### Usage

```
imputeSS(x, m = 10)
```

### **Arguments**

x A fitted gjrm object.

m Number of imputed response vectors.

### **Details**

This function generates m sets of imputed values for the outcome of interest under a fitted copulaSampleSel model. The algorithm draws parameters from the posterior distribution of copulaSampleSel which are then used to obtain simulated responses (from the posterior predictive distribution of the missing values) via a rejection algorithm. The bound for acceptance/rejection is obtained via a trust region optimisation.

The imputed values are used to create m complete imputed datasets and perform complete data analysis and inference about the parameters of interest using function gamlss() within this package.

### Value

It returns a list containing m imputed response vectors.

#### Author(s)

Authors: Jose Camarena, Giampiero Marra and Rosalba Radice Maintainer: Giampiero Marra <giampiero.marra@ucl.ac.uk>

## References

Robert C. and Casella G. (2004). Monte Carlo Statistical Methods. New York: Springer-Verlag. Ripley B. D. (1987) Stochastic Simulation. New York: John Wiley & Sons, Inc.

### See Also

jc.probs

# **Examples**

```
## see examples for gjrm
```

jc.probs	Joint or conditional probabilities from a fitted joint model

# Description

jc.probs can be used to calculate the joint or conditional probabilities from a fitted joint model with intervals obtained using posterior simulation.

# Usage

```
jc.probs(x, y1, y2, y3 = NULL, newdata, type = "joint", cond = 0,
    intervals = FALSE, n.sim = 100, prob.lev = 0.05, min.pr = 1e-323, max.pr = 1,
        cumul = "no")
```

# Arguments

х	A fitted gjrm object as produced by the respective fitting function.
y1	Value of response for first margin.
y2	Value of response for second margin.
у3	Value of response for third margin if a trivariate model is employed.
newdata	A data frame or list containing the values of the model covariates at which predictions are required. If not provided then predictions corresponding to the original data are returned. When newdata is provided, it should contain all the variables needed for prediction.
type	This argument can take two: "joint" (the probabilities are calculated from the fitted joint model) and "independence" (the calculation is done from univariate fits).
cond	There are three possible values: 0 (joint probabilities are delivered), 1 (conditional probabilities are delivered and conditioning is with the respect to the first margin), 2 (as before but conditioning is with the respect to the second margin).
intervals	If TRUE then intervals for the probabilities are also produced.
n.sim	Number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used for interval calculations.
prob.lev	Overall probability of the left and right tails of the probabilities' distributions used for interval calculations.
min.pr, max.pr	Allowed minimum and maximum for estimated probabities.
cumul	Only used for discrete and continuous margins' case.

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#### **Details**

This function calculates joint or conditional probabilities from a fitted joint model or a model assuming independence, with intervals obtained using posterior simulation.

## Value

res

It returns several values including: estimated probabilities (p12), with lower and upper interval limits (CIpr) if intervals = TRUE, and p1, p2 and p3 (the marginal probabilities).

## Author(s)

Maintainer: Giampiero Marra <giampiero.marra@ucl.ac.uk>

#### See Also

```
GJRM-package, gjrm
```

llpsi

Internal Function

## **Description**

Log-logistic robust function.

#### Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

LM.bpm

Lagrange Multiplier Test (Score Test)

### **Description**

Before fitting a bivariate probit model, LM. bpm can be used to test the hypothesis of absence of endogeneity, correlated model equations/errors or non-random sample selection.

# Usage

```
LM.bpm(formula, data = list(), weights = NULL, subset = NULL, Model,
    hess = TRUE)
```

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### Arguments

formula	A list of two formulas, one for equation 1 and the other for equation 2. s terms are used to specify smooth smooth functions of predictors. Note that if Model = "BSS" then the first formula MUST refer to the selection equation.
data	An optional data frame, list or environment containing the variables in the model. If not found in data, the variables are taken from environment(formula).
weights	Optional vector of prior weights to be used in fitting.
subset	Optional vector specifying a subset of observations to be used in the fitting process.
Model	It indicates the type of model to be used in the analysis. Possible values are "B" (bivariate model) and "BSS" (bivariate model with sample selection). The two marginal equations have probit links.
hess	If FALSE then the expected (rather than observed) information matrix is employed.

#### **Details**

This Lagrange multiplier test (also known as score test) is used here for testing the null hypothesis that  $\theta$  is equal to 0 (i.e. no endogeneity, non-random sample selection or correlated model equations/errors, depending on the model being fitted). Its main advantage is that it does not require an estimate of the model parameter vector under the alternative hypothesis. Asymptotically, it takes a Chi-squared distribution with one degree of freedom. Full details can be found in Marra et al. (2014) and Marra et al. (2017).

## Value

It returns a numeric p-value corresponding to the null hypothesis that the correlation,  $\theta$ , is equal to 0.

#### WARNINGS

This test's implementation is ONLY valid for bivariate binary probit models with normal errors.

### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### References

Marra G., Radice R. and Filippou P. (2017), Regression Spline Bivariate Probit Models: A Practical Approach to Testing for Exogeneity. *Communications in Statistics - Simulation and Computation*, 46(3), 2283-2298.

Marra G., Radice R. and Missiroli S. (2014), Testing the Hypothesis of Absence of Unobserved Confounding in Semiparametric Bivariate Probit Models. *Computational Statistics*, 29(3-4), 715-741.

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## See Also

```
gjrm
```

# **Examples**

```
## see examples for gjrm
```

1mc

Linear Model Fitting with Constraints

# Description

Linear model fitting with positivity and sum-to-one constraints on the model's coefficients.

# Usage

```
lmc(y, X, start.v = NULL, lambda = 1, pen = "none", gamma = 1, a = 3.7)
```

# Arguments

У	Response vector.
X	Design matrix.
start.v	Starting values.
lambda	Tuning parameter.
pen	Type of penalty. Choices are: none, ridge, lasso, alasso, scad.
gamma	Power parameter of adaptive lasso.
а	Scad parameter.

# **Details**

Linear model fitting with positivity and sum-to-one constraints on the model's coefficients.

# Value

The function returns an object of class 1mc.

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### **Examples**

```
## Not run:
library(GJRM)
set.seed(1)
   <- 1000
beta <- c(0.07, 0.08, 0.21, 0.12, 0.15, 0.17, 0.2)
    <- length(beta)
     <- matrix(runif(n*l), n, l)
     <- X%*%beta + rnorm(n)
out <- lmc(y, X)
conv.check(out)
out1 <- lmc(y, X, start.v = beta)
conv.check(out1)
coef(out)
                             # estimated coefficients
round(out$c.coefficients, 3) # constrained coefficients
sum(out$c.coefficients)
round(out1$c.coefficients, 3)
sum(out1$c.coefficients)
# penalised estimation
out1 <- lmc(y, X, pen = "alasso", lambda = 0.02)</pre>
conv.check(out1)
coef(out1)
round(out1$c.coefficients, 3)
sum(out1$c.coefficients)
AIC(out, out1)
BIC(out, out1)
round(cbind(out$c.coefficients, out1$c.coefficients), 3)
# scad
beta <- c(0.2, 0, 0, 0.02, 0.01, 0.01, 0.01, 0.08, 0.21, 0.12, 0.15, 0.17, 0.02)
   <- length(beta)
   <- matrix(runif(n*l), n, l)</pre>
```

