Package 'dnet'

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Type Package

Title Integrative Analysis of Omics Data in Terms of Network, Evolution and Ontology

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Depends R (>= 3.1.0), igraph, supraHex

Imports graph, Rgraphviz, Matrix

Suggests limma, survival, foreach, doParallel, Biobase

Description The focus of the dnet by Fang and Gough (2014) <doi:10.1186/s13073-014-0064-8> is to make sense of omics data (such as gene expression and mutations) from different angles including: integration with molecular networks, enrichments using ontologies, and relevance to gene evolutionary ages. Integration is achieved to identify a gene subnetwork from the whole gene network whose nodes/genes are labelled with informative data (such as the significant levels of differential expression or survival risks). To help make sense of identified gene networks, enrichment analysis is also supported using a wide variety of pre-compiled ontologies and phylostratific gene age information in major organisms including: human, mouse, rat, chicken, C.elegans, fruit fly, zebrafish and arabidopsis. Add-on functionalities are supports for calculating semantic similarity between ontology terms (and between genes) and for calculating network affinity based on random walk; both can be done via high-performance parallel computing.

URL http://dnet.r-forge.r-project.org, https://github.com/hfang-bristol/dnet

Collate 'dGSEA.r' 'dGSEAview.r' 'dGSEAwrite.r' 'visGSEA.r' 'dPvalAggregate.r' 'dNetInduce.r' 'dBUMfit.r' 'dBUMscore.r' 'dNetFind.r' 'dNetPipeline.r' 'dNetConfidence.r' 'visNet.r' 'visNetMul.r' 'visNetAnimate.r' 'visNetReorder.r' 'dNetReorder.r' 'visNetArc.r' 'visNetCircle.r' 'dRWR.r' 'dRWRcontact.r' 'dRWRpipeline.r' 'dContrast.r' 'dCommSignif.r' 'dSVDsignif.r' 'dFDRscore.r' 'dDAGinduce.r' 'dDAGreverse.r' 'dDAGroot.r' 'dDAGtip.r' 'dDAGlevel.r' 'dDAGannotate.r'

'dDAGancestor.r' 'dDAGtermSim.r' 'dDAGgeneSim.r' 'visDAG.r'
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'dRDataLoader.r' 'dCheckParallel.r' 'dFunArgs.r'

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R topics documented:

`dBUMfit` *Function to fit a p-value distribution under beta-uniform mixture model*

Description

`dBUMfit` is supposed to take as input a vector of p-values for deriving their distribution under beta-uniform mixture model (see Note below). The density distribution of input p-values is expressed as a mixture of two components: one for the null hypothesis (the noise component) and the other for the alternative hypothesis (the signal component). The noise component is the uniform density, while the signal component is the remainder of the mixture distribution. It returns an object of class "BUM".

Usage

```
dBUMfit(x, ntry = 1, hist.bum = T, contour.bum = T, verbose = T)
```

Arguments

`x`	a vector containing input p-values
`ntry`	an integeter specifying how many trys are used to find the optimised parameters by maximum likelihood estimation
`hist.bum`	logical to indicate whether the histogram graph should be drawn
`contour.bum`	logical to indicate whether a contour plot should be drawn to show the log likelihood as a function of two parameters (a and lambda) in the beta-uniform mixture model
`verbose`	logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

an object of class "BUM", a list with following elements:

- `lambda`: estimated mixture parameter
- `a`: estimated shape parameter
- `NLL`: Negative log-likelihood
- `pvalues`: the input pvalues
- `call`: the call that produced this result

Note

The probability density function of p-values under the Beta-Uniform Mixture model is formulated as: $f(x|\lambda, a) = \lambda + (1-\lambda) * a * x^{a-1}$. The model names after mixing two distributions:

- the uniform distribution with the density function as $\frac{1}{b-a}|_{a=0}^{b=1} = 1$
- the beta distribution with the density function as $\frac{\Gamma(a+b)}{\Gamma(a)+\Gamma(b)} * x^{a-1} * (1-x)^{b-1}|_{b=1} = a * x^{a-1}$

Both are mixed via λ. The mixture parameter λ measures the contribution from the uniform distribution. Accordingly, $1-\lambda$ measures the contribution from the beta distribution. Notably, the probability density function of the beta distribution can be splitted into two parts (rather than the exclusitive signal):

- the constant part as noise: $a * x^{a-1}|_{x=1} = a$
- the rest part as signal: $a * (x^{a-1} - 1)$

In other words, there is no signal at $x = 1$ but all being noise. It is a conservative, upper bound estimation of the noise. Therefore, the probability density function in the model can be decomposed into signal-noise components:

- the signal component: $(1-\lambda) * a * (x^{a-1} - 1)$
- the noise component: $\lambda + (1-\lambda) * a$

It is misleading to simply view λ as the noise component and $(1-\lambda) * a * x^{a-1}$ as the signal component, just as wrongly do in the literatures (e.g. http://www.ncbi.nlm.nih.gov/pubmed/18586718)

See Also

dBUMscore

Examples

```
# 1) generate an vector consisting of random values from beta distribution
x <- rbeta(1000, shape1=0.5, shape2=1)

# 2) fit a p-value distribution under beta-uniform mixture model
fit <- dBUMfit(x)
fit$lambda
fit$a
```

---

dBUMscore *Function to transform p-values into scores according to the fitted beta-uniform mixture model and/or after controlling false discovery rate*

Description

dBUMscore is supposed to take as input a vector of p-values, which are transformed into scores according to the fitted beta-uniform mixture model. Also if the FDR threshold is given, it is used to make sure that p-values below this are considered significant and thus scored positively. Instead, those p-values above the given FDR are considered insigificant and thus scored negatively.

Usage

```
dBUMscore(fit, method = c("pdf", "cdf"), fdr = NULL, scatter.bum = T)
```

Arguments

`fit`	an object of class "BUM"
`method`	the method used for the transformation. It can be either "pdf" for the method based on the probability density function of the fitted model, or "cdf" for the method based on the cumulative distribution function of the fitted model
`fdr`	the given FDR threshold. By default, it is set to NULL, meaning there is no constraint. If given, those p-values with the FDR below this are considered significant and thus scored positively. Instead, those p-values with the FDR above this given FDR are considered insigificant and thus scored negatively
`scatter.bum`	logical to indicate whether the scatter graph of scores against p-values should be drawn. Also indicated is the p-value (called tau) corresponding to the given FDR threshold (if any)

Value

- `scores`: a vector of scores

Note

The transformation from the input p-value x to the score $S(x)$ is based on the fitted beta-uniform mixture model with two parameters λ and a: $f(x|\lambda, a) = \lambda + (1-\lambda) * a * x^{a-1}$. Specifically, it considers the log-likelyhood ratio between the signal and noise compoment of the model. The probability density function (pdf) of the signal component and the noise component are $(1-\lambda) * a * (x^{a-1} - 1)$ and $\lambda + (1-\lambda) * a$, respectively. Accordingly, the cumulative distribution function (cdf) of the signal component and the noise component are $\int_0^x (1-\lambda) * a * (x^{a-1} - 1)\,\mathrm{d}x$ and $\int_0^x \lambda + (1-\lambda) * a\,\mathrm{d}x$. In order to take into account the significance of the p-value, the fdr threshold is also used for down-weighting the score. According to how to measure both components, there are two methods implemented for deriving the score $S(x)$:

- The method "pdf": $S(x) = log_2 \frac{(1-\lambda)*a*(x^{a-1}-1)}{\lambda+(1-\lambda)*a} - log_2 \frac{(1-\lambda)*a*(\tau^{a-1}-1)}{\lambda+(1-\lambda)*a} = log_2 \big(\frac{x^{a-1}-1}{\tau^{a-1}-1}\big)$. For the purpose of down-weighting scores, it must ensure $log_2 \frac{(1-\lambda)*a*(\tau^{a-1}-1)}{\lambda+(1-\lambda)*a} \geq 0$, that is, the constraint via $\tau \leq \big(\frac{\lambda+2*a*(1-\lambda)}{a*(1-\lambda)}\big)^{\frac{1}{a-1}}$
- The method "cdf": $S(x) = log_2 \frac{\int_0^x (1-\lambda)*a*(x^{a-1}-1)\,\mathrm{d}x}{\int_0^x \lambda+(1-\lambda)*a\,\mathrm{d}x} - log_2 \frac{\int_0^\tau (1-\lambda)*a*(\tau^{a-1}-1)\,\mathrm{d}x}{\int_0^\tau \lambda+(1-\lambda)*a\,\mathrm{d}x} = log_2 \frac{(1-\lambda)*(x^{a-1}-a)}{\lambda+(1-\lambda)*a} - log_2 \frac{(1-\lambda)*(\tau^{a-1}-a)}{\lambda+(1-\lambda)*a} = log_2 \big(\frac{x^{a-1}-a}{\tau^{a-1}-a}\big)$. For the purpose of down-weighting scores, it must ensure $log_2 \frac{(1-\lambda)*(\tau^{a-1}-a)}{\lambda+(1-\lambda)*a} \geq 0$, that is, the constraint via $\tau \leq \big(\frac{\lambda+2*a*(1-\lambda)}{1-\lambda}\big)^{\frac{1}{a-1}}$
- Where $\tau = \big[\frac{\lambda+(1-\lambda)*a-fdr*\lambda}{fdr*(1-\lambda)}\big]^{\frac{1}{a-1}}$, i.e. the p-value corresponding to the exact fdr threshold. It can be deduced from the definition of the false discovery rate: $fdr \doteq \frac{\int_0^\tau \lambda+(1-\lambda)*a\,\mathrm{d}x}{\int_0^\tau \lambda+(1-\lambda)*a*x^{a-1}\,\mathrm{d}x}$. Notably, if the calculated τ exceeds the contraint, it will be reset to the maximum end of that constraint

See Also

dBUMfit

Examples

```
# 1) generate an vector consisting of random values from beta distribution
x <- rbeta(1000, shape1=0.5, shape2=1)

# 2) fit a p-value distribution under beta-uniform mixture model
fit <- dBUMfit(x)

# 3) calculate the scores according to the fitted BUM and fdr=0.01
# using "pdf" method
scores <- dBUMscore(fit, method="pdf", fdr=0.01)
# using "cdf" method
scores <- dBUMscore(fit, method="cdf", fdr=0.01)
```

---

dCheckParallel *Function to check whether parallel computing should be used and how*

Description

dCheckParallel is used to check whether parallel computing should be used and how

Usage

```
dCheckParallel(multicores = NULL, verbose = T)
```

Arguments

multicores	an integer to specify how many cores will be registered as the multicore parallel backend to the 'foreach' package. If NULL, it will use a half of cores available in a user's computer
verbose	logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

TRUE for using parallel computing; FALSE otherwise

Note

Whether parallel computation with multicores is used is system-specific. Also, it will depend on whether these two packages "foreach" and "doParallel" have been installed. It can be installed via: source("http://bioconductor.org/biocLite.R"); biocLite(c("foreach","doParallel")).

See Also

dRWR, dRWRcontact, dRWRpipeline, dDAGtermSim, dDAGgeneSim

Examples

```
#dCheckParallel(multicores=2)
```

---

dCommSignif *Function to test the significance of communities within a graph*

Description

dCommSignif is supposed to test the significance of communities within a graph. For a community of the graph, it first calculates two types of degrees for each node: degrees based on parters only within the community itself, and the degrees based on its parters NOT in the community but in the graph. Then, it performs two-sample Wilcoxon tests on these two types of degrees to produce the signficance level (p-value)

Usage

```
dCommSignif(g, comm)
```

Arguments

g	an object of class "igraph" or "graphNEL"
comm	an object of class "communities". Details on this class can be found at http://igraph.org/r/doc/communities.html

Value

- significance: a vector of p-values (significance)

Note

none

See Also

dCommSignif

Examples

```
# 1) generate an vector consisting of random values from beta distribution
x <- rbeta(1000, shape1=0.5, shape2=1)

# 2) fit a p-value distribution under beta-uniform mixture model
fit <- dBUMfit(x, ntry=1, hist.bum=FALSE, contour.bum=FALSE)

# 3) calculate the scores according to the fitted BUM and fdr=0.01
# using "pdf" method
scores <- dBUMscore(fit, method="pdf", fdr=0.05, scatter.bum=FALSE)
names(scores) <- as.character(1:length(scores))
```

```
# 4) generate a random graph according to the ER model
g <- erdos.renyi.game(1000, 1/100)

# 5) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)

# 6) find the module with the maximum score
module <- dNetFind(subg, scores)

# 7) find the module and test its signficance
comm <- walktrap.community(module, modularity=TRUE)
significance <- dCommSignif(module, comm)
```

---

dContrast *Function to help build the contrast matrix*

Description

dContrast is used to help build the contrast matrix

Usage

```
dContrast(
level_sorted,
contrast.type = c("average", "zero", "sequential", "pairwise")
)
```

Arguments

level_sorted	a vector of levels (usually sorted) which are contrated to each other
contrast.type	the type of the contrast. It can be one of either 'average' for the contrast against the average of all levels, 'zero' for the contrast against the zero, 'sequential' for the contrast in a sequential order (it requires the levels being sorted properly), or 'pairwise' for the pairwise contrast.

Value

a list with following components:

- each: the contrast being specified
- name: the name of the contrast

Note

none

Examples

```
level_sorted <- c("L1","L2","L3","L4")

# the contrast against the average of all levels
contrasts <- dContrast(level_sorted, contrast.type="average")

# the contrast against the zero
contrasts <- dContrast(level_sorted, contrast.type="zero")

# the contrast in a sequential order
contrasts <- dContrast(level_sorted, contrast.type="sequential")

# the pairwise contrast
contrasts <- dContrast(level_sorted, contrast.type="pairwise")
```

`dDAGancestor` *Function to find common ancestors of two terms/nodes from a direct acyclic graph (DAG)*

Description

`dDAGancestor` is supposed to find a list of common ancestors shared by two terms/nodes, given a direct acyclic graph (DAG; an ontology). If two terms are given as NULL, then a sparse matrix of children x ancestors is built for all terms. If one of them is null, then a sparse matrix of children x ancestors is built but only for non-null input terms.

Usage

```
dDAGancestor(g, term1 = NULL, term2 = NULL, verbose = T)
```

Arguments

`g`	an object of class "igraph" or "graphNEL"
`term1`	the first term/node as input
`term2`	the second term/node as input
`verbose`	logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

- When two terms are given: a list of terms/nodes that are common ancestors for two input terms/nodes
- When two terms are given as NULL: a sparse matrix of children x ancestors is built for all terms, with '1' for the reachable and otherwise '0'.
- When one of terms is given as NULL: a sparse matrix of children x ancestors is built but only for non-null input terms, with '1' for the reachable and otherwise '0'.

Note

none

See Also

dDAGinduce

Examples

```
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) randomly give two terms
term1 <- sample(V(g)$name,1)
term2 <- sample(V(g)$name,1)

# 3) find common ancestors
dDAGancestor(g, term1, term2)
```

---

dDAGannotate *Function to generate a subgraph of a direct acyclic graph (DAG) induced by the input annotation data*

Description

dDAGannotate is supposed to produce a subgraph induced by the input annotation data, given a direct acyclic graph (DAG; an ontology). The input is a graph of "igraph" or "graphNET" object, a list of the vertices containing annotation data, and the mode defining the paths to the root of DAG. The induced subgraph contains vertices (with annotation data) and their ancestors along with the defined paths to the root of DAG. The annotations at these vertices (including their ancestors) are also updated according to the true-path rule: a gene annotated to a term should also be annotated by its all ancestor terms.