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```
y <- X%*%beta + rnorm(n)

out1 <- lmc(y, X, pen = "scad", lambda = 0.01)
conv.check(out1)

coef(out1)
sum(out1$c.coefficients)

round(cbind(beta, out1$c.coefficients), 2)

## End(Not run)</pre>
```

logLik.SemiParBIV

Extract the log likelihood for a fitted copula model

### **Description**

It extracts the log-likelihood for a fitted gjrm model.

## Usage

```
## S3 method for class 'SemiParBIV'
logLik(object, ...)
```

#### **Arguments**

object A fitted gjrm object.
... Un-used for this function.

# **Details**

Modification of the classic logLik which accounts for the estimated degrees of freedom used in gjrm. This function is provided so that information criteria work correctly by using the correct number of degrees of freedom.

# Value

Standard logLik object.

# Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

#### See Also

```
AIC, BIC
```

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mb	Nonparametric (worst-case and I	V) Manski's bounds
----	---------------------------------	--------------------

### **Description**

mb can be used to calculate the (worst-case and IV) Manski's bounds and confidence interval covering the true effect of interest with a fixed probability.

### Usage

```
mb(treat, outc, IV = NULL, Model, B = 100, sig.lev = 0.05)
```

#### **Arguments**

treat	Binary treatment/selection variable.
outc	Binary outcome variable.
IV	An instrumental binary variable can be used if available.
Model	Possible values are "B" (model with endogenous variable) and "BSS" (model with non-random sample selection).
В	Number of bootstrap replicates. This is used to obtain some components needed for confidence interval calculations.
sig.lev	Significance level.

#### **Details**

Based on Manski (1990), this function returns the nonparametric lower and upper (worst-case) Manski's bounds for the average treatment effect (ATE) when Model = "B" or prevalence when Model = "BSS". When an IV is employed the function returns IV Manski bounds.

For comparison, it also returns the estimated effect assuming random assignment (i.e., the treatment received or selection relies on the assumption of ignorable observed and unobserved selection). Note that this is equivalent to what provided by AT or prev when type = "naive", and is different from what obtained by AT or prev when type = "univariate" as observed confounders are accounted for and the assumption here is of ignorable unobserved selection.

A confidence interval covering the true ATE/prevalence with a fixed probability is also provided. This is based on the approach described in Imbens and Manski (2004). NOTE that this interval is typically very close (if not identical) to the lower and upper bounds.

The ATE can be at most 1 (or 100 in percentage) and the worst-case Manski's bounds have width 1. This means that 0 is always included within the possibilities of these bounds. Nevertheless, this may be useful to check whether the effect from a bivariate recursive model is included within the possibilities of the bounds.

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When estimating a prevalance the worst-case Manski's bounds have width equal to the non-response probability, which provides a measure of the uncertainty about the prevalence caused by non-response. Again, this may be useful to check whether the prevalence from a bivariate non-random sample selection model is included within the possibilities of the bounds.

See gjrm for some examples.

#### Value

LB, UP Lower and upper bounds for the true effect of interest.

CI Confidence interval covering the true effect of interest with a fixed probability.

ate.ra Estimated effect of interest assuming random assignment.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### References

Manski C.F. (1990), Nonparametric Bounds on Treatment Effects. *American Economic Review, Papers and Proceedings*, 80(2), 319-323.

Imbens G.W. and Manski C.F (2004), Confidence Intervals for Partially Identified Parameters. *Econometrica*, 72(6), 1845-1857.

# See Also

gjrm

# **Examples**

## see examples for gjrm

meps

MEPS data

# Description

2008 MEPS data.

#### Usage

data(meps)

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#### **Format**

```
bmi body mass index.

age age in years.

gender equal to 1 if male.

race levels: 2 white, 3 black, 4 native American, 5 others.

education years of education.

health levels: 5 excellent, 6 very good, 7 good, 8 fair, 9 poor.

limitation equal to 1 if health limits physical activity.

region levels: 2 northeast, 3 mid-west, 4 south, 5 west.

private equal to 1 if individual has private health insurance.

visits.hosp equal to 1 if at least one visit to hospital outpatient departments.

diabetes equal to 1 if diabetic.

hypertension equal to 1 if hypertensive.

hyperlipidemia equal to 1 if hyperlipidemic.

income income (000's).
```

#### Source

The data have been obtained from http://www.meps.ahrq.gov/.

# Examples

## Not run:

```
library("GJRM")
data("meps", package = "GJRM")
# Bivariate brobit models with endogenous treatment
treat.eq <- private ~ s(bmi) + s(income) + s(age) + s(education) +
               as.factor(health) + as.factor(race) +
               as.factor(limitation) + as.factor(region) +
               gender + hypertension + hyperlipidemia + diabetes
out.eq <- visits.hosp ~ private + s(bmi) + s(income) + s(age) +
                 s(education) + as.factor(health) +
                 as.factor(race) + as.factor(limitation) +
                 as.factor(region) + gender + hypertension +
                 hyperlipidemia + diabetes
```

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```
f.list <- list(treat.eq, out.eq)</pre>
      <- c("probit", "probit")
     <- gjrm(f.list, data = meps, margins = mr, Model = "B")</pre>
bpN
      <- gjrm(f.list, data = meps, margins = mr, BivD = "F", Model = "B")</pre>
bpC0 <- gjrm(f.list, data = meps, margins = mr, BivD = "C0", Model = "B")</pre>
bpC180 <- gjrm(f.list, data = meps, margins = mr, BivD = "C180", Model = "B")</pre>
bpJ0 <- gjrm(f.list, data = meps, margins = mr, BivD = "J0", Model = "B")</pre>
bpJ180 <- gjrm(f.list, data = meps, margins = mr, BivD = "J180", Model = "B")</pre>
bpG0 <- gjrm(f.list, data = meps, margins = mr, BivD = "G0", Model = "B")</pre>
bpG180 <- gjrm(f.list, data = meps, margins = mr, BivD = "G180", Model = "B")</pre>
conv.check(bpJ0)
AIC(bpN, bpF, bpC0, bpC180, bpJ0, bpJ180, bpG0, bpG180)
set.seed(1)
summary(bpJ0)
#dev.copy(postscript, "contplot.eps")
#dev.off()
par(mfrow = c(2, 2), mar = c(4.5, 4.5, 2, 2),
    cex.axis = 1.6, cex.lab = 1.6)
plot(bpJ0, eq = 1, seWithMean = TRUE, scale = 0, shade = TRUE,
     pages = 1, jit = TRUE)
#dev.copy(postscript, "spline1.eps")
#dev.off()
par(mfrow = c(2, 2), mar = c(4.5, 4.5, 2, 2),
    cex.axis = 1.6, cex.lab = 1.6)
plot(bpJ0, eq = 2, seWithMean = TRUE, scale = 0, shade = TRUE,
     pages = 1, jit = TRUE)
#dev.copy(postscript, "spline2.eps")
#dev.off()
set.seed(1)
AT(bpJ0, nm.end = "private", hd.plot = TRUE, cex.axis = 1.5,
   cex.lab = 1.5, cex.main = 1.6)
#dev.copy(postscript, "hd.plotAT.eps")
#dev.off()
AT(bpJ0, nm.end = "private", type = "univariate")
AT(bpJ0, nm.end = "private", type = "naive")
## End(Not run)
#
```

78 OR

numgh	Internal Function	

## **Description**

This and other similar internal functions calculate numerical derivatives.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

OR Causal odds ratio of a binary/continuous/discrete endogenous variable

# Description

OR can be used to calculate the causal odds ratio of a binary/continuous/discrete endogenous predictor/treatment, with corresponding interval obtained using posterior simulation.

# Usage

```
OR(x, nm.end, E = TRUE, treat = TRUE, type = "joint", ind = NULL,
    n.sim = 100, prob.lev = 0.05, length.out = NULL, hd.plot = FALSE,
    or.plot = FALSE,
    main = "Histogram and Kernel Density of Simulated Odds Ratios",
    xlab = "Simulated Odds Ratios", ...)
```

# Arguments

x	A fitted gjrm object.
nm.end	Name of the endogenous variable.
E	If TRUE then OR calculates the sample OR. If FALSE then it calculates the sample OR for the treated individuals only.
treat	If TRUE then OR calculates the OR using the treated only. If FALSE then it calculates the ratio using the control group. This only makes sense if E = FALSE.
type	This argument can take three values: "naive" (the effect is calculated ignoring the presence of observed and unobserved confounders), "univariate" (the effect is obtained from the univariate model which neglects the presence of unobserved confounders) and "joint" (the effect is obtained from the bivariate model which accounts for observed and unobserved confounders).

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ind	Binary logical variable. It can be used to calculate the OR for a subset of the data. Note that it does not make sense to use ind when some observations are excluded from the OR calculation (e.g., when using E = FALSE).
n.sim	Number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used when delta = FALSE. It may be increased if more precision is required.
prob.lev	Overall probability of the left and right tails of the OR distribution used for interval calculations.
length.out	Ddesired length of the sequence to be used when calculating the effect that a continuous/discrete treatment has on a binary outcome.
hd.plot	If TRUE then a plot of the histogram and kernel density estimate of the simulated odds ratios is produced. This can only be produced when binary responses are used.
or.plot	For the case of continuous/discrete endogenous variable and binary outcome, if TRUE then a plot (on the log scale) showing the odd ratios that the binary outcome is equal to 1 for each incremental value of the endogenous variable and respective intervals is produced.
main	Title for the plot.
xlab	Title for the x axis.
• • •	Other graphics parameters to pass on to plotting commands. These are used only when hd.plot = TRUE.

# **Details**

OR calculates the causal odds ratio for a binary/continuous/discrete treatment. Posterior simulation is used to obtain a confidence/credible interval.

# Value

prob.lev	Probability level used.
sim.OR	It returns a vector containing simulated values of the average OR. This is used to calculate intervals.
Ratios	For the case of continuous/discrete endogenous treatment and binary outcome, it returns a matrix made up of three columns containing the odds ratios for each incremental value in the endogenous variable and respective intervals.

# Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

# See Also

GJRM-package, gjrm

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PΕ

Partial effect from a binary bivariate model

## **Description**

PE can be used to calculate the sample treatment effect from a a binary bivariate model, with corresponding interval obtained using posterior simulation.

# Usage

```
PE(x1, idx, n.sim = 100, prob.lev = 0.05,
   hd.plot = FALSE,
   main = "Histogram and Kernel Density of Simulated Average Effects",
   xlab = "Simulated Average Effects", ...)
```

## **Arguments**

x1	A fitted gjrm object.
idx	This is useful to pick a particular individual and must be provided.
n.sim	Number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used when delta = FALSE. It may be increased if more precision is required.
prob.lev	Overall probability of the left and right tails of the AT distribution used for interval calculations.
hd.plot	If TRUE then a plot of the histogram and kernel density estimate of the simulated average effects is produced.
main	Title for the plot.
xlab	Title for the x axis.
• • •	Other graphics parameters to pass on to plotting commands. These are used only when hd.plot = TRUE.

#### **Details**

PE measures the sample average effect from a binary bivariate model when a binary response (associated with a continuous outcome) takes values 0 and 1. Posterior simulation is used to obtain a confidence/credible interval.

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### See Also

```
GJRM-package, gjrm
```

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pen	In	ternal Function

#### **Description**

It provides an overall penalty matrix in a format suitable for estimation conditional on smoothing parameters.

#### Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

plot.SemiParBIV

Plotting function

### **Description**

It takes a fitted gjrm object produced by gjrm() and plots the estimated smooth functions on the scale of the linear predictors. This function is a wrapper of plot.gam() in mgcv. Please see the documentation of plot.gam() for full details.

#### **Usage**

```
## S3 method for class 'SemiParBIV'
plot(x, eq, ...)
```

### **Arguments**

x A fitted gjrm object.

eq The equation from which smooth terms should be considered for printing.

Other graphics parameters to pass on to plotting commands, as described for plot.gam() in mgcv.

#### Details

This function produces plots showing the smooth terms of a fitted semiparametric bivariate probit model. In the case of 1-D smooths, the x axis of each plot is labelled using the name of the regressor, while the y axis is labelled as s(regr, edf) where regr is the regressor's name, and edf the effective degrees of freedom of the smooth. For 2-D smooths, perspective plots are produced with the x axes labelled with the first and second variable names and the y axis is labelled as s(var1, var2, edf), which indicates the variables of which the term is a function and the edf for the term.

If seWithMean = TRUE then the intervals include the uncertainty about the overall mean. Note that the smooths are still shown centred. The theoretical arguments and simulation study of Marra and Wood (2012) suggest that seWithMean = TRUE results in intervals with close to nominal frequentist coverage probabilities.

82 polys.map

## Value

The function generates plots.

#### WARNING

The function can not deal with smooths of more than 2 variables.

## Author(s)

Maintainer: Giampiero Marra <giampiero.marra@ucl.ac.uk>

### References

Marra G. and Wood S.N. (2012), Coverage Properties of Confidence Intervals for Generalized Additive Model Components. *Scandinavian Journal of Statistics*, 39(1), 53-74.

#### See Also

gjrm

polys.map

Geographic map with regions defined as polygons

## **Description**

This function produces a map with geographic regions defined by polygons. It is essentially the same function as polys.plot() in mgcv but with added arguments zlim and rev.col and a wider set of choices for scheme.

# Usage

```
polys.map(lm, z, scheme = "gray", lab = "", zlim, rev.col = TRUE, ...)
```

## **Arguments**

lm	Named list of matrices where each matrix has two columns. The matrix rows each define the vertex of a boundary polygon.
z	A vector of values associated with each area (item) of 1m.
scheme	Possible values are "heat", "terrain", "topo", "cm" and "gray", indicating how to fill the polygons in accordance with the value of z.
lab	label for plot.

polys.setup 83

zlim If missing then the range of z will be chosen using pretty(z) otherwise the

range provided will be used.

rev.col If FALSE then coloring scheme is not reversed.

. . . other arguments to pass to plot.

#### **Details**

See help file of polys.plot in mgcv.

#### Value

It produces a plot.

### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

polys.setup Set up geographic polygons

## **Description**

This function creates geographic polygons in a format suitable for smoothing.

## Usage

```
polys.setup(object)
```

# Arguments

object An RDS file object as extracted from http://www.gadm.org.

### Value

It produces a list with polygons (polys), and various names (names0, names1 - first level of aggregation, names2 - second level of aggregation).