Usage

```
dDAGannotate(
g,
annotations,
path.mode = c("all_paths", "shortest_paths", "all_shortest_paths"),
verbose = TRUE
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
annotations	the vertices/nodes for which annotation data are provided
path.mode	the mode of paths induced by vertices/nodes with input annotation data. It can be "all_paths" for all possible paths to the root, "shortest_paths" for only one path to the root (for each node in query), "all_shortest_paths" for all shortest paths to the root (i.e. for each node, find all shortest paths with the equal lengths)
verbose	logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

- subg: an induced subgraph, an object of class "igraph". In addition to the original attributes to nodes and edges, the return subgraph is also appended by new node attributes: "annotations", which contains a list of genes either as original annotations or inherited annotations; "IC", which stands for information content defined as negative 10-based log-transformed frequency of genes annotated to that term.

Note

For the mode "shortest_paths", the induced subgraph is the most concise, and thus informative for visualisation when there are many nodes in query, while the mode "all_paths" results in the complete subgraph.

See Also

dDAGinduce, dDAGlevel

Examples

```
## Not run:
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) load human genes annotated by HPPA
org.Hs.egHPPA <- dRDataLoader(RData='org.Hs.egHPPA')
GS <- org.Hs.egHPPA # as 'GS' object

# 3) prepare for annotation data
# randomly select vertices with annotation data
annotations <- GS$gs[sample(1:length(GS$gs),5)]

# 4) obtain the induced subgraph
# 4a) based on all possible paths (i.e. the complete subgraph induced)
dDAGannotate(g, annotations, path.mode="all_paths", verbose=TRUE)
# 4b) based on shortest paths (i.e. the most concise subgraph induced)
dag <- dDAGannotate(g, annotations, path.mode="shortest_paths",
verbose=TRUE)
```

```
# 5) color-code nodes/terms according to the number of annotations
data <- sapply(V(dag)$annotations, length)
names(data) <- V(dag)$name
visDAG(g=dag, data=data, node.info="both")

## End(Not run)
```

---

dDAGgeneSim *Function to calculate pair-wise semantic similarity between genes based on a direct acyclic graph (DAG) with annotated data*

Description

dDAGgeneSim is supposed to calculate pair-wise semantic similarity between genes based on a direct acyclic graph (DAG) with annotated data. It first calculates semantic similarity between terms and then derives semantic similarity between genes from terms-term semantic similarity. Parallel computing is also supported for Linux or Mac operating systems.

Usage

```
dDAGgeneSim(
g,
genes = NULL,
method.gene = c("BM.average", "BM.max", "BM.complete", "average",
"max"),
method.term = c("Resnik", "Lin", "Schlicker", "Jiang", "Pesquita"),
force = TRUE,
fast = TRUE,
parallel = TRUE,
multicores = NULL,
verbose = TRUE
)
```

Arguments

g	an object of class "igraph" or "graphNEL". It must contain a vertex attribute called 'annotations' for storing annotation data (see example for howto)
genes	the genes between which pair-wise semantic similarity is calculated. If NULL, all genes annotatable in the input dag will be used for calculation, which is very prohibitively expensive!
method.gene	the method used for how to derive semantic similarity between genes from semantic similarity between terms. It can be "average" for average similarity between any two terms (one from gene 1, the other from gene 2), "max" for the maximum similarity between any two terms, "BM.average" for best-matching (BM) based average similarity (i.e. for each term of either gene, first calculate maximum similarity to any term in the other gene, then take average of maximum similarity; the final BM-based average similiary is the pre-calculated

average between two genes in pair), "BM.max" for BM based maximum similarity (i.e. the same as "BM.average", but the final BM-based maximum similiary is the maximum of the pre-calculated average between two genes in pair), "BM.complete" for BM-based complete-linkage similarity (inspired by complete-linkage concept: the least of any maximum similarity between a term of one gene and a term of the other gene). When comparing BM-based similarity between genes, "BM.average" and "BM.max" are sensitive to the number of terms invovled; instead, "BM.complete" is much robust in this aspect. By default, it uses "BM.average".

`method.term` the method used to measure semantic similarity between terms. It can be "Resnik" for information content (IC) of most informative common ancestor (MICA) (see http://arxiv.org/pdf/cmp-lg/9511007.pdf), "Lin" for 2*IC at MICA divided by the sum of IC at pairs of terms (see https://www.cse.iitb.ac.in/~cs626-449/Papers/WordSimilarity/3.pdf), "Schlicker" for weighted version of 'Lin' by the 1-prob(MICA) (see http://www.ncbi.nlm.nih.gov/pubmed/16776819), "Jiang" for 1 - difference between the sum of IC at pairs of terms and 2*IC at MICA (see http://arxiv.org/pdf/cmp-lg/9709008.pdf), "Pesquita" for graph information content similarity related to Tanimoto-Jacard index (ie. summed information content of common ancestors divided by summed information content of all ancestors of term1 and term2 (see http://www.ncbi.nlm.nih.gov/pubmed/18460186))

`force` logical to indicate whether the only most specific terms (for each gene) will be used. By default, it sets to true. It is always advisable to use this since it is computationally fast but without compromising accuracy (considering the fact that true-path-rule has been applied when running `dDAGannotate`)

`fast` logical to indicate whether a vectorised fast computation is used. By default, it sets to true. It is always advisable to use this vectorised fast computation; since the conventional computation is just used for understanding scripts

`parallel` logical to indicate whether parallel computation with multicores is used. By default, it sets to true, but not necessarily does so. It will depend on whether these two packages "foreach" and "doParallel" have been installed. It can be installed via: `source("http://bioconductor.org/biocLite.R"); biocLite(c("foreach","doParallel"))` If not yet installed, this option will be disabled

`multicores` an integer to specify how many cores will be registered as the multicore parallel backend to the 'foreach' package. If NULL, it will use a half of cores available in a user's computer. This option only works when parallel computation is enabled

`verbose` logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

It returns a sparse matrix containing pair-wise semantic similarity between input genes. This sparse matrix can be converted to the full matrix via the function `as.matrix`

Note

For the mode "shortest_paths", the induced subgraph is the most concise, and thus informative for visualisation when there are many nodes in query, while the mode "all_paths" results in the complete

subgraph.

See Also

dDAGtermSim, dDAGinduce, dDAGtip, dCheckParallel

Examples

```
## Not run:
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) load human genes annotated by HPPA
org.Hs.egHPPA <- dRDataLoader(RData='org.Hs.egHPPA')

# 3) prepare for ontology and its annotation information
dag <- dDAGannotate(g, annotations=org.Hs.egHPPA,
path.mode="all_paths", verbose=TRUE)

# 4) calculate pair-wise semantic similarity between 5 randomly chosen genes
allgenes <- unique(unlist(V(dag)$annotations))
genes <- sample(allgenes,5)
sim <- dDAGgeneSim(g=dag, genes=genes, method.gene="BM.average",
method.term="Resnik", parallel=FALSE, verbose=TRUE)
sim

## End(Not run)
```

---

dDAGinduce *Function to generate a subgraph of a direct acyclic graph (DAG) induced by given vertices*

Description

dDAGinduce is supposed to produce a subgraph induced by given vertices, given a direct acyclic graph (DAG; an ontology). The input is a graph of "igraph" or "graphNET" object, a list of the vertices of the graph, and the mode defining the paths to the root of DAG. The resultant subgraph inherits the class from the input one. The induced subgraph contains exactly the vertices of interest and their defined paths to the root of DAG.

Usage

```
dDAGinduce(
g,
nodes_query,
path.mode = c("all_paths", "shortest_paths", "all_shortest_paths")
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
nodes_query	the vertices for which the calculation is performed
path.mode	the mode of paths induced by nodes in query. It can be "all_paths" for all possible paths to the root, "shortest_paths" for only one path to the root (for each node in query), "all_shortest_paths" for all shortest paths to the root (i.e. for each node, find all shortest paths with the equal lengths)

Value

- subg: an induced subgraph, an object of class "igraph" or "graphNEL"

Note

For the mode "shortest_paths", the induced subgraph is the most concise, and thus informative for visualisation when there are many nodes in query, while the mode "all_paths" results in the complete subgraph.

See Also

dDAGroot

Examples

```
## Not run:
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) randomly select vertices as the query nodes
# the query nodes can be igraph vertex sequences
nodes_query <- sample(V(g),5)
# more commonly, the query nodes can be term id
nodes_query <- sample(V(g),5)$name

# 3) obtain the induced subgraph
# 3a) based on all possible paths (i.e. the complete subgraph induced)
subg <- dDAGinduce(g, nodes_query, path.mode="all_paths")
# 3b) based on shortest paths (i.e. the most concise subgraph induced)
subg <- dDAGinduce(g, nodes_query, path.mode="shortest_paths")

## End(Not run)
```

dDAGlevel *Function to define/calculate the level of nodes in a direct acyclic graph (DAG)*

Description

dDAGlevel is supposed to calculate the level of nodes, given a direct acyclic graph (DAG; an ontology). The input is a graph of "igraph" or "graphNET" object, and the definition of the node level. The return can be the level for each node or the nodes for each level.

Usage

```
dDAGlevel(
g,
level.mode = c("longest_path", "shortest_path"),
return.mode = c("node2level", "level2node")
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
level.mode	the mode of how to define the level of nodes in DAG. It can be "longest_path" for defining the node level as the length of the longest path from the node to the root, and "shortest_paths" for defining the node level as the length of the shortest path from the node to the root
return.mode	the mode of how to return the node level information. It can be "node2level" for returning a named vector (i.e. the level for each node), and "level2node" for returning a named list (i.e. nodes for each level)

Value

When "return.mode" is "node2level", it returns a named vector: for each named node (i.e. Term ID), it stores its level When "return.mode" is "level2node", it returns a named list: for each named level, it contains the names (i.e. Term ID) of nodes belonging to this level

Note

The level for the root is 1. The level based on the longest path will ensure that nodes at the same level will never be reachable (i.e. in the same path), while the level based on the shortest path will not be necessary. The "longest path" based level can be useful in visiting nodes from the tipmost level to the root: 1) for the current node, all children have been visited; 2) nodes at the same level can be looked at independantly. The "shortest path" based level can be useful in deriving nodes according to their closeness to the root.

See Also

dDAGroot, dDAGreverse

Examples

```
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) randomly select vertices as the query nodes
nodes_query <- sample(V(g),5)$name

# 3) obtain the complete subgraph induced
subg <- dDAGinduce(g, nodes_query)

# 4) calculate the node levels
# 4a) definition based on the longest path
dDAGlevel(subg, level.mode="longest_path")
# 4b) definition based on the shortest path
dDAGlevel(subg, level.mode="shortest_path")
# 4c) definition based on the longest path, and return nodes for each level
dDAGlevel(subg, level.mode="longest_path", return.mode="level2node")
```

---

dDAGreverse *Function to reverse the edge direction of a direct acyclic graph (DAG)*

Description

dDAGreverse is supposed to reverse the edge direction of a direct acyclic graph (DAG; an ontology). The return graph remains all attributes associated on nodes and edges.

Usage

```
dDAGreverse(g)
```

Arguments

g	an object of class "igraph" or "graphNEL"

Value

- gr: a graph being reversed, an object of class "igraph" or "graphNEL"

Note

none

See Also

dDAGreverse

Examples

```
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) the graph with reverse edge direction
gr <- dDAGreverse(g)
gr
```

---

dDAGroot *Function to find the root node of a direct acyclic graph (DAG)*

Description

dDAGroot is supposed to find the root node of a direct acyclic graph (DAG; an ontology). It return the name (i.e Term ID) of the root node.

Usage

```
dDAGroot(g)
```

Arguments

g	an object of class "igraph" or "graphNEL"

Value

- root: the root name (i.e. Term ID)

Note

none

See Also

dDAGroot

Examples

```
## Not run:
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) find the root
root <- dDAGroot(g)
root

## End(Not run)
```

dDAGtermSim *Function to calculate pair-wise semantic similarity between input terms based on a direct acyclic graph (DAG) with annotated data*

Description

dDAGtermSim is supposed to calculate pair-wise semantic similarity between input terms based on a direct acyclic graph (DAG) with annotated data. Parallel computing is also supported for Linux or Mac operating systems.

Usage

```
dDAGtermSim(
g,
terms = NULL,
method = c("Resnik", "Lin", "Schlicker", "Jiang", "Pesquita"),
fast = T,
parallel = TRUE,
multicores = NULL,
verbose = T
)
```

Arguments

g	an object of class "igraph" or "graphNEL". It must contain a vertex attribute called 'annotations' for storing annotation data (see example for howto)
terms	the terms/nodes between which pair-wise semantic similarity is calculated. If NULL, all terms in the input DAG will be used for calcluation, which is very prohibitively expensive!
method	the method used to measure semantic similarity between input terms. It can be "Resnik" for information content (IC) of most informative common ancestor (MICA) (see http://arxiv.org/pdf/cmp-lg/9511007.pdf), "Lin" for 2*IC at MICA divided by the sum of IC at pairs of terms (see https://www.cse.iitb.ac.in/~cs626-449/Papers/WordSimilarity/3.pdf), "Schlicker" for weighted version of 'Lin' by the 1-prob(MICA) (see http://www.ncbi.nlm.nih.gov/pubmed/16776819), "Jiang" for 1 - difference between the sum of IC at pairs of terms and 2*IC at MICA (see http://arxiv.org/pdf/cmp-lg/9709008.pdf), "Pesquita" for graph information content similarity related to Tanimoto-Jacard index (ie. summed information content of common ancestors divided by summed information content of all ancestors of term1 and term2 (see http://www.ncbi.nlm.nih.gov/pubmed/18460186)). By default, it uses "Schlicker" method
fast	logical to indicate whether a vectorised fast computation is used. By default, it sets to true. It is always advisable to use this vectorised fast computation; since the conventional computation is just used for understanding scripts

parallel	logical to indicate whether parallel computation with multicores is used. By default, it sets to true, but not necessarily does so. It will depend on whether these two packages "foreach" and "doParallel" have been installed. It can be installed via: `source("http://bioconductor.org/biocLite.R"); biocLite(c("foreach","doParallel"))` If not yet installed, this option will be disabled
multicores	an integer to specify how many cores will be registered as the multicore parallel backend to the 'foreach' package. If NULL, it will use a half of cores available in a user's computer. This option only works when parallel computation is enabled
verbose	logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

It returns a sparse matrix containing pair-wise semantic similarity between input terms. This sparse matrix can be converted to the full matrix via the function `as.matrix`

Note

none

See Also

dDAGinduce, dDAGancestor, dDAGgeneSim, dCheckParallel

Examples

```
## Not run:
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) load human genes annotated by HPPA
org.Hs.egHPPA <- dRDataLoader(RData='org.Hs.egHPPA')

# 3) prepare for ontology and its annotation information
dag <- dDAGannotate(g, annotations=org.Hs.egHPPA,
path.mode="all_paths", verbose=TRUE)

# 4) calculate pair-wise semantic similarity between 5 randomly chosen terms
terms <- sample(V(dag)$name, 5)
sim <- dDAGtermSim(g=dag, terms=terms, method="Schlicker",
parallel=FALSE)
sim

## End(Not run)
```

dDAGtip *Function to find the tip node(s) of a direct acyclic graph (DAG)*

Description

dDAGtip is supposed to find the tip node(s) of a direct acyclic graph (DAG; an ontology). It return the name (i.e Term ID) of the tip node(s).

Usage

```
dDAGtip(g)
```

Arguments

g	an object of class "igraph" or "graphNEL"

Value

- tip: the tip name (i.e. Term ID)

Note

none

See Also

dDAGtip

Examples

```
## Not run:
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) find tips
tips <- dDAGtip(g)
tips

## End(Not run)
```

dEnricher *Function to conduct enrichment analysis given the input data and the ontology in query*

Description

dEnricher is supposed to conduct enrichment analysis given the input data and the ontology in query. It returns an object of class "eTerm". Enrichment analysis is based on either Fisher's exact test or Hypergeometric test. The test can respect the hierarchy of the ontology.

Usage