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

Thanks to Guy Harling for suggesting the implementation of this function.

84 post.check

#### **Examples**

?hiv

post.check	Diagnostic plots for discrete/continuous re	sponse margin

## **Description**

It produces diagnostic plots based on (randomised) quantile residuals.

### Usage

```
post.check(x, main = "Histogram and Density Estimate of Residuals",
    main2 = "Histogram and Density Estimate of Residuals",
    xlab = "Quantile Residuals", xlab2 = "Quantile Residuals",
    intervals = FALSE, n.sim = 100, prob.lev = 0.05, ...)
```

# Arguments

x	A fitted gjrm object.
main	Title for the plot.
main2	Title for the plot in the second row. This comes into play only when fitting models with two non-binary margins.
xlab	Title for the x axis.
xlab2	Title for the x axis in the second row. As above.
intervals	If TRUE then intervals for the qqplots are produced.
n.sim	Number of replicate datasets used to simulate quantiles of the residual distribution.
prob.lev	Overall probability of the left and right tails of the probabilities' distributions used for interval calculations.
	Other graphics parameters to pass on to plotting commands.

# **Details**

If the model fits the response well then the plots should look normally distributed. When fitting models with discrete and/or continuous margins, four plots will be produced. In this case, the arguments main2 and xlab2 come into play and allow for different labelling across the plots.

pred.gp 85

#### Value

gin when fitting a model that involves a binary response.

qr1 As above but for first equation (this applies when fitting models with continu-

ous/discrete margins).

qr2 As above but for second equation.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### See Also

gjrm

### Description

It takes a fitted gamlss object produced by gamlss() and produces the desired quntities and respective intervals.

## Usage

```
pred.gp(x, p = 0.5, newdata, n.sim = 100, prob.lev = 0.05)
```

### **Arguments**

x A fitted gamlss object.

p Value of p.

newdata A data frame or list containing the values of the model covariates at which pre-

dictions are required. If not provided then predictions corresponding to the original data are returned. When newdata is provided, it should contain all the vari-

ables needed for prediction.

n.sim The number of simulated coefficient vectors from the posterior distribution of

the estimated model parameters. This is used to calculate intervals. It may be

increased if more precision is required.

prob. lev Probability of the left and right tails of the posterior distribution used for interval

calculations.

#### Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

#### See Also

gamlss

86 predict.CopulaCLM

pred.mvt	Function to predict mean and variance of marginal distributions, as well as Kendall's tau

# Description

It takes a fitted gjrm object produced by gjrm() and produces predictions and respective intervals.

#### Usage

```
pred.mvt(x, eq, fun = "mean", n.sim = 100, prob.lev = 0.05, smooth.no = NULL, ...)
```

# Arguments

x	A fitted gjrm object.
eq	The equation number.
fun	Either mean, variance or tau.
n.sim	The number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used to calculate intervals. It may be increased if more precision is required.
prob.lev	Probability of the left and right tails of the posterior distribution used for interval calculations.
smooth.no	Smooth number if the interest is in a particular smooth and not the additive predictor(s).
	Other parameters.

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

# See Also

gjrm

Prediction function
---------------------

## **Description**

It takes a fitted gjrm object for the ordinal-continuous case and, for each equation, produces predictions for a new set of values of the model covariates or the original values used for the model fit. Standard errors of predictions can be produced and are based on the posterior distribution of the model coefficients.

predict.SemiParBIV 87

#### Usage

```
## S3 method for class 'CopulaCLM'
predict(object, eq, type = "link", ...)
```

# Arguments

object A fitted gjrm object.

eq The equation to be considered for prediction.

type Type of prediction.

... Other arguments as in predict.gam() in mgcv.

### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### See Also

gjrm

# **Description**

It takes a fitted gjrm object and, for each equation, produces predictions for a new set of values of the model covariates or the original values used for the model fit. Standard errors of predictions can be produced and are based on the posterior distribution of the model coefficients. This function is a wrapper for predict.gam() in mgcv. Please see the documentation of predict.gam() for full details.

# Usage

```
## S3 method for class 'SemiParBIV'
predict(object, eq, ...)
```

# Arguments

object A fitted gjrm object.

eq The equation to be considered for prediction.

... Other arguments as in predict.gam() in mgcv.

88 prev

#### WARNINGS

When type = "response" this function will provide prediction assuming that the identity link function is adopted. This means that type = "link" and type = "response" will produce the same results, which for some distributions is fine. This is because, for internal reasons, the model object used always assumes an identity link. There are other functions in the package which will produce predictions for the response correctly and we are currently working on extending them to all models in the package. For all the other type values the function will produce the correct results.

When predicting based on a new data set, this function can not return correct predictions for models based on a BivD value of "C0C90", "C0C270", "C180C90", "C180C270", "G0G90", "G0G270", "G180G90", "G180G270", "J0J90", "J0J270", "J180J90" or "J180J270".

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### See Also

gjrm

prev

Estimated overall prevalence from sample selection model

### Description

prev can be used to calculate the overall estimated prevalence from a sample selection model with binay outcome, with corresponding interval obtained using the delta method or posterior simulation.

#### Usage

```
prev(x, sw = NULL, type = "joint", ind = NULL, delta = FALSE,
    n.sim = 100, prob.lev = 0.05, hd.plot = FALSE,
    main = "Histogram and Kernel Density of Simulated Prevalences",
    xlab = "Simulated Prevalences", ...)
```

#### **Arguments**

x A fitted gjrm object.

sw Survey weights.

type

This argument can take three values: "naive" (the prevalence is calculated ignoring the presence of observed and unobserved confounders), "univariate" (the prevalence is obtained from the univariate probit/single imputation model which neglects the presence of unobserved confounders) and "joint" (the prevalence is obtained from the bivariate/trivariate model which accounts for observed and unobserved confounders).

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ind	Binary logical variable. It can be used to calculate the prevalence for a subset of the data.
delta	If TRUE then the delta method is used for confidence interval calculations, otherwise Bayesian posterior simulation is employed.
n.sim	Number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used when delta = FALSE. It may be increased if more precision is required.
prob.lev	Overall probability of the left and right tails of the prevalence distribution used for interval calculations.
hd.plot	If TRUE then a plot of the histogram and kernel density estimate of the simulated prevalences is produced. This can only be produced when delta = FALSE.
main	Title for the plot.
xlab	Title for the x axis.
	Other graphics parameters to pass on to plotting commands. These are used only when hd.plot = TRUE.

## **Details**

prev estimates the overall prevalence of a disease (e.g., HIV) when there are missing values that are not at random. An interval for the estimated prevalence can be obtained using the delta method or posterior simulation.

## Value

res	It returns three values: lower confidence interval limit, estimated prevalence and upper confidence interval limit.
prob.lev	Probability level used.
sim.prev	If delta = FALSE then it returns a vector containing simulated values of the prevalence. This is used to calculate an interval.

## Author(s)

Authors: Giampiero Marra, Rosalba Radice, Guy Harling, Mark E McGovern

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### References

Marra G., Radice R., Barnighausen T., Wood S.N. and McGovern M.E. (2017), A Simultaneous Equation Approach to Estimating HIV Prevalence with Non-Ignorable Missing Responses. *Journal of the American Statistical Association*, 112(518), 484-496.

## See Also

GJRM-package, gjrm

print.AT

Print an AT object

# Description

The print method for an AT object.

# Usage

```
## S3 method for class 'AT'
print(x, ...)
```

## **Arguments**

x AT object produced by AT().

... Other arguments.

# **Details**

print.AT prints the lower confidence interval limit, estimated AT and upper confidence interval limit.

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

## See Also

ΑT

print.copulaSampleSel Print a copulaSampleSel object

# Description

The print method for a copulaSampleSel object.

print.gamlss 91

## Usage

```
## S3 method for class 'copulaSampleSel'
print(x, ...)
```

### **Arguments**

- x copulaSampleSel object.
- ... Other arguments.

#### **Details**

It prints out the family, model equations, total number of observations, estimated association coefficient, etc for the penalized or unpenalized model.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

print.gamlss

Print a gamlss object

# **Description**

The print method for a gamlss object.

# Usage

```
## S3 method for class 'gamlss' print(x, ...)
```

#### **Arguments**

- x gamlss object produced by gamlss().
- ... Other arguments.

#### **Details**

print.gamlss prints out the family, model equations, total number of observations, etc for the penalized or unpenalized model.

92 print.gjrm

#### Author(s)

Maintainer: Giampiero Marra < giampiero . marra@ucl.ac.uk>

## See Also

```
gamlss
```

print.gjrm

Print a gjrm object

# Description

The print method for a gjrm object.

# Usage

```
## S3 method for class 'gjrm'
print(x, ...)
```

# Arguments

x gjrm object produced by gjrm().

... Other arguments.

# **Details**

print.gjrm prints out the family, model equations, total number of observations, estimated association coefficient, etc for the penalized or unpenalized model.

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

# See Also

gjrm

print.mb 93

print.mb

Print an mb object

# Description

The print method for an mb object.

# Usage

```
## S3 method for class 'mb'
print(x, ...)
```

# Arguments

x mb object produced by mb().

... Other arguments.

# **Details**

print.mb prints the lower and upper bounds, confidence interval, and effect assuming random assignment.

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

## See Also

mb

print.OR

Print an OR object

# Description

The print method for an OR object.

94 print.PE

## Usage

```
## S3 method for class 'OR'
print(x, ...)
```

# Arguments

- x OR object produced by OR().
- ... Other arguments.

## **Details**

print.OR prints the lower confidence interval limit, estimated OR and upper confidence interval limit

#### Author(s)

Maintainer: Giampiero Marra <giampiero.marra@ucl.ac.uk>

## See Also

OR

print.PE

Print an PE object

# **Description**

The print method for an PE object.

# Usage

```
## S3 method for class 'PE'
print(x, ...)
```

# **Arguments**

```
x PE object produced by PE().
```

... Other arguments.

print.prev 95

## **Details**

print.PE prints the lower confidence interval limit, estimated PE and upper confidence interval limit

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

#### See Also

PΕ

print.prev

Print an prev object

# Description

The print method for an prev object.

# Usage

```
## S3 method for class 'prev' print(x, ...)
```

#### **Arguments**

x prev object produced by prev().

... Other arguments.

# **Details**

print.prev prints the lower interval limit, estimated prevalence and upper interval limit.

#### Author(s)

Maintainer: Giampiero Marra <giampiero.marra@ucl.ac.uk>

## See Also

prev

96 print.SemiParBIV

print.RR

Print an RR object

# Description

The print method for an RR object.

# Usage

```
## S3 method for class 'RR'
print(x, ...)
```

## **Arguments**

x RR object produced by RR().

... Other arguments.

# **Details**

print.RR prints the lower confidence interval limit, estimated RR and upper confidence interval limit.

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

## See Also

RR

print.SemiParBIV

Print a SemiParBIV object

# Description

The print method for a SemiParBIV object.

print.SemiParROY 97

## Usage

```
## S3 method for class 'SemiParBIV'
print(x, ...)
```

# **Arguments**

- x SemiParBIV object.
- ... Other arguments.

#### **Details**

It prints out the family, model equations, total number of observations, estimated association coefficient and total effective degrees of freedom for the penalized or unpenalized model.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

print.SemiParROY

Print a SemiParROY object

# **Description**

The print method for a SemiParROY object.

# Usage

```
## S3 method for class 'SemiParROY'
print(x, ...)
```

#### **Arguments**

- x SemiParROY object.... Other arguments.
- ... Other arguments

### **Details**

It prints out the family, model equations, total number of observations, estimated association coefficient, etc for the penalized or unpenalized model.

98 probm

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

print.SemiParTRIV

Print a SemiParTRIV object

# Description

The print method for a SemiParTRIV object.

# Usage

```
## S3 method for class 'SemiParTRIV'
print(x, ...)
```

# **Arguments**

x SemiParTRIV object.

... Other arguments.

#### **Details**

It prints out the family, model equations, total number of observations, estimated association coefficient and total effective degrees of freedom for the penalized or unpenalized model.

### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

probm

Internal Function

# Description

Internal fitting function.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

regH

regH	Internal Function	

# **Description**

It applies one of two regularisations on the information matrix if desired. These are based on the Cholesky and eigen decompositions.

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

resp.check

Plots for response variable

## Description

It produces a histogram of the response along with the estimated density from the assumed distribution as well as a normal Q-Q plot for the (randomised) normalised quantile response. It also provides the log-likelihood for AIC calculation, for instance.

#### Usage

## **Arguments**

у	Response.
margin	The distributions allowed are: normal ("N"), log-normal ("LN"), generelised Pareto ("GP"), discrete generelised Pareto ("DGP"), Gumbel ("GU"), reverse Gumbel ("rGU"), logistic ("LO"), Weibull ("WEI"), inverse Gaussian ("iG"), gamma ("GA"), Dagum ("DAGUM"), Singh-Maddala ("SM"), beta ("BE"), Fisk ("FISK"), Poisson ("PO"), zero truncated Poisson ("ZTP"), negative binomial - type I ("NBI"), negative binomial - type II ("NBII"), Poisson inverse Gaussian ("PIG").
main	Title for the plot.
xlab	Title for the x axis.

rMVN

print.par If TRUE then the estimated parameters used to construct the plots are returned.

plots If FALSE then no plots are produced and only parameter estimates returned.

loglik If TRUE then it returns the logLik.

os If TRUE then the estimated parameters are returned on the original scale.

intervals If TRUE then intervals for the qqplot are produced.

n.sim Number of replicate datasets used to simulate quantiles of the residual distribu-

tion.

prob.lev Overall probability of the left and right tails of the probabilities' distribution

used for interval calculations.

i.f Internal fitting option. This is not for user purposes.

min.dn, min.pr, max.pr

Allowed minimum and maximum for estimated probabities and densities for

parameter estimation.

... Other graphics parameters to pass on to plotting commands.

#### **Details**

Prior to fitting a model with discrete and/or continuous margins, the distributions for the responses may be chosen by looking at the histogram of the response along with the estimated density from the assumed distribution, and at the normalised quantile responses. These will provide a rough guide to the adequacy of the chosen distribution. The latter are defined as the quantile standard normal function of the cumulative distribution function of the response with scale and location estimated by MLE. These should behave approximately as normally distributed variables (even though the original observations are not). Therefore, a normal Q-Q plot is appropriate here.

If loglik = TRUE then this function also provides the log-likelihood for AIC calculation, for instance.