```
dEnricher(
data,
identity = c("symbol", "entrez"),
check.symbol.identity = FALSE,
genome = c("Hs", "Mm", "Rn", "Gg", "Ce", "Dm", "Da", "At"),
ontology = c("GOBP", "GOMF", "GOCC", "PS", "PS2", "SF", "DO", "HPPA",
"HPMI", "HPCM",
"HPMA", "MP", "MsigdbH", "MsigdbC1", "MsigdbC2CGP", "MsigdbC2CP",
"MsigdbC2KEGG",
"MsigdbC2REACTOME", "MsigdbC2BIOCARTA", "MsigdbC3TFT", "MsigdbC3MIR",
"MsigdbC4CGN",
"MsigdbC4CM", "MsigdbC5BP", "MsigdbC5MF", "MsigdbC5CC", "MsigdbC6",
"MsigdbC7",
"DGIdb"),
sizeRange = c(10, 1000),
min.overlap = 3,
which_distance = NULL,
test = c("HypergeoTest", "FisherTest", "BinomialTest"),
p.adjust.method = c("BH", "BY", "bonferroni", "holm", "hochberg",
"hommel"),
ontology.algorithm = c("none", "pc", "elim", "lea"),
elim.pvalue = 0.01,
lea.depth = 2,
verbose = T,
RData.location =
"https://github.com/hfang-bristol/RDataCentre/blob/master/dnet/1.0.7"
)
```

Arguments

data	an input vector. It contains either Entrez Gene ID or Symbol
identity	the type of gene identity (i.e. row names of input data), either "symbol" for gene symbols (by default) or "entrez" for Entrez Gene ID. The option "symbol" is preferred as it is relatively stable from one update to another; also it is possible to search against synonyms (see the next parameter)

`check.symbol.identity`
: logical to indicate whether synonyms will be searched against when gene symbols cannot be matched. By default, it sets to FALSE since it may take a while to do such check using all possible synoyms

`genome`
: the genome identity. It can be one of "Hs" for human, "Mm" for mouse, "Rn" for rat, "Gg" for chicken, "Ce" for c.elegans, "Dm" for fruitfly, "Da" for zebrafish, and "At" for arabidopsis

`ontology`
: the ontology supported currently. It can be "GOBP" for Gene Ontology Biological Process, "GOMF" for Gene Ontology Molecular Function, "GOCC" for Gene Ontology Cellular Component, "PS" for phylostratific age information, "PS2" for the collapsed PS version (inferred ancestors being collapsed into one with the known taxonomy information), "SF" for domain superfamily assignments, "DO" for Disease Ontology, "HPPA" for Human Phenotype Phenotypic Abnormality, "HPMI" for Human Phenotype Mode of Inheritance, "HPCM" for Human Phenotype Clinical Modifier, "HPMA" for Human Phenotype Mortality Aging, "MP" for Mammalian Phenotype, and Drug-Gene Interaction database (DGIdb) and the molecular signatures database (Msigdb) only in human (including "MsigdbH", "MsigdbC1", "MsigdbC2CGP", "MsigdbC2CP", "MsigdbC2KEGG", "MsigdbC2REACTOME", "MsigdbC2BIOCARTA", "MsigdbC3TFT", "MsigdbC3MIR", "MsigdbC4CGN", "MsigdbC4CM", "MsigdbC5BP", "MsigdbC5MF", "MsigdbC5CC", "MsigdbC6", "MsigdbC7"). Note: These four ("GOBP", "GOMF", "GOCC" and "PS") are availble for all genomes/species; for "Hs" and "Mm", these six ("DO", "HPPA", "HPMI", "HPCM", "HPMA" and "MP") are also supported; all "Msigdb" are only supported in "Hs". For details on the eligibility for pairs of input genome and ontology, please refer to the online Documentations at http://supfam.org/dnet/docs.html

`sizeRange`
: the minimum and maximum size of members of each gene set in consideration. By default, it sets to a minimum of 10 but no more than 1000

`min.overlap`
: the minimum number of overlaps. Only those gene sets that overlap with input data at least min.overlap (3 by default) will be processed

`which_distance`
: which distance of terms in the ontology is used to restrict terms in consideration. By default, it sets to 'NULL' to consider all distances

`test`
: the statistic test used. It can be "FisherTest" for using fisher's exact test, "HypergeoTest" for using hypergeometric test, or "BinomialTest" for using binomial test. Fisher's exact test is to test the independence between gene group (genes belonging to a group or not) and gene annotation (genes annotated by a term or not), and thus compare sampling to the left part of background (after sampling without replacement). Hypergeometric test is to sample at random (without replacement) from the background containing annotated and non-annotated genes, and thus compare sampling to background. Unlike hypergeometric test, binomial test is to sample at random (with replacement) from the background with the constant probability. In terms of the ease of finding the significance, they are in order: hypergeometric test > binomial test > fisher's exact test. In other words, in terms of the calculated p-value, hypergeometric test < binomial test < fisher's exact test

`p.adjust.method`
: the method used to adjust p-values. It can be one of "BH", "BY", "bonferroni", "holm", "hochberg" and "hommel". The first two methods "BH" (widely used)

and "BY" control the false discovery rate (FDR: the expected proportion of false discoveries amongst the rejected hypotheses); the last four methods "bonferroni", "holm", "hochberg" and "hommel" are designed to give strong control of the family-wise error rate (FWER). Notes: FDR is a less stringent condition than FWER

`ontology.algorithm`
: the algorithm used to account for the hierarchy of the ontology. It can be one of "none", "pc", "elim" and "lea". For details, please see 'Note'

`elim.pvalue`
: the parameter only used when "ontology.algorithm" is "elim". It is used to control how to declare a signficantly enriched term (and subsequently all genes in this term are eliminated from all its ancestors)

`lea.depth`
: the parameter only used when "ontology.algorithm" is "lea". It is used to control how many maximum depth is uded to consider the children of a term (and subsequently all genes in these children term are eliminated from the use for the recalculation of the signifance at this term)

`verbose`
: logical to indicate whether the messages will be displayed in the screen. By default, it sets to false for no display

`RData.location`
: the characters to tell the location of built-in RData files. By default, it remotely locates at https://github.com/hfang-bristol/RDataCentre/blob/master/dnet and http://dnet.r-forge.r-project.org/RData. Be aware of several versions and the latest one is matched to the current package version. For the user equipped with fast internet connection, this option can be just left as default. But it is always advisable to download these files locally. Especially when the user needs to run this function many times, there is no need to ask the function to remotely download every time (also it will unnecessarily increase the runtime). For examples, these files (as a whole or part of them) can be first downloaded into your current working directory, and then set this option as: $RData.location = "."$. Surely, the location can be anywhere as long as the user provides the correct path pointing to (otherwise, the script will have to remotely download each time). Here is the UNIX command for downloading all RData files (preserving the directory structure): $wget - r - l2 - A" * .RData" - np - nH - - cut - dirs = 0" http://dnet.r - forge.r - project.org/RData"$

Value

an object of class "eTerm", a list with following components:

- `set_info`: a matrix of nSet X 4 containing gene set information, where nSet is the number of gene set in consideration, and the 4 columns are "setID" (i.e. "Term ID"), "name" (i.e. "Term Name"), "namespace" and "distance"
- `gs`: a list of gene sets, each storing gene members. Always, gene sets are identified by "setID" and gene members identified by "Entrez ID"
- `data`: a vector containing input data in consideration. It is not always the same as the input data as only those mappable are retained
- `overlap`: a list of overlapped gene sets, each storing genes overlapped between a gene set and the given input data (i.e. the genes of interest). Always, gene sets are identified by "setID" and gene members identified by "Entrez ID"

- `zscore`: a vector containing z-scores
- `pvalue`: a vector containing p-values
- `adjp`: a vector containing adjusted p-values. It is the p value but after being adjusted for multiple comparisons
- `call`: the call that produced this result

Note

The interpretation of the algorithms used to account for the hierarchy of the ontology is:

- "none": does not consider the ontology hierarchy at all.
- "lea": computers the significance of a term in terms of the significance of its children at the maximum depth (e.g. 2). Precisely, once genes are already annotated to any children terms with a more signficance than itself, then all these genes are eliminated from the use for the recalculation of the signifance at that term. The final p-values takes the maximum of the original p-value and the recalculated p-value.
- "elim": computers the significance of a term in terms of the significance of its all children. Precisely, once genes are already annotated to a signficantly enriched term under the cutoff of e.g. pvalue<1e-2, all these genes are eliminated from the ancestors of that term).
- "pc": requires the significance of a term not only using the whole genes as background but also using genes annotated to all its direct parents/ancestors as background. The final p-value takes the maximum of both p-values in these two calculations.
- "Notes": the order of the number of significant terms is: "none" > "lea" > "elim" > "pc".

See Also

dEnricherView

Examples

```
## Not run:
# load data
#library(Biobase)
#TCGA_mutations <- dRDataLoader(RData='TCGA_mutations')
#symbols <- as.character(fData(TCGA_mutations)$Symbol)

# Enrichment analysis using Disease Ontology (DO)
#data <- symbols[1:100] # select the first 100 human genes
#eTerm <- dEnricher(data, identity="symbol", genome="Hs", ontology="DO")

# visualise the top significant terms in the ontology hierarchy
#ig.DO <- dRDataLoader(RData='ig.DO')
#g <- ig.DO
#nodes_query <- names(sort(eTerm$adjp)[1:5])
#nodes.highlight <- rep("red", length(nodes_query))
#names(nodes.highlight) <- nodes_query
#subg <- dDAGinduce(g, nodes_query)
# color-code terms according to the adjust p-values (taking the form of 10-based negative logarithm)
#data <- -1*log10(eTerm$adjp[V(subg)$name])
```

```
#visDAG(g=subg, data=data, node.info="both", zlim=c(0,2), node.attrs=list(color=nodes.highlight))
# color-code terms according to the z-scores
#data <- eTerm$zscore[V(subg)$name]
#visDAG(g=subg, data=data, node.info="both", node.attrs=list(color=nodes.highlight))

## End(Not run)
```

---

dEnricherView *Function to view enrichment results of dEnricher*

Description

dEnricherView is supposed to view results of enrichment analysis by dEnricher.

Usage

```
dEnricherView(
eTerm,
top_num = 10,
sortBy = c("adjp", "pvalue", "zscore", "nAnno", "nOverlap", "none"),
decreasing = NULL,
details = F
)
```

Arguments

eTerm	an object of class "eTerm"
top_num	the maximum number of gene sets (terms) will be viewed
sortBy	which statistics will be used for sorting and viewing gene sets (terms). It can be "adjp" for adjusted p value, "pvalue" for p value, "zscore" for enrichment z-score, "nAnno" for the number of sets (terms), "nOverlap" for the number in overlaps, and "none" for ordering according to ID of gene sets (terms)
decreasing	logical to indicate whether to sort in a decreasing order. If it is null, it would be true for "zscore", "nAnno" or "nOverlap"; otherwise it would be false
details	logical to indicate whether the detailed information of gene sets (terms) is also viewed. By default, it sets to false for no inclusion

Value

a data frame with following components:

- setID: term ID; as rownames
- name: term name
- nAnno: number in gene members annotated by a term
- nOverlap: number in overlaps
- zscore: enrichment z-score

- pvalue: nominal p value
- adjp: adjusted p value
- namespace: term namespace; optional, it is only appended when "details" is true
- distance: term distance; optional, it is only appended when "details" is true
- members: members (represented as Gene Symbols) in overlaps; optional, it is only appended when "details" is true

Note

none

See Also

dEnricher

Examples

```
#dEnricherView(eTerm, top_num=10, sortBy="adjp", decreasing=FALSE, details=TRUE)
```

---

dFDRscore *Function to transform fdr into scores according to log-likelihood ratio between the true positives and the false positivies and/or after controlling false discovery rate*

Description

dFDRscore is supposed to take as input a vector of fdr, which are transformed into scores according to log-likelihood ratio between the true positives and the false positivies. Also if the FDR threshold is given, it is used to make sure that fdr below threshold are considered significant and thus scored positively. Instead, those fdr above the given threshold are considered insigificant and thus scored negatively.

Usage

```
dFDRscore(fdr, fdr.threshold = NULL, scatter = F)
```

Arguments

fdr	a vector containing a list of input fdr
fdr.threshold	the given FDR threshold. By default, it is set to NULL, meaning there is no constraint. If given, those fdr with the FDR below threshold are considered significant and thus scored positively. Instead, those fdr with the FDR above given threshold are considered insigificant and thus scored negatively
scatter	logical to indicate whether the scatter graph of scores against p-values should be drawn. Also indicated is the score corresponding to the given FDR threshold (if any)

Value

- scores: a vector of scores

Note

none

See Also

dSVDsignif, dNetPipeline

Examples

```
# 1) generate data with an iid matrix of 1000 x 9
data <- cbind(matrix(rnorm(1000*3,mean=0,sd=1), nrow=1000, ncol=3),
matrix(rnorm(1000*3,mean=0.5,sd=1), nrow=1000, ncol=3),
matrix(rnorm(1000*3,mean=-0.5,sd=1), nrow=1000, ncol=3))

# 2) calculate the significance according to SVD
# using "fdr" significance
fdr <- dSVDsignif(data, signif="fdr", num.permutation=10)

# 3) calculate the scores according to the fitted BUM and fdr=0.01
# no fdr threshold
scores <- dFDRscore(fdr)
# using fdr threshold of 0.01
scores <- dFDRscore(fdr, fdr.threshold=0.1, scatter=TRUE)
```

---

dFunArgs *Function to assign (and evaluate) arguments with default values for an input function*

Description

dFunArgs is supposed to assign (and evaluate) arguments with default values for an input function.

Usage

```
dFunArgs(fun, action = F, verbose = T)
```

Arguments

fun	an input function name (character string)
action	logical to indicate whether the function will act as it should be (with assigned values in the current environment). By default, it sets to FALSE
verbose	logical to indicate whether the messages will be displayed in the screen. By default, it sets to TRUE for display

Value

a list containing arguments and their default values

Note

This function is potentially useful when debugging. Because the developer does not have to specify default values for all arguments except those arguments are of interest

See Also

dNetPipeline

Examples

```
## Not run:
fun <- "dNetPipeline"
dFunArgs(fun)

## End(Not run)
```

---

dGSEA *Function to conduct gene set enrichment analysis given the input data and the ontology in query*

Description

dGSEA is supposed to conduct gene set enrichment analysis given the input data and the ontology in query. It returns an object of class "eTerm".

Usage

```
dGSEA(
data,
identity = c("symbol", "entrez"),
check.symbol.identity = FALSE,
genome = c("Hs", "Mm", "Rn", "Gg", "Ce", "Dm", "Da", "At"),
ontology = c("GOBP", "GOMF", "GOCC", "PS", "PS2", "SF", "DO", "HPPA",
"HPMI", "HPCM",
"HPMA", "MP", "MsigdbH", "MsigdbC1", "MsigdbC2CGP", "MsigdbC2CP",
"MsigdbC2KEGG",
"MsigdbC2REACTOME", "MsigdbC2BIOCARTA", "MsigdbC3TFT", "MsigdbC3MIR",
"MsigdbC4CGN",
"MsigdbC4CM", "MsigdbC5BP", "MsigdbC5MF", "MsigdbC5CC", "MsigdbC6",
"MsigdbC7",
"DGIdb", "Customised"),
customised.genesets = NULL,
sizeRange = c(10, 20000),
```

```
which_distance = NULL,
weight = 1,
nperm = 1000,
fast = T,
sigTail = c("two-tails", "one-tail"),
p.adjust.method = c("BH", "BY", "bonferroni", "holm", "hochberg",
"hommel"),
verbose = T,
RData.location =
"https://github.com/hfang-bristol/RDataCentre/blob/master/dnet/1.0.7"
)
```

Arguments

data	a data frame or matrix of input data. It must have row names, either Entrez Gene ID or Symbol
identity	the type of gene identity (i.e. row names of input data), either "symbol" for gene symbols (by default) or "entrez" for Entrez Gene ID. The option "symbol" is preferred as it is relatively stable from one update to another; also it is possible to search against synonyms (see the next parameter)
check.symbol.identity	logical to indicate whether synonyms will be searched against when gene symbols cannot be matched. By default, it sets to FALSE since it may take a while to do such check using all possible synoyms
genome	the genome identity. It can be one of "Hs" for human, "Mm" for mouse, "Rn" for rat, "Gg" for chicken, "Ce" for c.elegans, "Dm" for fruitfly, "Da" for zebrafish, and "At" for arabidopsis
ontology	the ontology supported currently. It can be "GOBP" for Gene Ontology Biological Process, "GOMF" for Gene Ontology Molecular Function, "GOCC" for Gene Ontology Cellular Component, "PS" for phylostratific age information, "PS2" for the collapsed PS version (inferred ancestors being collapsed into one with the known taxonomy information), "SF" for domain superfamily assignments, "DO" for Disease Ontology, "HPPA" for Human Phenotype Phenotypic Abnormality, "HPMI" for Human Phenotype Mode of Inheritance, "HPCM" for Human Phenotype Clinical Modifier, "HPMA" for Human Phenotype Mortality Aging, "MP" for Mammalian Phenotype, and Drug-Gene Interaction database (DGIdb) and the molecular signatures database (Msigdb) only in human (including "MsigdbH", "MsigdbC1", "MsigdbC2CGP", "MsigdbC2CP", "MsigdbC2KEGG", "MsigdbC2REACTOME", "MsigdbC2BIOCARTA", "MsigdbC3TFT", "MsigdbC3MIR", "MsigdbC4CGN", "MsigdbC4CM", "MsigdbC5BP", "MsigdbC5MF", "MsigdbC5CC", "MsigdbC6", "MsigdbC7"). Note: These four ("GOBP", "GOMF", "GOCC" and "PS") are availble for all genomes/species; for "Hs" and "Mm", these six ("DO", "HPPA", "HPMI", "HPCM", "HPMA" and "MP") are also supported; all "Msigdb" are only supported in "Hs". For details on the eligibility for pairs of input genome and ontology, please refer to the online Documentations at http://supfam.org/dnet/docs.html. Also supported are the user-customised gene sets; in doing so, the option "Customised" should be used together with the input of the next parameter "customised.genesets"

`customised.genesets`
: an input vector/matrix/list which only works when the user chooses "Customised" in the previous parameter "ontology". It contains either Entrez Gene ID or Symbol

`sizeRange`
: the minimum and maximum size of members of each gene set in consideration. By default, it sets to a minimum of 10 but no more than 1000

`which_distance`
: which distance of terms in the ontology is used to restrict terms in consideration. By default, it sets to 'NULL' to consider all distances

`weight`
: type of score weight. It can be "0" for unweighted (an equivalent to Kolmogorov-Smirnov, only considering the rank), "1" for weighted by input gene score (by default), and "2" for over-weighted, and so on

`nperm`
: the number of random permutations. For each permutation, gene-score associations will be permutated so that permutation of gene-term associations is realised

`fast`
: logical to indicate whether to fast calculate expected results from permutated data. By default, it sets to true

`sigTail`
: the tail used to calculate the statistical significance. It can be either "two-tails" for the significance based on two-tails or "one-tail" for the significance based on one tail

`p.adjust.method`
: the method used to adjust p-values. It can be one of "BH", "BY", "bonferroni", "holm", "hochberg" and "hommel". The first two methods "BH" (widely used) and "BY" control the false discovery rate (FDR: the expected proportion of false discoveries amongst the rejected hypotheses); the last four methods "bonferroni", "holm", "hochberg" and "hommel" are designed to give strong control of the family-wise error rate (FWER). Notes: FDR is a less stringent condition than FWER

`verbose`
: logical to indicate whether the messages will be displayed in the screen. By default, it sets to false for no display

`RData.location`
: the characters to tell the location of built-in RData files. By default, it remotely locates at https://github.com/hfang-bristol/RDataCentre/blob/master/dnet and http://dnet.r-forge.r-project.org/RData. Be aware of several versions and the latest one is matched to the current package version. For the user equipped with fast internet connection, this option can be just left as default. But it is always advisable to download these files locally. Especially when the user needs to run this function many times, there is no need to ask the function to remotely download every time (also it will unnecessarily increase the runtime). For examples, these files (as a whole or part of them) can be first downloaded into your current working directory, and then set this option as: $RData.location = "."$. Surely, the location can be anywhere as long as the user provides the correct path pointing to (otherwise, the script will have to remotely download each time). Here is the UNIX command for downloading all RData files (preserving the directory structure): $wget -r -l2 -A"*.RData" -np -nH --cut-dirs=0 "http://dnet.r-forge.r-project.org/RData"$

Value

an object of class "eTerm", a list with following components:

- `set_info`: a matrix of nSet X 4 containing gene set information, where nSet is the number of gene set in consideration, and the 4 columns are "setID" (i.e. "Term ID"), "name" (i.e. "Term Name"), "namespace" and "distance"
- `gs`: a list of gene sets, each storing gene members. Always, gene sets are identified by "setID" and gene members identified by "Entrez ID"
- `data`: a matrix of nGene X nSample containing input data in consideration. It is not always the same as the input data as only those mappable are retained
- `es`: a matrix of nSet X nSample containing enrichment score, where nSample is the number of samples (i.e. the number of columns in input data
- `nes`: a matrix of nSet X nSample containing normalised enrichment score. It is the version of enrichment score but after being normalised by gene set size
- `pvalue`: a matrix of nSet X nSample containing nominal p value
- `adjp`: a matrix of nSet X nSample containing adjusted p value. It is the p value but after being adjusted for multiple comparisons
- `gadjp`: a matrix of nSet X nSample containing globally adjusted p value in terms of all samples
- `fdr`: a matrix of nSet X nSample containing false discovery rate (FDR). It is the estimated probability that the normalised enrichment score represents a false positive finding
- `qvalue`: a matrix of nSet X nSample containing q value. It is the monotunically increasing FDR
- `weight`: the input type of score weight
- `call`: the call that produced this result

Note

The interpretation of returned components:

- "es": enrichment score for the gene set is the degree to which this gene set is overrepresented at the top or bottom of the ranked list of genes in each column of input data;
- "nes": normalised enrichment score for the gene set is enrichment score that has already normalised by gene set size. It is comparable across analysed gene sets;
- "pvalue": nominal p value is the statistical significance of the enrichment score. It is not adjusted for multiple hypothesis testing, and thus is of limited use in comparing gene sets;
- "adjp": adjusted p value by Benjamini & Hochberg method. It is comparable across gene sets;
- "gadjp": globally adjusted p value by Benjamini & Hochberg method. Unlike "adjp", it is adjusted in terms of all samples;
- "fdr": false discovery rate is the estimated probability that the normalised enrichment score represents a false positive finding. Unlike "adjp" or "gadjp" (also aliased as "fdr") that is derived from a list of p values, this version of fdr is directly calculate from the statistic (i.e. normalised enrichment score);
- "qvalue": q value is the monotunically increasing FDR so that the higher "nes", the lower "qvalue".

See Also

dGSEAview, dGSEAwrite, visGSEA

Examples