The shapiro test can also be performed.

#### Author(s)

Maintainer: Giampiero Marra < giampiero . marra@ucl.ac.uk>

#### See Also

gjrm

### Description

This function simply generates random multivariate normal variates.

#### Author(s)

Maintainer: Giampiero Marra < giampiero . marra@ucl.ac.uk>

rob.const 101

rob.const

Bootstrap procedure to help select the robust constant in a GAMLSS

# Description

It helps finding the robust constant for a GAMLSS.

# Usage

```
rob.const(x, B = 100)
```

# Arguments

x A fitted gjrm object.

B Number of bootstrap replicates.

## **Details**

It helps finding the robust constant for a GAMLSS based on the mean or median.

### Value

rc Robust constant used in fitting.

sw Sum of weights for each bootstrap replicate.

m1 Mean.
m2 Median.

## Author(s)

Maintainer: Giampiero Marra <giampiero.marra@ucl.ac.uk>

# See Also

gamlss

rob.int

rob.int

Tool for tuning bounds of integral in robust models

# Description

Tool for tuning bounds of integral in robust GAMLSS.

# Usage

```
rob.int(x, rc, l.grid = 1000, tol = 1e-4, var.range = NULL)
```

## **Arguments**

x A fitted gjrm object, typically from a non-robust fit.

rc Robust tuning constant.

1.grid Length of grid.

tol Tolerance

var.range Range of values, min and max, to use in calculations.

# **Details**

Tool for tuning bounds of integral in robust GAMLSS.

# Value

1B, uB Lower and upper bounds.

## Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

# See Also

gamlss

RR 103

RR

Causal risk ratio of a binary/continuous/discrete endogenous variable

# Description

RR can be used to calculate the causal risk ratio of a binary/continuous/discrete endogenous predictor/treatment, with corresponding interval obtained using posterior simulation.

# Usage

```
RR(x, nm.end, E = TRUE, treat = TRUE, type = "joint", ind = NULL,
    n.sim = 100, prob.lev = 0.05, length.out = NULL, hd.plot = FALSE,
    rr.plot = FALSE,
    main = "Histogram and Kernel Density of Simulated Risk Ratios",
    xlab = "Simulated Risk Ratios", ...)
```

# Arguments

X	A fitted gjrm object.
nm.end	Name of the endogenous variable.
Е	If TRUE then RR calculates the sample RR. If FALSE then it calculates the sample RR for the treated individuals only.
treat	If TRUE then RR calculates the RR using the treated only. If FALSE then it calculates the ratio using the control group. This only makes sense if E = FALSE.
type	This argument can take three values: "naive" (the effect is calculated ignoring the presence of observed and unobserved confounders), "univariate" (the effect is obtained from the univariate model which neglects the presence of unobserved confounders) and "joint" (the effect is obtained from the bivariate model which accounts for observed and unobserved confounders).
ind	Binary logical variable. It can be used to calculate the RR for a subset of the data. Note that it does not make sense to use ind when some observations are excluded from the RR calculation (e.g., when using E = FALSE).
n.sim	Number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used when delta = FALSE. It may be increased if more precision is required.
prob.lev	Overall probability of the left and right tails of the RR distribution used for interval calculations.
length.out	Ddesired length of the sequence to be used when calculating the effect that a continuous/discrete treatment has on a binary outcome.
hd.plot	If TRUE then a plot of the histogram and kernel density estimate of the simulated risk ratios is produced. This can only be produced when binary responses are used.

104 S.m

rr.plot For the case of continuous/discrete endogenous variable and binary outcome,

if TRUE then a plot (on the log scale) showing the risk ratios that the binary outcome is equal to 1 for each incremental value of the endogenous variable and

respective intervals is produced.

main Title for the plot. xlab Title for the x axis.

. . Other graphics parameters to pass on to plotting commands. These are used only

when hd.plot = TRUE.

#### **Details**

RR calculates the causal risk ratio of the probabilities of positive outcome under treatment (the binary predictor or treatment assumes value 1) and under control (the binary treatment assumes value 0). Posterior simulation is used to obtain a confidence/credible interval.

RR works also for the case of continuous/discrete endogenous treatment variable.

#### Value

prob.lev Probability level used.

sim.RR It returns a vector containing simulated values of the average RR. This is used

to calculate intervals.

Ratios For the case of continuous/discrete endogenous variable and binary outcome, it

returns a matrix made up of three columns containing the risk ratios for each

incremental value in the endogenous variable and respective intervals.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

## See Also

GJRM-package, gjrm

S.m Internal Function

## **Description**

It provides penalty matrices in a format suitable for automatic multiple smoothing parameter estimation.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

SemiParBIV 105

SemiParBIV

Internal fitting function

#### **Description**

Internal fitting set up function.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

SemiParBIV.fit

Internal Function

# Description

Wrapper of core algorithm.

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

SemiParBIV.fit.post

Internal Function

# **Description**

This and other similar internal functions calculate useful post estimation quantities.

# Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

SemiParROY

Internal fitting function

# Description

Internal fitting set up function.

# Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

SemiParTRIV	Internal fitting function
	Title Titel filling filletter

# Description

Internal fitting set up function.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

```
summary. \verb|copulaSampleSel| \\ copulaSampleSel| summary
```

# Description

It takes a fitted copulaSampleSel object and produces some summaries from it.

## Usage

## **Arguments**

object	A fitted copulaSampleSel object.
X	$summary.copula Sample Sel\ object\ produced\ by\ summary.copula Sample Sel().$
n.sim	The number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used to calculate intervals for the association parameter, dispersion coefficient, for instance It may be increased if more precision is required.
prob.lev	Probability of the left and right tails of the posterior distribution used for interval calculations.
digits	Number of digits printed in output.
signif.stars	By default significance stars are printed alongside output.
	Other arguments.

summary.gamlss 107

## **Details**

```
\verb"print.summary.copulaSampleSel" prints model term summaries.
```

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

# **Examples**

```
## see examples for gjrm
```

summary.gamlss

gamlss summary

# Description

It takes a fitted gamlss object and produces some summaries from it.

# Usage

# Arguments

object	A fitted gamlss object.
X	summary.gamlss object produced by summary.gamlss().
n.sim	The number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used to calculate intervals for various parameters. It may be increased if more precision is required.
prob.lev	Probability of the left and right tails of the posterior distribution used for interval calculations.
digits	Number of digits printed in output.
signif.stars	By default significance stars are printed alongside output.
	Other arguments.

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#### **Details**

print.summary.gamlss prints model term summaries.

#### Value

tableP1 Table containing parametric estimates, their standard errors, z-values and p-

values for equation 1.

tableP2, tableP3

As above but for equations 2 and 3 if present.

tableNP1 Table of nonparametric summaries for each smooth component including effec-

tive degrees of freedom, estimated rank, approximate Wald statistic for testing the null hypothesis that the smooth term is zero and corresponding p-value, for

equation 1.

tableNP2, tableNP3

As above but for equations 2 and 3.

n Sample size.

sigma, nu Estimated distribution specific parameters.

formula1, formula2, formula3

Formulas used for the model equations.

1.sp1,1.sp2,1.sp3

Number of smooth components in model equation.

t.edf Total degrees of freedom of the estimated bivariate model.

CIsig, CInu Intervals for distribution specific parameters.