```
## Not run:
# load data
#library(Biobase)
#TCGA_mutations <- dRDataLoader(RData='TCGA_mutations')

# gene set enrichment analysis (GSEA) using KEGG pathways
## calculate the total mutations for each gene
#tol <- apply(exprs(TCGA_mutations), 1, sum)
#data <- data.frame(tol=tol)
#eTerm <- dGSEA(data, identity="symbol", genome="Hs", ontology="MsigdbC2KEGG")
#res <- dGSEAview(eTerm, which_sample=1, top_num=5, sortBy="adjp", decreasing=FALSE, details=TRUE)
#visGSEA(eTerm, which_sample=1, which_term=rownames(res)[1])
#output <- dGSEAwrite(eTerm, which_content="gadjp", which_score="gadjp", filename="eTerm.txt")

## based on customised gene sets
#eTerm <- dGSEA(data, ontology="Customised", customised.genesets=sample(rownames(data),100))
#res <- dGSEAview(eTerm, which_sample=1, top_num=5, sortBy="adjp", decreasing=FALSE, details=TRUE)
#visGSEA(eTerm, which_sample=1, which_term=rownames(res)[1])

## End(Not run)
```

---

dGSEAview *Function to view enrichment results in a sample-specific manner*

Description

dGSEAview is supposed to view results of gene set enrichment analysis but for a specific sample.

Usage

```
dGSEAview(
eTerm,
which_sample = 1,
top_num = 10,
sortBy = c("adjp", "gadjp", "ES", "nES", "pvalue", "FWER", "FDR",
"qvalue", "none"),
decreasing = NULL,
details = F
)
```

Arguments

eTerm	an object of class "eTerm"
which_sample	which sample will be viewed
top_num	the maximum number of gene sets will be viewed
sortBy	which statistics will be used for sorting and viewing gene sets. It can be "adjp" for adjusted p value, "gadjp" for globally adjusted p value, "ES" for enrichment score, "nES" for normalised enrichment score, "pvalue" for p value, "FWER" for family-wise error rate, "FDR" for false discovery rate, "qvalue" for q value, "none" for sorting by setID
decreasing	logical to indicate whether to sort in a decreasing order. If it is null, it would be true for "ES" or "nES"; otherwise it would be false
details	logical to indicate whether the detail information of gene sets is also viewed. By default, it sets to false for no inclusion

Value

a data frame with following components:

- setID: term ID
- ES: enrichment score
- nES: normalised enrichment score
- pvalue: nominal p value
- adjp: adjusted p value
- gadjp: globally adjusted p value
- FDR: false discovery rate
- qvalue: q value
- setSize: the number of genes in the set; optional, it is only appended when "details" is true
- name: term name; optional, it is only appended when "details" is true
- namespace: term namespace; optional, it is only appended when "details" is true
- distance: term distance; optional, it is only appended when "details" is true

Note

none

See Also

dGSEA

Examples

```
#dGSEAview(eTerm, which_sample=1, top_num=10, sortBy="adjp", decreasing=FALSE, details=TRUE)
```

dGSEAwrite *Function to write out enrichment results*

Description

dGSEAwrite is supposed to write out enrichment results.

Usage

```
dGSEAwrite(
eTerm,
which_content = c("gadjp", "adjp", "pvalue", "FWER", "FDR", "qvalue",
"nES", "ES"),
which_score = c("gadjp", "adjp", "FWER", "FDR", "qvalue", "nES"),
cutoff = 0.1,
filename = NULL,
keep.significance = T
)
```

Arguments

eTerm	an object of class "eTerm"
which_content	the content will be written out. It includes two categories: i) based on "adjp" for adjusted p value, "gadjp" for globally adjusted p value, "pvalue" for p value, "FWER" for family-wise error rate, "FDR" for false discovery rate, "qvalue" for q value; ii) based on "ES" for enrichment score, "nES" for normalised enrichment score. For the former, the content is : first -1*log10-transformed, and then multiplied by -1 if nES is negative.
which_score	which statistics/score will be used for declaring the significance. It can be "adjp" for adjusted p value, "gadjp" for globally adjusted p value, "FWER" for family-wise error rate, "FDR" for false discovery rate, "qvalue" for q value
cutoff	a cutoff to declare the signficance. It should be used together with 'which_score'
filename	a character string naming a filename
keep.significance	logical to indicate whether or not to mask those insignfiicant by NA. By default, it sets to true to mask those insignfiicant by NA

Value

a data frame with following components:

- setID: term ID
- setSize: the number of genes in the set
- name: term name
- namespace: term namespace
- distance: term distance
- sample names: sample names in the next columns

Note

If "filename" is not NULL, a tab-delimited text file will be also written out.

See Also

dGSEA

Examples

```
#output <- dGSEAwrite(eTerm, which_content="gadjp", which_score="gadjp", filename="eTerm.txt")
```

---

dNetConfidence *Function to append the confidence information from the source graphs into the target graph*

Description

eConsensusGraph is supposed to append the confidence information (extracted from a list of the source graphs) into the target graph. The confidence information is about how often a node (or an edge) in the target graph that can be found in the input source graphs. The target graph is an object of class "igraph" or "graphNEL", and the source graphs are a list of objects of class "igraph" or "graphNEL". It also returns an object of class "igraph" or "graphNEL"; specifically, the same as the input target graph but appended with the "nodeConfidence" attribute to the nodes and the "edgeConfidence" attribute to the edges.

Usage

```
dNetConfidence(target, sources, plot = F)
```

Arguments

target	the target graph, an object of class "igraph" or "graphNEL"
sources	a list of the source graphs, each with an object of class "igraph" or "graphNEL". These source graphs will be used to calculate how often a node (or an edge) in the target graph that can be found with them.
plot	logical to indicate whether the returned graph (i.e. the target graph plus the confidence information on nodes and edges) should be plotted. If it sets true, the plot will display the returned graph with the size of nodes indicative of the node confidence (the frequency that a node appears in the source graphs), and with the width of edges indicative of the edge confidence (the frequency that an edge appears in the source graphs)

Value

an object of class "igraph" or "graphNEL", which is a target graph but appended with the "nodeConfidence" attribute to the nodes and the "edgeConfidence" attribute to the edges (in the form of 100 percentage)

Note

None

See Also

visNet

Examples

```
# 1) generate a target graph according to the ER model
g <- erdos.renyi.game(100, 1/100)
target <- dNetInduce(g, V(g), knn=0)

# 2) generate a list source graphs according to the ER model
sources <- lapply(1:100, function(x) erdos.renyi.game(100*runif(1),
1/10))

# 3) append the confidence information from the source graphs into the target graph
g <- dNetConfidence(target=target, sources=sources)

# 4) visualise the confidence target graph
visNet(g, vertex.size=V(g)$nodeConfidence/10,
edge.width=E(g)$edgeConfidence)
```

---

dNetFind *Function to find heuristically maximum scoring subgraph*

Description

dNetFind is supposed to find the maximum scoring subgraph from an input graph and scores imposed on its nodes. The input graph and the output subgraph are both of "igraph" or "graphNET" object. The input scores imposed on the nodes in the input graph can be divided into two parts: the positive nodes and the negative nodes. The searching for maximum scoring subgraph is deduced to find the connected subgraph containing the positive nodes as many as possible, but the negative nodes as few as possible. To this end, a heuristic search is used (see Note below).

Usage

```
dNetFind(g, scores)
```

Arguments

g	an object of class "igraph" or "graphNEL"
scores	a vector of scores. For each element, it must have the name that could be mapped onto the input graph. Also, the names in input "scores" should contain all those in the input graph "g", but the reverse is not necessary

Value

a subgraph with a maximum score, an object of class "igraph" or "graphNEL". It has node attributes 'score' and 'type' (either 'desired' or 'linker')

Note

The search procedure is heuristic to find the subgraph with the maximum score:

- i) transform the input graph into a new graph by collapsing connected positive nodes into a meta-node. As such, meta-nodes are isolated to each other but are linked via negative nodes (single-nodes). Clearly, meta-nodes have positive scores, and negative scores for the single-nodes.
- ii) append the weight attribute to the edges in the transformed graph. There are two types of edges: 1) the single-single edge with two single-nodes as two ends, and 2) single-meta edge with a single-node as one end and a meta-node as the other end. The weight for a single-single edge is the absolute sum of the scores in its two-end single-nodes but normalised by their degrees. The weight for a single-meta edge is simply the absolute score in its single-node end normalised by the degree. As such, weights are all non-negative.
- iii) find minimum spanning tree (MST) in the weighted transformed graph using Prim's greedy algorithm. A spanning tree of the weighted graph is a subgraph that is tree and connects all the node together. The MST is a spanning tree with the sum of its edge weights minimised amongst all possible spanning trees.
- iv) find all shortest paths between any pair of meta-nodes in the MST. Within the weighted transformed graph in ii), a subgraph is induced containing nodes (only occuring in these shortest paths) and all edges between them.
- v) within the induced subgraph, identify single-nodes that are direct neighbors of meta-nodes. For each of these single-nodes, also make sure it has the absolute scores no more than the sum of scores in its neighboring meta-nodes. These single-nodes meeting both criteria are called "linkers".
- vi) still within the induced subgraph in v), find the linker graph that contains only linkers and edges between them. Similarly to iii), find MST of the linker graph, called 'linker MST'. Notably, this linker MST serves as the scaffold, which only contains linkers but has meta-nodes being direcly attached to.
- vii) in linker MST plus its attached meta-nodes, find the optimal path that has the sum of scores of its nodes and attached meta-nodes maximised amongest all possible paths. Nodes along this optimal path plus their attached meta-nodes are called 'subgraph nodes'.
- viii) finally, from the input graph extract a subgraph (called 'subgraph') that only contains subgraph nodes and edges betwen them. This subgraph is the maximum scoring subgraph containing the positive nodes as many as possible, but the negative nodes as few as possible.

See Also

dNetFind

Examples

```
# 1) generate an vector consisting of random values from beta distribution
x <- rbeta(1000, shape1=0.5, shape2=1)

# 2) fit a p-value distribution under beta-uniform mixture model
fit <- dBUMfit(x, ntry=1, hist.bum=FALSE, contour.bum=FALSE)

# 3) calculate the scores according to the fitted BUM and fdr=0.01
# using "pdf" method
scores <- dBUMscore(fit, method="pdf", fdr=0.05, scatter.bum=FALSE)
names(scores) <- as.character(1:length(scores))

# 4) generate a random graph according to the ER model
g <- erdos.renyi.game(1000, 1/100)

# 5) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)

# 6) find the subgraph with the maximum score
subgraph <- dNetFind(subg, scores)
```

---

dNetInduce *Function to generate a subgraph induced by given vertices and their k nearest neighbors*

Description

dNetInduce is supposed to produce a subgraph induced by given vertices and its k nearest neighbors. The input is a graph of "igraph" or "graphNET" object, a list of the vertices of the graph, and a k value for finding k nearest neighbors for these vertices. The output is a subgraph induced by given vertices plus their k neighbours. The resultant subgraph inherits the class from the input one. The induced subgraph contains exactly the vertices of interest, and all the edges between them.

Usage

```
dNetInduce(
g,
nodes_query,
knn = 0,
remove.loops = F,
largest.comp = T,
min.comp.size = 1
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
nodes_query	the vertices for which the calculation is performed

knn	an integeter specifying how many k steps are used to find the nearest neighbours of the given vertices. By default, knn is set to zero; it means no neighbors will be considered. When knn is 1, the immediate neighbors of the given vertices will be also considered for inducing the subgraph. The same is true when knn is 2, etc
remove.loops	logical to indicate whether the loop edges are to be removed. By default, it sets to false
largest.comp	logical to indicate whether the largest component is only retained. By default, it sets to true for the largest component being left
min.comp.size	an integer specifying the minimum size of component that will be retained. This parameter only works when setting the false to keep the largest component. By default, it sets to 1 meaning all nodes will be retained

Value

- subg: an induced subgraph, an object of class "igraph" or "graphNEL". Appended with a node attribute 'comp' if multiple components are kept

Note

The given vertices plus their k nearest neighbors will be used to induce the subgraph.

See Also

dNetInduce

Examples

```
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/100)

# 2) select the first 10 vertices as the query nodes
nodes_query <- V(g)[1:10]

# 3) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, nodes_query, knn=0)

# 4) produce the induced subgraph based on the nodes in query ane their immediate neighbours
subg <- dNetInduce(g, nodes_query, knn=1)
```

---

dNetPipeline *Function to setup the pipeline for finding maximum-scoring subgraph from an input graph and the signficance imposed on its nodes*

Description

dNetPipeline is supposed to finish ab inito maximum-scoring subgraph identification for the input graph with the node information on the significance (p-value or fdr). It returns an object of class "igraph" or "graphNEL".

Usage

```
dNetPipeline(
g,
pval,
method = c("pdf", "cdf", "customised"),
significance.threshold = NULL,
nsize = NULL,
plot = F,
verbose = T
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
pval	a vector containing input p-values (or fdr). For each element, it must have the name that could be mapped onto the input graph. Also, the names in input "pval" should contain all those in the input graph "g", but the reverse is not necessary
method	the method used for the transformation. It can be either "pdf" for the method based on the probability density function of the fitted model, or "cdf" for the method based on the cumulative distribution function of the fitted model
significance.threshold	the given significance threshold. By default, it is set to NULL, meaning there is no constraint. If given, those p-values below this are considered significant and thus scored positively. Instead, those p-values above this given significance threshold are considered insigificant and thus scored negatively
nsize	the desired number of nodes constrained to the resulting subgraph. It is not nulll, a wide range of significance thresholds will be scanned to find the optimal significance threshold leading to the desired number of nodes in the resulting subgraph. Notably, the given significance threshold will be overwritten by this option.
plot	logical to indicate whether the histogram plot, contour plot and scatter plot should be drawn. By default, it sets to false for no plotting
verbose	logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

a subgraph with a maximum score, an object of class "igraph" or "graphNEL". It has node attributes 'score' and 'type' (either 'desired' or 'linker'). Also appended is a graph attribute 'threshold' (that is, 'significance.threshold' used particularly useful when 'nsize' is defined)

Note

The pipeline sequentially consists of:

- ia) if the method is either "pdf" or "cdf", dBUMfit used to fit the p-value distribution under beta-uniform mixture model, and dBUMscore used to calculate the scores according to the fitted BUM and the significance threshold.

- ib) if the method is either "customised", then the user input list of fdr (or p-values) and the significance threshold will be directly used for score transformation by dFDRscore.
- ii) if there is the desired number of nodes constrained to the resulting subgraph, a wide range of significance thresholds (including rough stage with large intervals, and finetune stage with smaller intervals) will be scanned to find the significance threshold to meet the desired number of nodes.
- iii) dNetFind used to find maximum-scoring subgraph from the input graph and scores imposed on its nodes.

See Also

dBUMfit, dBUMscore, dFDRscore, dNetFind

Examples

```
## Not run:
# 1) generate an vector consisting of random values from beta distribution
x <- rbeta(1000, shape1=0.5, shape2=1)
names(x) <- as.character(1:length(x))

# 2) generate a random graph according to the ER model
g <- erdos.renyi.game(1000, 1/100)

# 3) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)

# 4) find maximum-scoring subgraph based on the given significance threshold
# 4a) assume the input is a list of p-values (controlling fdr=0.1)
subgraph <- dNetPipeline(g=subg, pval=x, significance.threshold=0.1)
# 4b) assume the input is a list of customised significance (eg FDR directly)
subgraph <- dNetPipeline(g=subg, pval=x, method="customised",
significance.threshold=0.1)

# 5) find maximum-scoring subgraph with the desired node number nsize=20
subgraph <- dNetPipeline(g=subg, pval=x, nsize=20)

## End(Not run)
```

---

dNetReorder *Function to reorder the multiple graph colorings within a sheet-shape rectangle grid*

Description

dNetReorder is reorder the multiple graph colorings within a sheet-shape rectangle grid

Usage

```
dNetReorder(
g,
data,
feature = c("node", "edge"),
node.normalise = c("none", "degree"),
xdim = NULL,
ydim = NULL,
amplifier = NULL,
metric = c("none", "pearson", "spearman", "kendall", "euclidean",
"manhattan", "cos",
"mi"),
init = c("linear", "uniform", "sample"),
algorithm = c("sequential", "batch"),
alphaType = c("invert", "linear", "power"),
neighKernel = c("gaussian", "bubble", "cutgaussian", "ep", "gamma")
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
data	an input data matrix used to color-code vertices/nodes. One column corresponds to one graph node coloring. The input matrix must have row names, and these names should include all node names of input graph, i.e. V(g)$name, since there is a mapping operation. After mapping, the length of the patern vector should be the same as the number of nodes of input graph. The way of how to color-code is to map values in the pattern onto the whole colormap (see the next arguments: colormap, ncolors, zlim and colorbar)
feature	the type of the features used. It can be one of either 'edge' for the edge feature or 'node' for the node feature. See 'Note' for explanations.
node.normalise	the normalisation of the nodes. It can be one of either 'none' for no normalisation or 'degree' for a node being penalised by its degree.
xdim	an integer specifying x-dimension of the grid
ydim	an integer specifying y-dimension of the grid
amplifier	an integer specifying the amplifier (3 by default) of the number of component planes. The product of the component number and the amplifier constitutes the number of rectangles in the sheet grid
metric	distance metric used to define the similarity between component planes. It can be "none", which means directly using column-wise vectors of codebook/data matrix. Otherwise, first calculate the covariance matrix from the codebook/data matrix. The distance metric used for calculating the covariance matrix between component planes can be: "pearson" for pearson correlation, "spearman" for spearman rank correlation, "kendall" for kendall tau rank correlation, "euclidean" for euclidean distance, "manhattan" for cityblock distance, "cos" for cosine similarity, "mi" for mutual information.

`init`	an initialisation method. It can be one of "uniform", "sample" and "linear" initialisation methods
`algorithm`	the training algorithm. Currently, only "sequential" algorithm has been implemented
`alphaType`	the alpha type. It can be one of "invert", "linear" and "power" alpha types
`neighKernel`	the training neighbor kernel. It can be one of "gaussian", "bubble", "cutgaussian", "ep" and "gamma" kernels

Value

an object of class "sReorder", a list with following components:

- `nHex`: the total number of rectanges in the grid
- `xdim`: x-dimension of the grid
- `ydim`: y-dimension of the grid
- `uOrder`: the unique order/placement for each component plane that is reordered to the "sheet"-shape grid with rectangular lattice
- `coord`: a matrix of nHex x 2, with each row corresponding to the coordinates of each "uOrder" rectangle in the 2D map grid
- `call`: the call that produced this result

Note

According to which features are used and whether nodes should be penalised by degrees, the feature data are constructed differently from the input data and input graph:

- When the node features are used, the feature data is the input data (or penalised data) with the same dimension.
- When the edge featrues are used, each entry (i.e. given an edge and a sample) in the feature data is the absolute difference between its two-end nodes (or after being penalised).
- After that, the constructed feature are subject to sample correlation analysis by supraHex. That is, a map grid (with sheet shape consisting of a rectangular lattice) is used to train either column-wise vectors of the feature data matrix or the covariance matrix thereof.
- As a result, similar samples are placed closer to each other within this map grid. More precisely, to ensure the unique placement, each sample mapped to the "sheet"-shape grid with rectangular lattice is determinied iteratively in an order from the best matched to the next compromised one. If multiple samples are hit in the same rectangular lattice, the worse one is always sacrificed by moving to the next best one till all samples are placed somewhere exclusively on their own.

The size of "sheet"-shape rectangle grid depends on the input arguments:

- How the input parameters are used to determine nHex is taken priority in the following order: "xdim & ydim" > "nHex" > "data".
- If both of xdim and ydim are given, $nHex = xdim * ydim$.
- If only data is input, $nHex = 5 * sqrt(dlen)$, where dlen is the number of rows of the input data.
- After nHex is determined, xy-dimensions of rectangle grid are then determined according to the square root of the two biggest eigenvalues of the input data.

See Also

visNetReorder

Examples

```
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/100)

# 2) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)

# 3) reorder the module with vertices being color-coded by input data
nnodes <- vcount(subg)
nsamples <- 10
data <- matrix(runif(nnodes*nsamples), nrow=nnodes, ncol=nsamples)
rownames(data) <- V(subg)$name
sReorder <- dNetReorder(g=subg, data, feature="node",
node.normalise="none")
```

---

dPvalAggregate *Function to aggregate p values*

Description

dPvalAggregate is supposed to aggregate a input matrix p-values into a vector of aggregated p-values. The aggregate operation is applied to each row of input matrix, each resulting in an aggregated p-value. The method implemented can be based on the order statistics of p-values or according to Fisher's method or Z-transform method.

Usage

```
dPvalAggregate(
pmatrix,
method = c("orderStatistic", "fishers", "Ztransform", "logistic"),
order = ncol(pmatrix),
weight = rep(1, ncol(pmatrix))
)
```

Arguments

pmatrix	a data frame or matrix of p-values
method	the method used. It can be either "orderStatistic" for the method based on the order statistics of p-values, or "fishers" for Fisher's method (summation of logs), or "Ztransform" for Z-transform test (summation of z values, Stouffer's method) and the weighted Z-test, or "logistic" for summation of logits
order	an integeter specifying the order used for the aggregation according to the order statistics of p-values
weight	a vector specifying the weights used for the aggregation according to Z-transform method

Value

- ap: a vector with the length nrow(pmatrix), containing aggregated p-values

Note

For each row of input matrix with the c columns, there are c p-values that are uniformly independently distributed over [0,1] under the null hypothesis (uniform distribution). According to the order statisitcs, they follow the Beta distribution with the paramters $a = order$ and $b = c - order + 1$. According to the Fisher's method, after transformation by $-2 * \sum^c log(pvalue)$, they follow Chi-Squared distribution. According to the Z-transform method, first converts the one-tailed P-values into standard normal deviates Z, then combines Z via $\frac{\sum^c (w*Z)}{\sqrt{\sum^c (w^2)}}$, where w is the weight (usually square root of the sample size if the weighted Z-test; 1 if Z-transform test), and finally the combined Z follows the standard normal distribution to test the cumulative/aggregated evidence on the common null hypothesis. The logistic method is defined as $\sum^c log(\frac{pvalue}{1-pvalue}) * 1/C$, where $C = sqrt((kpi^2(5k+2))/(3(5k+4)))$, following Student's t distribution. Generally speaking, Fisher's method places greater emphasis on small p-values, while the Z-transform method on equal footings, the logistic method provides a compromise between these two. In other words, the Z-transform method does well in problems where evidence against the combined null is spread more than a small fraction of the individual tests, or when the total evidence is weak; Fisher's method does best in problems where the evidence is concentrated in a relatively small fraction of the individual tests or when the evidence is at least moderately strong.

See Also

dPvalAggregate

Examples

```
# 1) generate an iid uniformly-distributed random matrix of 1000x3
pmatrix <- cbind(runif(1000), runif(1000), runif(1000))

# 2) aggregate according to the order statistics
ap <- dPvalAggregate(pmatrix, method="orderStatistic")

# 3) aggregate according to the Fisher's method
ap <- dPvalAggregate(pmatrix, method="fishers")

# 4) aggregate according to the Z-transform method
ap <- dPvalAggregate(pmatrix, method="Ztransform")

# 5) aggregate according to the logistic method
ap <- dPvalAggregate(pmatrix, method="logistic")
```

dRDataLoader *Function to load dnet built-in RData*

Description

dRDataLoader is supposed to load the package built-in RData.

Usage

```
dRDataLoader(
RData = c(NA, "TCGA_mutations", "ig.DO", "ig.GOBP", "ig.GOCC",
"ig.GOMF", "ig.HPCM",
"ig.HPMA", "ig.HPMI", "ig.HPPA", "ig.MP", "org.At.eg", "org.At.egGOBP",
"org.At.egGOCC", "org.At.egGOMF", "org.At.egPS", "org.At.egSF",
"org.At.string",
"org.Ce.eg", "org.Ce.egGOBP", "org.Ce.egGOCC", "org.Ce.egGOMF",
"org.Ce.egPS",
"org.Ce.egSF", "org.Ce.string", "org.Da.eg", "org.Da.egGOBP",
"org.Da.egGOCC",
"org.Da.egGOMF", "org.Da.egPS", "org.Da.egSF", "org.Da.string",
"org.Dm.eg",
"org.Dm.egGOBP", "org.Dm.egGOCC", "org.Dm.egGOMF", "org.Dm.egPS",
"org.Dm.egSF",
"org.Dm.string", "org.Gg.eg", "org.Gg.egGOBP", "org.Gg.egGOCC",
"org.Gg.egGOMF",
"org.Gg.egPS", "org.Gg.egSF", "org.Gg.string", "org.Hs.eg",
"org.Hs.egDGIdb",
"org.Hs.egDO", "org.Hs.egGOBP", "org.Hs.egGOCC", "org.Hs.egGOMF",
"org.Hs.egHPCM",
"org.Hs.egHPMA", "org.Hs.egHPMI", "org.Hs.egHPPA", "org.Hs.egMP",
"org.Hs.egMsigdbC1",
"org.Hs.egMsigdbC2BIOCARTA", "org.Hs.egMsigdbC2CGP",
"org.Hs.egMsigdbC2CP",
"org.Hs.egMsigdbC2KEGG", "org.Hs.egMsigdbC2REACTOME",
"org.Hs.egMsigdbC3MIR",
"org.Hs.egMsigdbC3TFT", "org.Hs.egMsigdbC4CGN", "org.Hs.egMsigdbC4CM",
"org.Hs.egMsigdbC5BP", "org.Hs.egMsigdbC5CC", "org.Hs.egMsigdbC5MF",
"org.Hs.egMsigdbC6", "org.Hs.egMsigdbC7", "org.Hs.egMsigdbH",
"org.Hs.egPS",
"org.Hs.egSF", "org.Hs.string", "org.Mm.eg", "org.Mm.egDO",
"org.Mm.egGOBP",
"org.Mm.egGOCC", "org.Mm.egGOMF", "org.Mm.egHPCM", "org.Mm.egHPMA",
"org.Mm.egHPMI",
"org.Mm.egHPPA", "org.Mm.egMP", "org.Mm.egPS", "org.Mm.egSF",
"org.Mm.string",
"org.Rn.eg", "org.Rn.egGOBP", "org.Rn.egGOCC", "org.Rn.egGOMF",
"org.Rn.egPS",
```

```
"org.Rn.egSF", "CLL", "org.Rn.string"),
genome = c(NA, "Hs", "Mm", "Rn", "Gg", "Ce", "Dm", "Da", "At"),
ontology = c(NA, "GOBP", "GOMF", "GOCC", "PS", "PS2", "SF", "DO",
"HPPA", "HPMI",
"HPCM", "HPMA", "MP", "MsigdbH", "MsigdbC1", "MsigdbC2CGP",
"MsigdbC2CP",
"MsigdbC2KEGG", "MsigdbC2REACTOME", "MsigdbC2BIOCARTA", "MsigdbC3TFT",
"MsigdbC3MIR",
"MsigdbC4CGN", "MsigdbC4CM", "MsigdbC5BP", "MsigdbC5MF", "MsigdbC5CC",
"MsigdbC6",
"MsigdbC7", "DGIdb"),
verbose = T,
RData.location =
"https://github.com/hfang-bristol/RDataCentre/blob/master/dnet/1.0.7"
)
```

Arguments

RData	which built-in RData to load. It can be one of "TCGA_mutations", "ig.DO", "ig.GOBP", "ig.GOCC", "ig.GOMF", "ig.HPCM", "ig.HPMA", "ig.HPMI", "ig.HPPA", "ig.MP", "org.At.eg", "org.At.egGOBP", "org.At.egGOCC", "org.At.egGOMF", "org.At.egPS", "org.At.egSF", "org.At.string", "org.Ce.eg", "org.Ce.egGOBP", "org.Ce.egGOCC", "org.Ce.egGOMF", "org.Ce.egPS", "org.Ce.egSF", "org.Ce.string", "org.Da.eg", "org.Da.egGOBP", "org.Da.egGOCC", "org.Da.egGOMF", "org.Da.egPS", "org.Da.egSF", "org.Da.string", "org.Dm.eg", "org.Dm.egGOBP", "org.Dm.egGOCC", "org.Dm.egGOMF", "org.Dm.egPS", "org.Dm.egSF", "org.Dm.string", "org.Gg.eg", "org.Gg.egGOBP", "org.Gg.egGOCC", "org.Gg.egGOMF", "org.Gg.egPS", "org.Gg.egSF", "org.Gg.string", "org.Hs.eg", "org.Hs.egDGIdb", "org.Hs.egDO", "org.Hs.egGOBP", "org.Hs.egGOCC", "org.Hs.egGOMF", "org.Hs.egHPCM", "org.Hs.egHPMA", "org.Hs.egHPMI", "org.Hs.egHPPA", "org.Hs.egMP", "org.Hs.egMsigdbC1", "org.Hs.egMsigdbC2BIOC "org.Hs.egMsigdbC2CGP", "org.Hs.egMsigdbC2CP", "org.Hs.egMsigdbC2KEGG", "org.Hs.egMsigdbC2REACTOME", "org.Hs.egMsigdbC3MIR", "org.Hs.egMsigdbC3TFT", "org.Hs.egMsigdbC4CGN", "org.Hs.egMsigdbC4CM", "org.Hs.egMsigdbC5BP", "org.Hs.egMsigdbC5CC", "org.Hs.egMsigdbC5MF", "org.Hs.egMsigdbC6", "org.Hs.egMsigdbC7", "org.Hs.egMsigdbH", "org.Hs.egPS", "org.Hs.egSF", "org.Hs.string", "org.Mm.eg", "org.Mm.egDO", "org.Mm.egGOBP", "org.Mm.egGOCC", "org.Mm.egGOMF", "org.Mm.egHPCM", "org.Mm.egHPMA", "org.Mm.egHPMI", "org.Mm.egHPPA", "org.Mm.egMP", "org.Mm.egPS", "org.Mm.egSF", "org.Mm.string", "org.Rn.eg", "org.Rn.egGOBP", "org.Rn.egGOCC", "org.Rn.egGOMF", "org.Rn.egPS", "org.Rn.egSF", "CLL", "org.Rn.string". On the meanings, please refer to the Documentations at http://supfam.org/dnet/docs.html
genome	the genome identity. It can be one of "Hs" for human, "Mm" for mouse, "Rn" for rat, "Gg" for chicken, "Ce" for c.elegans, "Dm" for fruitfly, "Da" for zebrafish, and "At" for arabidopsis
ontology	the ontology supported currently. It can be "GOBP" for Gene Ontology Biological Process, "GOMF" for Gene Ontology Molecular Function, "GOCC" for Gene Ontology Cellular Component, "PS" for phylostratific age information, "PS2" for the collapsed PS version (inferred ancestors being collapsed

into one with the known taxonomy information), "SF" for domain superfamily assignments, "DO" for Disease Ontology, "HPPA" for Human Phenotype Phenotypic Abnormality, "HPMI" for Human Phenotype Mode of Inheritance, "HPCM" for Human Phenotype Clinical Modifier, "HPMA" for Human Phenotype Mortality Aging, "MP" for Mammalian Phenotype, and Drug-Gene Interaction database (DGIdb) and the molecular signatures database (Msigdb) only in human (including "MsigdbH", "MsigdbC1", "MsigdbC2CGP", "MsigdbC2CP", "MsigdbC2KEGG", "MsigdbC2REACTOME", "MsigdbC2BIOCARTA", "MsigdbC3TFT", "MsigdbC3MIR", "MsigdbC4CGN", "MsigdbC4CM", "MsigdbC5BP", "MsigdbC5MF", "MsigdbC5CC", "MsigdbC6", "MsigdbC7"). Note: These four ("GOBP", "GOMF", "GOCC" and "PS") are availble for all genomes/species; for "Hs" and "Mm", these six ("DO", "HPPA", "HPMI", "HPCM", "HPMA" and "MP") are also supported; all "Msigdb" are only supported in "Hs". For details on the eligibility for pairs of input genome and ontology, please refer to the online Documentations at http://supfam.org/dnet/docs.html