### Author(s)

Maintainer: Giampiero Marra < giampiero . marra@ucl.ac.uk>

#### **Examples**

## see examples for gamlss

summary.gjrm

gjrm summary

## **Description**

It takes a fitted gjrm object and produces some summaries from it.

summary.gjrm 109

## Usage

## Arguments

object	A fitted gjrm object.
X	summary.gjrm object produced by summary.gjrm().
n.sim	The number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used to calculate intervals for the association parameter, dispersion coefficient etc. It may be increased if more precision is required.
prob.lev	Probability of the left and right tails of the posterior distribution used for interval calculations.
digits	Number of digits printed in output.
signif.stars	By default significance stars are printed alongside output.
	Other arguments.

#### **Details**

print.summary.gjrm prints model term summaries.

#### Value

tableP1 Table containing parametric estimates, their standard errors, z-values and pvalues for equation 1. tableP2,tableP3, ... As above but for equation 2 and equations 3 and 4 if present. tableNP1 Table of nonparametric summaries for each smooth component including effective degrees of freedom, estimated rank, approximate Wald statistic for testing the null hypothesis that the smooth term is zero and corresponding p-value, for equation 1. tableNP2, tableNP3, ... As above but for equation 2 and equations 3 and 4 if present. Sample size. n theta Estimated dependence parameter linking the two equations.

tau Estimated Kendall's tau dependence measure between the two equations.

```
sigma1, sigma2 Estimated distribution specific parameters for equations 1 and 2. nu1, nu2 Estimated distribution specific parameters for equations 1 and 2. formula1, formula2, formula3, ... Formulas used for the model equations.  
1.sp1,1.sp2,1.sp3, ... Number of smooth components in model equations.  
t.edf Total degrees of freedom of the estimated bivariate model.  
CItheta, CItau Interval(s) for \theta and Kendall's tau.  
CIsig1,CIsig2,CInu1,CInu2 Intervals for distribution specific parameters
```

#### WARNINGS

Note that the summary output will also indeed provide the Kendall's tau and related interval. This is a valid measure of dependence for continuous margins but it is not for discrete margins, for instance. However, it is still displayed for the sake of keeping the printed output consistent with that of other models in the package.

## Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

```
summary.SemiParBIV SemiParBIV summary
```

## **Description**

It takes a fitted SemiParBIV object and produces some summaries from it.

## Usage

summary.SemiParBIV 111

#### **Arguments**

object A fitted SemiParBIV object. summary. SemiParBIV object produced by summary. SemiParBIV(). n.sim The number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used to calculate intervals for the association parameter, dispersion coefficient and other measures (e.g., gamma measure). It may be increased if more precision is required. Probability of the left and right tails of the posterior distribution used for interval prob.lev calculations. gm If TRUE then intervals for the gamma measure and odds ratio are calculated. digits Number of digits printed in output. signif.stars By default significance stars are printed alongside output.

#### **Details**

Using some low level functions in mgcv, based on the results of Marra and Wood (2012), 'Bayesian p-values' are returned for the smooth terms. These have better frequentist performance than their frequentist counterpart. See the help file of summary.gam in mgcv for further details. Covariate selection can also be achieved using a single penalty shrinkage approach as shown in Marra and Wood (2011).

Posterior simulation is used to obtain intervals of nonlinear functions of parameters, such as the association and dispersion parameters as well as the odds ratio and gamma measure discussed by Tajar et al. (2001) if gm = TRUE.

print.summary.SemiParBIV prints model term summaries.

Other arguments.

## Value

tableP1 Table containing parametric estimates, their standard errors, z-values and p-values for equation 1.

tableP2,tableP3, ...

As above but for equation 2 and equations 3 and 4 if present.

tableNP1 Table of nonparametric summaries for each smooth component including effec-

tive degrees of freedom, estimated rank, approximate Wald statistic for testing the null hypothesis that the smooth term is zero and corresponding p-value, for

equation 1.

tableNP2,tableNP3, ...

As above but for equation 2 and equations 3 and 4 if present.

n Sample size.

theta Estimated dependence parameter linking the two equations.

formula1, formula2, formula3, ...

Formulas used for the model equations.

1.sp1, 1.sp2, 1.sp3, ...

Number of smooth components in model equations.

t.edf Total degrees of freedom of the estimated bivariate model.

CItheta Interval(s) for  $\theta$ .

n.sel Number of selected observations in the sample selection case.

OR, CIor Odds ratio and related CI. The odds ratio is a measure of association between

binary random variables and is defined as p00p11/p10p01. In the case of independence this ratio is equal to 1. It can take values in the range (-Inf, Inf) and it does not depend on the marginal probabilities (Tajar et al., 2001). Interval is

calculated using posterior simulation.

GM, CIgm Gamma measure and related CI. This measure of association was proposed by

Goodman and Kruskal (1954). It is defined as (OR - 1)/(OR + 1), can take values in the range (-1, 1) and does not depend on the marginal probabilities. Interval

is calculated using posterior simulation.

tau, CItau Kendall's tau and respective intervals.

#### **WARNINGS**

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Note that the summary output will also indeed provide the Kendall's tau and related interval. This is a valid measure of dependence for continuous margins but it is not for binary margins. However, it is still displayed for the sake of keeping the printed output consistent with that of other models in the package.

#### Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

#### References

Marra G. and Wood S.N. (2011), Practical Variable Selection for Generalized Additive Models. *Computational Statistics and Data Analysis*, 55(7), 2372-2387.

Marra G. and Wood S.N. (2012), Coverage Properties of Confidence Intervals for Generalized Additive Model Components. *Scandinavian Journal of Statistics*, 39(1), 53-74.

Tajar M., Denuit M. and Lambert P. (2001), Copula-Type Representation for Random Couples with Bernoulli Margins. Discussion Papaer 0118, Universite Catholique De Louvain.

# See Also

AT, prev

summary.SemiParROY SemiParROY summary

# Description

It takes a fitted SemiParROY object and produces some summaries from it.

# Usage

## **Arguments**

object	A fitted SemiParROY object.
x	summary.SemiParROY object produced by summary.SemiParROY().
n.sim	The number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used to calculate intervals for the association parameter, dispersion coefficient, for instance It may be increased if more precision is required.
prob.lev	Probability of the left and right tails of the posterior distribution used for interval calculations.
digits	Number of digits printed in output.
signif.stars	By default significance stars are printed alongside output.
	Other arguments.

## **Details**

print.summary.SemiParROY prints model term summaries.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

## **Examples**

```
## see examples for gjrm
```

summary.SemiParTRIV SemiParTRIV summary

# Description

It takes a fitted SemiParTRIV object and produces some summaries from it.

TRIapprox

## Usage

## **Arguments**

object	A fitted SemiParTRIV object.
Х	summary.SemiParTRIV object produced by summary.SemiParTRIV().
n.sim	The number of simulated coefficient vectors from the posterior distribution of the estimated model parameters. This is used to calculate intervals for the asso- ciation parameter and other measures. It may be increased if more precision is required.
prob.lev	Probability of the left and right tails of the posterior distribution used for interval calculations.
digits	Number of digits printed in output.
signif.stars	By default significance stars are printed alongside output.
	Other arguments.

# **Details**

print.summary.SemiParTRIV prints model term summaries.