`verbose` logical to indicate whether the messages will be displayed in the screen. By default, it sets to TRUE for display

`RData.location` the characters to tell the location of built-in RData files. By default, it remotely locates at https://github.com/hfang-bristol/RDataCentre/blob/master/dnet and http://dnet.r-forge.r-project.org/RData. Be aware of several versions and the latest one is matched to the current package version. For the user equipped with fast internet connection, this option can be just left as default. But it is always advisable to download these files locally. Especially when the user needs to run this function many times, there is no need to ask the function to remotely download every time (also it will unnecessarily increase the runtime). For examples, these files (as a whole or part of them) can be first downloaded into your current working directory, and then set this option as: $RData.location = "."$. Surely, the location can be anywhere as long as the user provides the correct path pointing to (otherwise, the script will have to remotely download each time). Here is the UNIX command for downloading all RData files (preserving the directory structure): $wget -r -l2 -A "*.RData" -np -nH --cut-dirs=0 "http://dnet.r-forge.r-project.org/RData"$

Value

any use-specified variable that is given on the right side of the assigement sign '<-', which contains the loaded RData.

Note

If there are no use-specified variable that is given on the right side of the assigement sign '<-', then no RData will be loaded onto the working environment.

See Also

dRDataLoader

Examples

```
## Not run:
org.Hs.egSF <- dRDataLoader(RData='org.Hs.egSF')
org.Hs.eg <- dRDataLoader(RData='org.Hs.eg')
org.Hs.egDGIdb <- dRDataLoader(RData='org.Hs.egDGIdb')
org.Hs.egMsigdbC2KEGG <- dRDataLoader(RData='org.Hs.egMsigdbC2KEGG')
org.Hs.egHPPA <- dRDataLoader(genome='Hs', ontology='HPPA')
ig.MP <- dRDataLoader(RData='ig.MP')

## End(Not run)
```

---

dRWR *Function to implement Random Walk with Restart (RWR) on the input graph*

Description

dRWR is supposed to implement Random Walk with Restart (RWR) on the input graph. If the seeds (i.e. a set of starting nodes) are given, it intends to calculate the affinity score of all nodes in the graph to the seeds. If the seeds are not given, it will pre-compute affinity matrix for nodes in the input graph with respect to each starting node (as a seed) by looping over every node in the graph. Parallel computing is also supported for Linux or Mac operating systems.

Usage

```
dRWR(
g,
normalise = c("laplacian", "row", "column", "none"),
setSeeds = NULL,
restart = 0.75,
normalise.affinity.matrix = c("none", "quantile"),
parallel = TRUE,
multicores = NULL,
verbose = T
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
normalise	the way to normalise the adjacency matrix of the input graph. It can be 'laplacian' for laplacian normalisation, 'row' for row-wise normalisation, 'column' for column-wise normalisation, or 'none'
setSeeds	an input matrix used to define sets of starting seeds. One column corresponds to one set of seeds that a walker starts with. The input matrix must have row names, coming from node names of input graph, i.e. V(g)$name, since there is a mapping operation. The non-zero entries mean that the corresonding rows (i.e. the gene/row names) are used as the seeds, and non-zero values can be viewed

as how to weight the relative importance of seeds. By default, this option sets to "NULL", suggesting each node in the graph will be used as a set of the seed to pre-compute affinity matrix for the input graph. This default does not scale for large input graphs since it will loop over every node in the graph; however, the pre-computed affinity matrix can be extensively reused for obtaining affinity scores between any combinations of nodes/seeds, allows for some flexibility in the downstream use, in particular when sampling a large number of random node combinations for statistical testing

`restart` the restart probability used for RWR. The restart probability takes the value from 0 to 1, controlling the range from the starting nodes/seeds that the walker will explore. The higher the value, the more likely the walker is to visit the nodes centered on the starting nodes. At the extreme when the restart probability is zero, the walker moves freely to the neighbors at each step without restarting from seeds, i.e., following a random walk (RW)

`normalise.affinity.matrix`
the way to normalise the output affinity matrix. It can be 'none' for no normalisation, 'quantile' for quantile normalisation to ensure that columns (if multiple) of the output affinity matrix have the same quantiles

`parallel` logical to indicate whether parallel computation with multicores is used. By default, it sets to true, but not necessarily does so. It will depend on whether these two packages "foreach" and "doParallel" have been installed. It can be installed via: `source("http://bioconductor.org/biocLite.R"); biocLite(c("foreach","doParallel"))` If not yet installed, this option will be disabled

`multicores` an integer to specify how many cores will be registered as the multicore parallel backend to the 'foreach' package. If NULL, it will use a half of cores available in a user's computer. This option only works when parallel computation is enabled

`verbose` logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

It returns a sparse matrix, called 'PTmatrix':

- When the seeds are NOT given: a pre-computated affinity matrix with the dimension of n X n, where n is the number of nodes in the input graph. Columns stand for starting nodes walking from, and rows for ending nodes walking to. Therefore, a column for a starting node represents a steady-state affinity vector that the starting node will visit all the ending nodes in the graph
- When the seeds are given: an affinity matrix with the dimension of n X nset, where n is the number of nodes in the input graph, and nset for the number of the sets of seeds (i.e. the number of columns in setSeeds). Each column stands for the steady probability vector, storing the affinity score of all nodes in the graph to the starting nodes/seeds. This steady probability vector can be viewed as the "influential impact" over the graph imposed by the starting nodes/seeds.

Note

The input graph will treat as an unweighted graph if there is no 'weight' edge attribute associated with

See Also

dRWRcontact, dRWRpipeline, dCheckParallel

Examples

```
## Not run:
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/100)

# 2) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)
V(subg)$name <- 1:vcount(subg)

# 3) obtain the pre-computated affinity matrix
PTmatrix <- dRWR(g=subg, normalise="laplacian", restart=0.75,
parallel=FALSE)
# visualise affinity matrix
visHeatmapAdv(PTmatrix, Rowv=FALSE, Colv=FALSE, colormap="wyr",
KeyValueName="Affinity")

# 4) obtain affinity matrix given sets of seeds
# define sets of seeds
# each seed with equal weight (i.e. all non-zero entries are '1')
aSeeds <- c(1,0,1,0,1)
bSeeds <- c(0,0,1,0,1)
setSeeds <- data.frame(aSeeds,bSeeds)
rownames(setSeeds) <- 1:5
# calcualte affinity matrix
PTmatrix <- dRWR(g=subg, normalise="laplacian", setSeeds=setSeeds,
restart=0.75, parallel=FALSE)
PTmatrix

## End(Not run)
```

---

dRWRcontact — *Function to estimate RWR-based contact strength between samples from an input gene-sample data matrix, an input graph and its pre-computed affinity matrix*

Description

dRWRcontact is supposed to estimate sample relationships (ie. contact strength between samples) from an input gene-sample matrix, an input graph and its affinity matrix pre-computed according to random walk restart (RWR) of the input graph. It includes: 1) RWR-smoothed columns of input gene-sample matrix based on the pre-computed affinity matrix; 2) calculation of contact strength (inner products of RWR-smooth columns of input gene-sample matrix); 3) estimation of the contact signficance by a randomalisation procedure. Parallel computing is also supported for Linux or Mac operating systems.

Usage

```
dRWRcontact(
data,
g,
Amatrix,
permutation = c("random", "degree"),
num.permutation = 10,
p.adjust.method = c("BH", "BY", "bonferroni", "holm", "hochberg",
"hommel"),
adjp.cutoff = 0.05,
parallel = TRUE,
multicores = NULL,
verbose = T
)
```

Arguments

data	an input gene-sample data matrix used for seeds. Each value in input gene-sample matrix does not necessarily have to be binary (non-zeros will be used as a weight, but should be non-negative for easy interpretation).
g	an object of class "igraph" or "graphNEL"
Amatrix	an affinity matrix pre-computed from the input graph. Notes: columns for starting nodes walking from, and rows for ending nodes walking to
permutation	how to do permutation. It can be 'degree' for degree-preserving permutation, 'random' for permutation purely in random
num.permutation	the number of permutations used to for generating the distribution of contact strength under randomalisation
p.adjust.method	the method used to adjust p-values. It can be one of "BH", "BY", "bonferroni", "holm", "hochberg" and "hommel". The first two methods "BH" (widely used) and "BY" control the false discovery rate (FDR: the expected proportion of false discoveries amongst the rejected hypotheses); the last four methods "bonferroni", "holm", "hochberg" and "hommel" are designed to give strong control of the family-wise error rate (FWER). Notes: FDR is a less stringent condition than FWER
adjp.cutoff	the cutoff of adjusted pvalue to construct the contact graph
parallel	logical to indicate whether parallel computation with multicores is used. By default, it sets to true, but not necessarily does so. It will depend on whether these two packages "foreach" and "doParallel" have been installed. It can be installed via: `source("http://bioconductor.org/biocLite.R"); biocLite(c("foreach","doParallel"))` If not yet installed, this option will be disabled
multicores	an integer to specify how many cores will be registered as the multicore parallel backend to the 'foreach' package. If NULL, it will use a half of cores available in a user's computer. This option only works when parallel computation is enabled
verbose	logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

an object of class "dContact", a list with following components:

- `ratio`: a symmetric matrix storing ratio (the observed against the expected) between pairwise samples
- `zscore`: a symmetric matrix storing zscore between pairwise samples
- `pval`: a symmetric matrix storing pvalue between pairwise samples
- `adjpval`: a symmetric matrix storing adjusted pvalue between pairwise samples
- `cgraph`: the constructed contact graph (as a 'igraph' object) under the cutoff of adjusted value
- `call`: the call that produced this result

Note

none

See Also

dRWR, dCheckParallel

Examples

```
## Not run:
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/100)

# 2) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)
V(subg)$name <- 1:vcount(subg)

# 3) pre-compute affinity matrix from the input graph
Amatrix <- dRWR(g=subg, parallel=FALSE)

# 4) estimate RWR-based sample relationships
# define sets of seeds as data
# each seed with equal weight (i.e. all non-zero entries are '1')
aSeeds <- c(1,0,1,0,1)
bSeeds <- c(0,0,1,0,1)
data <- data.frame(aSeeds,bSeeds)
rownames(data) <- 1:5
# calcualte their two contacts
dContact <- dRWRcontact(data=data, g=subg, Amatrix=Amatrix,
parallel=FALSE)
dContact

## End(Not run)
```

dRWRpipeline *Function to setup a pipeine to estimate RWR-based contact strength between samples from an input gene-sample data matrix and an input graph*

Description

dRWRpipeline is supposed to estimate sample relationships (ie. contact strength between samples) from an input gene-sample matrix and an input graph. The pipeline includes: 1) random walk restart (RWR) of the input graph using the input matrix as seeds; 2) calculation of contact strength (inner products of RWR-smoothed columns of input matrix); 3) estimation of the contact signficance by a randomalisation procedure. It supports two methods how to use RWR: 'direct' for directly applying RWR in the given seeds; 'indirectly' for first pre-computing affinity matrix of the input graph, and then deriving the affinity score. Parallel computing is also supported for Linux or Mac operating systems.

Usage

```
dRWRpipeline(
data,
g,
method = c("direct", "indirect"),
normalise = c("laplacian", "row", "column", "none"),
restart = 0.75,
normalise.affinity.matrix = c("none", "quantile"),
permutation = c("random", "degree"),
num.permutation = 10,
p.adjust.method = c("BH", "BY", "bonferroni", "holm", "hochberg",
"hommel"),
adjp.cutoff = 0.05,
parallel = TRUE,
multicores = NULL,
verbose = T
)
```

Arguments

data	an input gene-sample data matrix used for seeds. Each value in input gene-sample matrix does not necessarily have to be binary (non-zeros will be used as a weight, but should be non-negative for easy interpretation).
g	an object of class "igraph" or "graphNEL"
method	the method used to calculate RWR. It can be 'direct' for directly applying RWR, 'indirect' for indirectly applying RWR (first pre-compute affinity matrix and then derive the affinity score)
normalise	the way to normalise the adjacency matrix of the input graph. It can be 'laplacian' for laplacian normalisation, 'row' for row-wise normalisation, 'column' for column-wise normalisation, or 'none'

`restart` the restart probability used for RWR. The restart probability takes the value from 0 to 1, controlling the range from the starting nodes/seeds that the walker will explore. The higher the value, the more likely the walker is to visit the nodes centered on the starting nodes. At the extreme when the restart probability is zero, the walker moves freely to the neighbors at each step without restarting from seeds, i.e., following a random walk (RW)

`normalise.affinity.matrix` the way to normalise the output affinity matrix. It can be 'none' for no normalisation, 'quantile' for quantile normalisation to ensure that columns (if multiple) of the output affinity matrix have the same quantiles

`permutation` how to do permutation. It can be 'degree' for degree-preserving permutation, 'random' for permutation in random

`num.permutation` the number of permutations used to for generating the distribution of contact strength under randomalisation

`p.adjust.method` the method used to adjust p-values. It can be one of "BH", "BY", "bonferroni", "holm", "hochberg" and "hommel". The first two methods "BH" (widely used) and "BY" control the false discovery rate (FDR: the expected proportion of false discoveries amongst the rejected hypotheses); the last four methods "bonferroni", "holm", "hochberg" and "hommel" are designed to give strong control of the family-wise error rate (FWER). Notes: FDR is a less stringent condition than FWER

`adjp.cutoff` the cutoff of adjusted pvalue to construct the contact graph

`parallel` logical to indicate whether parallel computation with multicores is used. By default, it sets to true, but not necessarily does so. It will depend on whether these two packages "foreach" and "doParallel" have been installed. It can be installed via: `source("http://bioconductor.org/biocLite.R"); biocLite(c("foreach","doParallel"))` If not yet installed, this option will be disabled

`multicores` an integer to specify how many cores will be registered as the multicore parallel backend to the 'foreach' package. If NULL, it will use a half of cores available in a user's computer. This option only works when parallel computation is enabled

`verbose` logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

an object of class "dContact", a list with following components:

- `ratio`: a symmetric matrix storing ratio (the observed against the expected) between pairwise samples
- `zscore`: a symmetric matrix storing zscore between pairwise samples
- `pval`: a symmetric matrix storing pvalue between pairwise samples
- `adjpval`: a symmetric matrix storing adjusted pvalue between pairwise samples
- `cgraph`: the constructed contact graph (as a 'igraph' object) under the cutoff of adjusted value
- `Amatrix`: a pre-computated affinity matrix when using 'inderect' method; NULL otherwise
- `call`: the call that produced this result

Note

The choice of which method to use RWR depends on the number of seed sets and the number of permutations for statistical test. If the total product of both numbers are huge, it is better to use 'indirect' method (for a single run). However, if the user wants to re-use pre-computed affinity matrix (ie. re-use the input graph a lot), then it is highly recommended to sequentially use dRWR and dRWRcontact instead.

See Also

dRWR, dRWRcontact, dCheckParallel

Examples

```
## Not run:
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/100)

# 2) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)
V(subg)$name <- 1:vcount(subg)

# 3) estimate RWR dating based sample relationships
# define sets of seeds as data
# each seed with equal weight (i.e. all non-zero entries are '1')
aSeeds <- c(1,0,1,0,1)
bSeeds <- c(0,0,1,0,1)
data <- data.frame(aSeeds,bSeeds)
rownames(data) <- 1:5
# calcualte their two contact graph
dContact <- dRWRpipeline(data=data, g=subg, parallel=FALSE)
dContact

## End(Not run)
```

---

dSVDsignif *Function to obtain SVD-based gene significance from the input gene-sample matrix*

Description

dSVDsignif is supposed to obtain gene signficance from the given gene-sample matrix according to singular value decomposition (SVD)-based method. The method includes: 1) singular value decomposition of the input matrix; 2) determination of the eigens in consideration (if not given); 3) construction of the gene-specific project vector based on the considered eigens; 4) calculation of the distance statistic from the projection vector to zero point vector; and 5) based on distance statistic to obtain the gene significance.

Usage

```
dSVDsignif(
data,
num.eigen = NULL,
pval.eigen = 0.01,
signif = c("fdr", "pval"),
orient.permutation = c("row", "column", "both"),
num.permutation = 100,
fdr.procedure = c("stepup", "stepdown"),
verbose = T
)
```

Arguments

data	an input gene-sample data matrix used for singular value decomposition
num.eigen	an integer specifying the number of eigens in consideration. If NULL, this number will be automatically decided on based on the observed relative eigenexpression against randomised relative eigenexpression calculated from a list (here 100) of permutated input matrix
pval.eigen	p-value used to call those eigens as dominant. This parameter is used only when parameter 'num.eigen' is NULL. Here, p-value is calcualted to assess how likely the observed relative eigenexpression are more than the maximum relative eigenexpression calculated from permutated matrix
signif	the singificance to return. It can be either "pval" for using the p-value as the gene significance, or "fdr" for using the fdr as the gene significance
orient.permutation	the orientation of matrix being permutated. It can be either "row" to permutate values within each row, or "column" to permutate values within each column, or "both" to permutate values both within rows and columns. Notably, when using the p-value as the gene significance, it is always to permutate values within each row.
num.permutation	an integer specifying how many permutations are used
fdr.procedure	the procedure to adjust the fdr. To ensure that the high distance statistic the more significance, the fdr should be adjusted either using "stepup" for step-up procedure (from the most significant to the least significant) or using "stepdown" for step-down procedure (from the least significant to the most significant)
verbose	logical to indicate whether the messages will be displayed in the screen. By default, it sets to true for display

Value

a vector storing gene significance

Note

none

See Also

dFDRscore

Examples

```
## Not run:
# 1) generate data with an iid matrix of 1000 x 9
data <- cbind(matrix(rnorm(1000*3,mean=0,sd=1), nrow=1000, ncol=3),
matrix(rnorm(1000*3,mean=0.5,sd=1), nrow=1000, ncol=3),
matrix(rnorm(1000*3,mean=-0.5,sd=1), nrow=1000, ncol=3))

# 2) calculate the significance according to SVD
# using "fdr" significance
fdr <- dSVDsignif(data, signif="fdr", num.permutation=10)
# using "pval" significance
pval <- dSVDsignif(data, signif="pval", num.permutation=10)

## End(Not run)
```

---

ig.HPPA *Human Phenotype Phenotypic Abnormality (HPPA).*

Description

An R object that contains information on Human Phenotype Phenotypic Abnormality terms. These terms are organised as a direct acyclic graph (DAG), which is further stored as an object of the class 'igraph' (see http://igraph.org/r/doc/aaa-igraph-package.html). This data is prepared based on http://purl.obolibrary.org/obo/hp.obo.

Usage

```
data(ig.HPPA)
```

Value

an object of class "igraph". As a direct graph, it has attributes to vertices/nodes and edges:

- vertex attributes: "name" (i.e. "Term ID"), "term_id" (i.e. "Term ID"), "term_name" (i.e "Term Name") and "term_distance" (i.e. Term Distance: the distance to the root; always 0 for the root itself)
- edge attributes: "relation" (either 'is_a' or 'part_of')

References

Robinson et al. (2012) The Human Phenotype Ontology: a tool for annotating and analyzing human hereditary disease. *Am J Hum Genet*, 83:610-615.

Examples

```
ig.HPPA <- dRDataLoader(RData='ig.HPPA')
ig.HPPA
```

`org.Hs.egHPPA` *Annotations of Human Entrez Genes (EG) by Human Phenotype Phenotypic Abnormality (HPPA).*

Description

An R object that contains associations between HPPA terms and Human Entrez Genes. This data is first prepared based on http://purl.obolibrary.org/obo/hp.obo and http://compbio.charite.de/hudson/job/hpo.annotations.monthly/lastStableBuild/artifact/annotation/ALL_SOURCES_ALL_FREQUENCIES_genes_to_phenotype.txt.

Usage

```
data(org.Hs.egHPPA)
```

Value

an object of class "GS", a list with following components:

- `set_info`: a matrix of nSet X 4 containing gene set information, where nSet is the number of gene sets (i.e. HPPA terms), and the 4 columns are "setID" (i.e. "Term ID"), "name" (i.e. "Term Name"), "namespace" and "distance"
- `gs`: a list of gene sets, each storing gene members thereof. Always, gene sets are identified by "setID" and gene members identified by "Entrez ID"

References

Robinson et al. (2012) The Human Phenotype Ontology: a tool for annotating and analyzing human hereditary disease. *Am J Hum Genet*, 83:610-615.

Examples

```
org.Hs.egHPPA <- dRDataLoader(RData='org.Hs.egHPPA')
names(org.Hs.egHPPA)
```

visBoxplotAdv *Function to visualise a data frame using advanced boxplot*

Description

visBoxplotAdv is supposed to visualise a data frame using advanced boxplot. In addition to boxplot, a scatter plot is also drawn with various methods to avoid co-incident points so that each point is visible (with fine-controling the color and plotting character). Also, these points can be pies or thermometers, which allows an additional proportation data to be visualised as well.

Usage

```
visBoxplotAdv(
formula,
data,
orientation = c("vertical", "horizontal"),
method = c("center", "hex", "square", "swarm"),
corral = c("none", "gutter", "wrap", "random", "omit"),
corralWidth,
cex = 1,
spacing = 1,
breaks = NULL,
labels,
at = NULL,
add = FALSE,
log = FALSE,
xlim = NULL,
ylim = NULL,
xlab = NULL,
ylab = NULL,
pch = c("circles", "thermometers", "pies")[1],
col = graphics::par("col"),
bg = NA,
pwpch = NULL,
pwcol = NULL,
pwbg = NULL,
pwpie = NULL,
do.plot = TRUE,
do.boxplot = TRUE,
boxplot.notch = FALSE,
boxplot.border = "#888888C0",
boxplot.col = "transparent",
...
)
```

Arguments

formula	a formula, such as 'y ~ grp', where 'y' is a numeric vector of data values to be split into groups according to the grouping variable 'grp' (usually a factor)
data	a data.frame (or list) from which the variables in 'formula' should be taken.
orientation	the orientation. It can be one of "vertical" for the vertical orientation, "horizontal" for the horizontal orientation
method	the method for arranging the points. It can be one of "swarm" for arranging points in increasing order (if a point would overlap an existing point, it is shifted sideways (along the group axis) by a minimal amount sufficient to avoid overlap), "center" for first discretizing the values along the data axis (in order to create more efficient packing) and then using a square grid to produce a symmetric swarm, "hex" for first discretization and then arranging points in a hexagonal grid, and "square" for first discretization and then arranging points in a square grid
corral	the method to adjust points that would be placed outside their own group region. It can be one of "none" for not adjusting runaway points, "gutter" for collecting runaway points along the boundary between groups, "wrap" for wrapping runaway points to produce periodic boundaries, "random" for placing runaway points randomly in the region, and "omit" for omitting runaway points
corralWidth	the width of the "corral" in user coordinates
cex	size of points relative to the default. This must be a single value
spacing	relative spacing between points
breaks	breakpoints (optional). If NULL, breakpoints are chosen automatically
labels	labels for each group. Recycled if necessary. By default, these are inferred from the data
at	numeric vector giving the locations where the swarms should be drawn; defaults to '1:n' where n is the number of groups
add	whether to add to an existing plot
log	whether to use a logarithmic scale on the data axis
xlim	limits for x-axis
ylim	limits for y-axis
xlab	labels for x-aixs
ylab	labels for y-aixs
pch	plotting characters, specified by group and recycled if necessary. In additon to the convertional pch values, it can also be "circles", "thermometers", or "pies". For "pies" (or "thermometers"), users can also specify the proportional values (see below "pwpie") to visualise another information in the pie (or themometer) chart
col	plotting colors, specified by group and recycled if necessary
bg	plotting background, specified by group and recycled if necessary
pwpch	point-wise version of pch
pwcol	point-wise version of col

pwbg	point-wise version of bg
pwpie	point-wise proportion used when drawing pies or themometers
do.plot	whether to draw main plot
do.boxplot	whether to draw boxplot. It only works when the main plot is drawn
boxplot.notch	whether to draw a notch in the boxplot. If the notches of two plots do not overlap this is 'strong evidence' that the two medians differ
boxplot.border	the color for the outlines of the boxplots
boxplot.col	the color for the bodies of the boxplots
...	additional graphic parameters for the plot

Value

A data frame with plotting information. It has the same row names as the input data

Note

none

See Also

visBoxplotAdv

Examples

```
## Not run:
#data(TCGA_mutations)
#pd <- Biobase::pData(TCGA_mutations)
# only tumor types "LAML" or "BLCA"
#data <- pd[pd$TCGA_tumor_type=="LAML" | pd$TCGA_tumor_type=="BLCA",]
#labels <- levels(as.factor(data$TCGA_tumor_type))
# colors for gender
#pwcol <- as.numeric((data$Gender))
# pie for relative age
#pwpie <- data$Age/(max(data$Age))
#out <- visBoxplotAdv(formula=time~TCGA_tumor_type, data=data, pch="pies", pwcol=pwcol, pwpie=pwpie)
#legend("topright", legend=levels(data$Gender), box.col="transparent", pch=19, col=unique(pwcol))

## End(Not run)
```

---

visDAG *Function to visualise a direct acyclic graph (DAG) with node colorings according to a named input data vector (if provided)*

Description

visDAG is supposed to visualise a direct acyclic graph (DAG) with node colorings according to a named input data vector (if provided)

Usage

```
visDAG(
g,
data = NULL,
height = 7,
width = 7,
margin = rep(0.1, 4),
colormap = c("yr", "bwr", "jet", "gbr", "wyr", "br", "rainbow", "wb",
"lightyellow-orange"),
ncolors = 40,
zlim = NULL,
colorbar = T,
colorbar.fraction = 0.1,
newpage = T,
layout.orientation = c("left_right", "top_bottom", "bottom_top",
"right_left"),
node.info = c("none", "term_id", "term_name", "both",
"full_term_name"),
numChar = 15,
graph.node.attrs = NULL,
graph.edge.attrs = NULL,
node.attrs = NULL
)
```

Arguments

g	an object of class "igraph"
data	a named input data verctor used to color-code vertices/nodes. The input data vector must have names, and these names should include all node names of input graph, i.e. V(g)$name, since there is a mapping operation. The way of how to color-code is to map values in the data onto the whole colormap (see the next arguments: colormap, ncolors, zlim and colorbar)
height	a numeric value specifying the height of device
width	a numeric value specifying the width of device
margin	margins as units of length 4 or 1
colormap	short name for the colormap. It can be one of "jet" (jet colormap), "bwr" (blue-white-red colormap), "gbr" (green-black-red colormap), "wyr" (white-yellow-red colormap), "br" (black-red colormap), "yr" (yellow-red colormap), "wb" (white-black colormap), and "rainbow" (rainbow colormap, that is, red-yellow-green-cyan-blue-magenta). Alternatively, any hyphen-separated HTML color names, e.g. "lightyellow-orange" (by default), "blue-black-yellow", "royalblue-white-sandybrown", "darkgreen-white-darkviolet". A list of standard color names can be found in http://html-color-codes.info/color-names
ncolors	the number of colors specified over the colormap
zlim	the minimum and maximum z/data values for which colors should be plotted, defaulting to the range of the finite values of z. Each of the given colors will be

used to color an equispaced interval of this range. The midpoints of the intervals cover the range, so that values just outside the range will be plotted

`colorbar` logical to indicate whether to append a colorbar. If data is null, it always sets to false

`colorbar.fraction`
the relative fraction of colorbar block against the device size

`newpage` logical to indicate whether to open a new page. By default, it sets to true for opening a new page

`layout.orientation`
the orientation of the DAG layout. It can be one of "left_right" for the left-right layout (viewed from the DAG root point), "top_bottom" for the top-bottom layout, "bottom_top" for the bottom-top layout, and "right_left" for the right-left layout

`node.info` tells the ontology term information used to label nodes. It can be one of "none" for no node labeling, "term_id" for using Term ID, "term_name" for using Term Name (the first 15 characters), "both" for using both of Term ID and Name (the first 15 characters), and "full_term_name" for using the full Term Name

`numChar` a positive integer specifying wrap width of node labelling

`graph.node.attrs`
a list of global node attributes. These node attributes will be changed globally. See 'Note' below for details on the attributes

`graph.edge.attrs`
a list of global edge attributes. These edge attributes will be changed globally. See 'Note' below for details on the attributes

`node.attrs` a list of local edge attributes. These node attributes will be changed locally; as such, for each attribute, the input value must be a named vector (i.e. using Term ID as names). See 'Note' below for details on the attributes

Value

An object of class 'Ragraph'

Note

A list of global node attributes used in "graph.node.attrs":

- "shape": the shape of the node: "circle", "rectangle", "rect", "box" and "ellipse"
- "fixedsize": the logical to use only width and height attributes. By default, it sets to true for not expanding for the width of the label
- "fillcolor": the background color of the node
- "color": the color for the node, corresponding to the outside edge of the node
- "fontcolor": the color for the node text/labelings
- "fontsize": the font size for the node text/labelings
- "height": the height (in inches) of the node: 0.5 by default
- "width": the width (in inches) of the node: 0.75 by default

- "style": the line style for the node: "solid", "dashed", "dotted", "invis" and "bold"

A list of global edge attributes used in "graph.edge.attrs":

- "color": the color of the edge: gray by default
- "weight": the weight of the edge: 1 by default
- "style": the line style for the edge: "solid", "dashed", "dotted", "invis" and "bold"

A list of local node attributes used in "node.attrs" (only those named Term IDs will be changed locally!):

- "label": a named vector specifying the node text/labelings
- "shape": a named vector specifying the shape of the node: "circle", "rectangle", "rect", "box" and "ellipse"
- "fixedsize": a named vector specifying whether it sets to true for not expanding for the width of the label
- "fillcolor": a named vector specifying the background color of the node
- "color": a named vector specifying the color for the node, corresponding to the outside edge of the node
- "fontcolor": a named vector specifying the color for the node text/labelings
- "fontsize": a named vector specifying the font size for the node text/labelings
- "height": a named vector specifying the height (in inches) of the node: 0.5 by default
- "width": a named vector specifying the width (in inches) of the node: 0.75 by default
- "style": a named vector specifying the line style for the node: "solid", "dashed", "dotted", "invis" and "bold"

See Also

dDAGreverse, dDAGroot, dDAGinduce, dDAGlevel

Examples

```
## Not run:
# 1) load HPPA as igraph object
ig.HPPA <-dRDataLoader(RData='ig.HPPA')
g <- ig.HPPA

# 2) randomly select vertices as the query nodes
# the more common, the query nodes can be term id
nodes_query <- V(g)[sample(V(g),5)]$name

# 3) obtain the induced subgraph based on all possible paths
subg <- dDAGinduce(g, nodes_query, path.mode="all_paths")

# 4) just visualise the induced subgraph
visDAG(g=subg, node.info="both")

# 5) color-code nodes/terms according to its level
```

```
data <- dDAGlevel(subg)
visDAG(g=subg, data=data, node.info="both")
# 5a) globally change the node and edge attributes
visDAG(g=subg, data=data, layout.orientation="top_bottom",
node.info="both",
graph.node.attrs=list(fixedsize=FALSE,shape="box",color="transparent"),
graph.edge.attrs=list(color="black"))
# 5b) locally highlight the root by changing its shape into "box"
root <- dDAGroot(subg)
root.shape <- "box"
names(root.shape) <- V(subg)[root]$name
visDAG(g=subg, data=data, node.info="both",
node.attrs=list(shape=root.shape))
# 5c) further locally remove the root labelling
root.label <- ""
names(root.label) <- V(subg)[root]$name
visDAG(g=subg, data=data, node.info="both",
node.attrs=list(shape=root.shape,label=root.label))

## End(Not run)
```

---

visGSEA *Function to visualise running enrichment score for a given sample and a gene set*

Description

visGSEA is supposed to visualise running enrichment score for a given sample and a gene set. To help understand the underlying running enrichment score, the input gene scores are also displayed. Positions for members in the given gene set are color-coded in both displays (red line for the positive gene scores, and green line for the negative).

Usage