# Author(s)

Maintainer: Giampiero Marra < giampiero . marra@ucl.ac.uk>

## **Examples**

```
## see examples for gjrm
```

TRIapprox Internal Function

# Description

It approximates the trivariate normal integral.

## Author(s)

Maintainer: Giampiero Marra <giampiero.marra@ucl.ac.uk>

triprobgHs 115

## **Description**

It provides score and Hessian for trivariate binary models.

## Author(s)

Author: Panagiota Filippou

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

vis.gjrm

Visualization function

# **Description**

It takes a fitted gjrm object produced by gjrm() and produces perspective or contour plot views of model predictions. This function is a wrapper of vis.gam() in mgcv. Please see the documentation of vis.gam() for full details.

## Usage

```
vis.gjrm(x, eq, fun = NULL, ...)
```

# Arguments

x A fitted gjrm object.

eq The equation number.

fun Either mean or variance. If left as equal to NULL then predictions on the scale

of the predictor will be produced.

Other graphics parameters to pass on to plotting commands, as described for

vis.gam() in mgcv.

## Value

The function generates plots.

## Author(s)

Maintainer: Giampiero Marra < giampiero.marra@ucl.ac.uk>

# See Also

gjrm

VuongClarke VuongClarke

VuongClarke

Vuong and Clarke tests

## **Description**

The Vuong and Clarke tests are likelihood-ratio-based tests that can be used for choosing between two non-nested models.

## Usage

```
VuongClarke(obj1, obj2, sig.lev = 0.05)
```

## **Arguments**

obj1, obj2 Objects of the two fitted bivariate non-nested models.

sig.lev Significance level used for testing.

#### **Details**

The Vuong (1989) and Clarke (2007) tests are likelihood-ratio-based tests for model selection that use the Kullback-Leibler information criterion. The implemented tests can be used for choosing between two bivariate models which are non-nested.

In the Vuong test, the null hypothesis is that the two models are equally close to the actual model, whereas the alternative is that one model is closer. The test follows asymptotically a standard normal distribution under the null. Assume that the critical region is (-c,c), where c is typically set to 1.96. If the value of the test is higher than c then we reject the null hypothesis that the models are equivalent in favor of model obj1. Viceversa if the value is smaller than c. If the value falls in [-c,c] then we cannot discriminate between the two competing models given the data.

In the Clarke test, if the two models are statistically equivalent then the log-likelihood ratios of the observations should be evenly distributed around zero and around half of the ratios should be larger than zero. The test follows asymptotically a binomial distribution with parameters n and 0.5. Critical values can be obtained as shown in Clarke (2007). Intuitively, model obj1 is preferred over obj2 if the value of the test is significantly larger than its expected value under the null hypothesis (n/2), and vice versa. If the value is not significantly different from n/2 then obj1 can be thought of as equivalent to obj2.

## Value

It returns two decisions based on the tests and criteria discussed above.

## Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

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#### References

Clarke K. (2007), A Simple Distribution-Free Test for Non-Nested Model Selection. *Political Analysis*, 15, 347-363.

Vuong Q.H. (1989), Likelihood Ratio Tests for Model Selection and Non-Nested Hypotheses. *Econometrica*, 57(2), 307-333.

#### **Examples**

## see examples for gjrm

war

Civil war data

# Description

Civil war data from Fearon and Laitin (2003).

## Usage

data(war)

#### **Format**

war is a 6326 row data frame with the following columns

**onset** equal to 1 for all country-years in which a civil war started.

**instab** equal to 1 if unstable government.

oil equal to 1 for oil exporter country.

warl equal to 1 if the country had a distinct civil war ongoing in the previous year.

gdpenl GDP per capita (measured as thousands of 1985 U.S. dollars) lagged one year.

**ncontig** equal to 1 for non-contiguous state.

nwstate equal to 1 for new state.

**lpopl** log(population size).

**Imtnest** log(mountainous).

**ethfrac** measure of ethnic fractionalization (calculated as the probability that two randomly drawn individuals from a country are not from the same ethnicity).

relfrac measure of religious fractionalization.

polity21 measure of political democracy (ranges from -10 to 10) lagged one year.

## Source

Data are from:

Fearon J.D., Laitin D.D. (2003), Ethnicity, Insurgency, and Civil War. *The American Political Science Review*, 97, 75-90.

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## **Examples**

```
## Not run:
library("GJRM")
data("war", package = "GJRM")
# Bivariate brobit model with partial observability
reb.eq <- onset ~ instab + oil + warl + lpopl + lmtnest + ethfrac +
               polity21 + s(gdpenl) + s(relfrac)
gov.eq <- onset ~ instab + oil + warl + ncontig + nwstate + s(gdpenl)</pre>
bpo <- gjrm(list(reb.eq, gov.eq), data = war, Model = "BPO",</pre>
          margins = c("probit", "probit"))
conv.check(bpo)
# perhaps model is to complex
set.seed(1)
sbpo <- summary(bpo)</pre>
sbpo$theta; sbpo$CItheta
# let's exclude the correlation parameter in fitting
bpo0 <- gjrm(list(reb.eq, gov.eq), data = war, Model = "BPO0",</pre>
           margins = c("probit", "probit"))
conv.check(bpo0)
summary(bpo0)
war.eq <- onset ~ instab + oil + warl + ncontig + nwstate + lpopl +</pre>
               lmtnest + ethfrac + polity2l + s(gdpenl) + s(relfrac)
Probit <- gam(war.eq, family = binomial(link = "probit"), data = war)</pre>
summary(Probit)
coef(Probit)[(which(names(coef(Probit)) == "s(gdpen1).9"))]
coef(bpo0)[(which(names(coef(bpo)) == "s(gdpen1).9"))]
probitW <- bpoW <- bpoReb <- bpoGov <- NA</pre>
gdp.grid <- seq(0, 8)</pre>
```

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```
median.values <- data.frame(t(apply(war, 2, FUN = median)))</pre>
for (i in 1:length(gdp.grid)){
newd <- median.values; newd$gdpenl <- gdp.grid[i]</pre>
eta1 <- predict(bpo0, eq = 1, newd)
eta2 <- predict(bpo0, eq = 2, newd)
probitW[i] <- predict(Probit, newd, type = "response")</pre>
bpoW[i]
          <- pnorm(eta1)*pnorm(eta2)</pre>
bpoReb[i] <- pnorm(eta1)</pre>
bpoGov[i] <- pnorm(eta2)</pre>
}
plot(gdp.grid, probitW, type = "1", ylim = c(0, 0.55), lwd = 2,
     col = "grey", xlab = "GDP per Capita (in thousands)",
     ylab = "Pr(Outcome)", main = "Probabilities for All Outcomes",
     cex.main = 1.5, cex.lab = 1.3, cex.axis = 1.3)
lines(gdp.grid, bpoW, lwd = 2)
lines(gdp.grid, bpoReb, lwd = 2, lty = 2)
lines(gdp.grid, bpoGov, lwd = 2, lty = 3)
#dev.copy(postscript, "probWAR.eps", width = 8)
#dev.off()
## End(Not run)
#
```

working.comp

Internal Function

# **Description**

It efficiently calculates the working model quantities needed to implement the automatic multiple smoothing parameter estimation procedure by exploiting a result which leads to very fast and stable calculations.

#### Author(s)

Maintainer: Giampiero Marra < giampiero .marra@ucl.ac.uk>

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