```
visGSEA(
eTerm,
which_sample = 1,
which_term = "GO:0006281",
plot = T,
orientation = c("vertical", "horizontal"),
hit.linewidth = 0.5,
newpage = T
)
```

Arguments

eTerm	an object of class "eTerm"
which_sample	which sample will be used. It can be index or sample names

which_term	which term will be used. It can be index or term ID or term names
plot	logical to indicate whether to plot
orientation	the orientation of the plots. It can be either "vertical" (default) or "horizontal"
hit.linewidth	the line width for the hits (ie genes in the gene set)
newpage	logical to indicate whether to open a new page. By default, it sets to true for opening a new page

Value

leading genes (being sorted)

Note

none

See Also

dGSEA, dGSEAview

Examples

```
#visGSEA(eTerm, which_sample=1, which_term=1)
```

---

visNet *Function to visualise a graph object of class "igraph" or "graphNEL"*

Description

visNet is supposed to visualise a graph object of class "igraph" or "graphNEL". It also allows the color-coding of vertices by providing the input pattern.

Usage

```
visNet(
g,
pattern = NULL,
colormap = c("bwr", "jet", "gbr", "wyr", "br", "yr", "rainbow", "wb"),
ncolors = 40,
zlim = NULL,
colorbar = T,
newpage = T,
glayout = layout.fruchterman.reingold,
vertex.frame.color = NA,
vertex.size = NULL,
vertex.color = NULL,
vertex.shape = NULL,
vertex.label = NULL,
```

```
vertex.label.cex = NULL,
vertex.label.dist = NULL,
vertex.label.color = "black",
vertex.label.family = "sans",
...
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
pattern	a numeric vector used to color-code vertices/nodes. Notably, if the input vector contains names, then these names should include all node names of input graph, i.e. V(g)$name, since there is a mapping operation. After mapping, the length of the patern vector should be the same as the number of nodes of input graph; otherwise, this input pattern will be ignored. The way of how to color-code is to map values in the pattern onto the whole colormap (see the next arguments: colormap, ncolors, zlim and colorbar)
colormap	short name for the colormap. It can be one of "jet" (jet colormap), "bwr" (blue-white-red colormap), "gbr" (green-black-red colormap), "wyr" (white-yellow-red colormap), "br" (black-red colormap), "yr" (yellow-red colormap), "wb" (white-black colormap), and "rainbow" (rainbow colormap, that is, red-yellow-green-cyan-blue-magenta). Alternatively, any hyphen-separated HTML color names, e.g. "blue-black-yellow", "royalblue-white-sandybrown", "darkgreen-white-darkviolet". A list of standard color names can be found in http://html-color-codes.info/color-names
ncolors	the number of colors specified over the colormap
zlim	the minimum and maximum z/patttern values for which colors should be plotted, defaulting to the range of the finite values of z. Each of the given colors will be used to color an equispaced interval of this range. The midpoints of the intervals cover the range, so that values just outside the range will be plotted
colorbar	logical to indicate whether to append a colorbar. If pattern is null, it always sets to false
newpage	logical to indicate whether to open a new page. By default, it sets to true for opening a new page
glayout	either a function or a numeric matrix configuring how the vertices will be placed on the plot. If layout is a function, this function will be called with the graph as the single parameter to determine the actual coordinates. This function can be one of "layout.auto", "layout.random", "layout.circle", "layout.sphere", "layout.fruchterman.reingold", "layout.kamada.kawai", "layout.spring", "layout.reingold.tilford", "layout.fruchterman.reingold.grid", "layout.lgl", "layout.graphopt", "layout.svd" and "layout.norm". A full explanation of these layouts can be found in http://igraph.org/r/doc/layout_nicely.html
vertex.frame.color	the color of the frame of the vertices. If it is NA, then there is no frame
vertex.size	the size of each vertex. If it is a vector, each vertex may differ in size

vertex.color
: the fill color of the vertices. If it is NA, then there is no fill color. If the pattern is given, this setup will be ignored

vertex.shape
: the shape of each vertex. It can be one of "circle", "square", "csquare", "rectangle", "crectangle", "vrectangle", "pie" (http://igraph.org/r/doc/vertex.shape.pie.html), "sphere", and "none". If it sets to NULL, these vertices with negative will be "csquare" and the rest "circle".

vertex.label
: the label of the vertices. If it is NA, then there is no label. The default vertex labels are the name attribute of the nodes

vertex.label.cex
: the font size of vertex labels.

vertex.label.dist
: the distance of the label from the center of the vertex. If it is 0 then the label is centered on the vertex. If it is 1 then the label is displayed beside the vertex.

vertex.label.color
: the color of vertex labels.

vertex.label.family
: the font family of vertex labels

...
: additional graphic parameters. See http://igraph.org/r/doc/plot.common.html for the complete list.

Value

invisible

Note

none

See Also

dNetFind

Examples

```
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/100)

# 2) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)

# 3) visualise the subg with vertices being color-coded by the pattern
pattern <- runif(vcount(subg))
names(pattern) <- V(subg)$name
visNet(g=subg, pattern=pattern, colormap="bwr", vertex.shape="sphere")
```

visNetAnimate *Function to animate the same graph but with multiple graph node colorings according to input data matrix*

Description

visNetAnimate is supposed to animate the same graph but with multiple colorings according to input data matrix. The output can be a pdf file containing a list of frames/images, a mp4 video file or a gif file. To support video output file, the software 'ffmpeg' must be first installed (also put its path into the system PATH variable; see Note). To support gif output file, the software 'ImageMagick' must be first installed (also put its path into the system PATH variable; see Note).

Usage

```
visNetAnimate(
g,
data,
filename = "visNetAnimate",
filetype = c("pdf", "mp4", "gif"),
image.type = c("jpg", "png"),
num.frame = ncol(data),
sec_per_frame = 1,
height.device = 7,
margin = rep(0.1, 4),
border.color = "#EEEEEE",
colormap = c("bwr", "jet", "gbr", "wyr", "br", "yr", "rainbow", "wb"),
ncolors = 40,
zlim = NULL,
colorbar = T,
colorbar.fraction = 0.25,
glayout = layout.fruchterman.reingold,
glayout.dynamics = F,
mtext.side = 3,
mtext.adj = 0,
mtext.cex = 1,
mtext.font = 2,
mtext.col = "black",
...
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
data	an input data matrix used to color-code vertices/nodes. One column corresponds to one graph node coloring. The input matrix must have row names, and these names should include all node names of input graph, i.e. V(g)$name, since there is a mapping operation. After mapping, the length of the patern vector should be

the same as the number of nodes of input graph. The way of how to color-code is to map values in the pattern onto the whole colormap (see the next arguments: colormap, ncolors, zlim and colorbar)

`filename` the without-extension part of the name of the output file. By default, it is 'visNetAnimate'

`filetype` the type of the output file, i.e. the extension of the output file name. It can be one of either 'pdf' for the pdf file, 'mp4' for the mp4 video file, 'gif' for the gif file

`image.type` the type of the image files temporarily generated. It can be one of either 'jpg' or 'png'. These temporary image files are used for producing mp4/gif output file. The reason doing so is to accommodate that sometimes only one of image types is supported so that you can choose the right one

`num.frame` a numeric value specifying the number of frames/images. By default, it sets to the number of columns in the input data matrix

`sec_per_frame` a numeric value specifying how long (seconds) it takes to stream a frame/image. This argument only works when producing mp4 video or gif file.

`height.device` a numeric value specifying the height (or width) of device/frame/image.

`margin` margins as units of length 4 or 1

`border.color` the border color of each figure

`colormap` short name for the colormap. It can be one of "jet" (jet colormap), "bwr" (blue-white-red colormap), "gbr" (green-black-red colormap), "wyr" (white-yellow-red colormap), "br" (black-red colormap), "yr" (yellow-red colormap), "wb" (white-black colormap), and "rainbow" (rainbow colormap, that is, red-yellow-green-cyan-blue-magenta). Alternatively, any hyphen-separated HTML color names, e.g. "blue-black-yellow", "royalblue-white-sandybrown", "darkgreen-white-darkviolet". A list of standard color names can be found in http://html-color-codes.info/color-names

`ncolors` the number of colors specified over the colormap

`zlim` the minimum and maximum z/patttern values for which colors should be plotted, defaulting to the range of the finite values of z. Each of the given colors will be used to color an equispaced interval of this range. The midpoints of the intervals cover the range, so that values just outside the range will be plotted

`colorbar` logical to indicate whether to append a colorbar. If pattern is null, it always sets to false

`colorbar.fraction` the relative fraction of colorbar block against the figure block

`glayout` either a function or a numeric matrix configuring how the vertices will be placed on the plot. If layout is a function, this function will be called with the graph as the single parameter to determine the actual coordinates. This function can be one of "layout.auto", "layout.random", "layout.circle", "layout.sphere", "layout.fruchterman.reingold", "layout.kamada.kawai", "layout.spring", "layout.reingold.tilford", "layout.fruchterman.reingold.grid", "layout.lgl", "layout.graphopt", "layout.svd" and "layout.norm". A full explanation of these layouts can be found in http://igraph.org/r/doc/layout_nicely.html

`glayout.dynamics`
: logical to indicate whether graph layout should be dynamic. By default, it always sets to false. If YES, the Fruchterman-Reingold layout algorithm http://igraph.org/r/doc/layout_with_fr.html will be used to stimulate the dynamic layout

`mtext.side`
: on which side of the mtext plot (1=bottom, 2=left, 3=top, 4=right)

`mtext.adj`
: the adjustment for mtext alignment (0 for left or bottom alignment, 1 for right or top alignment)

`mtext.cex`
: the font size of mtext labels

`mtext.font`
: the font weight of mtext labels

`mtext.col`
: the color of mtext labels

`...`
: additional graphic parameters. See http://igraph.org/r/doc/plot.common.html for the complete list.

Value

If specifying the output file name (see argument 'filename' above), the output file is either 'filename.pdf' or 'filename.mp4' or 'filename.gif' in the current working directory. If no output file name specified, by default the output file is either 'visNetAnimate.pdf' or 'visNetAnimate.mp4' or 'visNetAnimate.gif'

Note

When producing mp4 video, this function requires the installation of the software 'ffmpeg' at https://www.ffmpeg.org. Shell command lines for ffmpeg installation in Terminal (for both Linux and Mac) are:

- 1) `wget -O ffmpeg.tar.gz http://www.ffmpeg.org/releases/ffmpeg-2.7.1.tar.gz`
- 2) `mkdir ~/ffmpeg | tar xvfz ffmpeg.tar.gz -C ~/ffmpeg --strip-components=1`
- 3) `cd ffmpeg`
- 4a) # Assuming you want installation with a ROOT (sudo) privilege:
  `./configure --disable-yasm`
- 4b) # Assuming you want local installation without ROOT (sudo) privilege:
  `./configure --disable-yasm --prefix=$HOME/ffmpeg`
- 5) `make`
- 6) `make install`
- 7) # add the system PATH variable to your ~/.bash_profile file if you follow 4b) route:
  `export PATH=$HOME/ffmpeg:$PATH`
- 8) # make sure ffmpeg has been installed successfully:
  `ffmpeg -h`

When producing gif file, this function requires the installation of the software 'ImageMagick' at http://www.imagemagick.org. Shell command lines for ImageMagick installation in Terminal are:

- 1) `wget http://www.imagemagick.org/download/ImageMagick.tar.gz`
- 2) `mkdir ~/ImageMagick | tar xvzf ImageMagick.tar.gz -C ~/ImageMagick --strip-components=1`

- 3) cd ImageMagick
- 4) ./configure --prefix=$HOME/ImageMagick
- 5) make
- 6) make install
- 7) # add the system PATH variable to your ~/.bash_profile file.
  For Linux:
  export MAGICK_HOME=$HOME/ImageMagick
  export PATH=$MAGICK_HOME/bin:$PATH
  export LD_LIBRARY_PATH=${LD_LIBRARY_PATH:+$LD_LIBRARY_PATH:}$MAGICK_HOME/lib
  For Mac:
  export MAGICK_HOME=$HOME/ImageMagick
  export PATH=$MAGICK_HOME/bin:$PATH
  export DYLD_LIBRARY_PATH=$MAGICK_HOME/lib/
- 8a) # check configuration:
  convert -list configure
- 8b) # check image format supported:
  identify -list format
- Tips:
  Prior to 4), please make sure libjpeg and libpng are installed. If NOT, for Mac try this:
  brew install libjpeg libpng
  To check whether ImageMagick does work, please get additional information from:
  identify -list format
  convert -list configure
  On details, please refer to http://www.imagemagick.org/script/advanced-unix-installation.php

See Also

visNetMul

Examples

```
## Not run:
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/100)

# 2) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)

# 3) visualise the module with vertices being color-coded by scores
nnodes <- vcount(subg)
nsamples <- 10
data <- matrix(runif(nnodes*nsamples), nrow=nnodes, ncol=nsamples)
rownames(data) <- V(subg)$name
# output as a <a href="visNetAnimate.pdf">pdf</a> file
visNetAnimate(g=subg, data=data, filetype="pdf")
# output as a <a href="visNetAnimate.mp4">mp4</a> file but with dynamic layout
visNetAnimate(g=subg, data=data, filetype="mp4", glayout.dynamics=TRUE)
```

```
# output as a <a href="visNetAnimate.gif">gif</a> file but with dynamic layout
visNetAnimate(g=subg, data=data, filetype="gif", glayout.dynamics=TRUE)

## End(Not run)
```

---

visNetArc *Function to visualise an igraph object via arc diagram*

Description

visNetArc is supposed to visualise a graph object of class "igraph" via arc diagram in one-dimensional layout. More precisely, it displays vertices (nodes) along an axis, with edges linked by arcs. With proper ordering of vertices (e.g. according to communities and degrees), arc diagram is able to identify clusters and bridges (as effective as two-dimensional layout). One advantage of using arc diagram is to allow for easy annotations along vertices.

Usage

```
visNetArc(
g,
orientation = c("vertical", "horizontal"),
newpage = T,
ordering = NULL,
labels = V(g)$name,
vertex.label.color = "black",
vertex.label.cex = 1,
vertex.color = "transparent",
vertex.frame.color = "black",
vertex.size = log(degree(g)) + 0.1,
vertex.pch = 21,
vertex.lwd = 1,
edge.color = "grey",
edge.width = 1,
edge.lty = 1,
...
)
```

Arguments

g	an object of class "igraph"
orientation	the orientation of the plots. It can be either "vertical" (default) or "horizontal"
newpage	logical to indicate whether to open a new page. By default, it sets to true for opening a new page
ordering	a numeric vector about the ordering of vertices. It is optional. It is highly recommend to order vertices according to communities and degrees
labels	the label of the vertices. The default vertex labels are the name attribute of the nodes

vertex.label.color
: the color of vertex labels

vertex.label.cex
: the font size of vertex labels

vertex.color
: the fill color of the vertices. The default vertex colors are transparent

vertex.frame.color
: the color of the frame of the vertices. The default vertex frame colors are black

vertex.size
: the size of each vertex. By default, it is decided according to node degrees

vertex.pch
: the shape of each vertex. Either an integer specifying a symbol or a single character to be used as the default in plotting points. See http://www.statmethods.net/advgraphs/parameters.html

vertex.lwd
: line width for the vertices (default 1)

edge.color
: the color of the edges (default "grey")

edge.width
: line width for the edges (default 1)

edge.lty
: line type for the edges (default 1)

...
: additional graphic parameters associated with 'mtext'

Value

invisible

Note

none

See Also

visNet

Examples

```
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/80)

# 2) produce the induced subgraph only based on the nodes in query
g <- dNetInduce(g, V(g), knn=0)

# 3) color nodes according to communities identified via a spin-glass model and simulated annealing
com <- spinglass.community(g, spins=4)
vgroups <- com$membership
palette.name <- visColormap(colormap="rainbow")
vcolors <- palette.name(length(com))[vgroups]

# 4) size nodes according to degrees
vdegrees <- igraph::degree(g)

# 5) sort nodes: first by communities and then degrees
tmp <- data.frame(ind=1:vcount(g), vgroups, vdegrees)
```

```
ordering <- tmp[order(vgroups,vdegrees),]$ind

# 6) visualise graph using 1-dimensional arc diagram
visNetArc(g, ordering=ordering, labels=V(g)$name,
vertex.label.color=vcolors,
vertex.color=vcolors, vertex.frame.color=vcolors,
vertex.size=log(vdegrees)+0.1)

# 7) as comparison, also visualise graph on 2-dimensional layout
visNet(g, colormap="bwr", layout=layout.kamada.kawai(g),
vertex.label=V(g)$name,
vertex.color=vcolors, vertex.frame.color=vcolors,
vertex.shape="sphere")
```

`visNetCircle` *Function to visualise an igraph object via circle diagram*

Description

`visNetCircle` is supposed to visualise a graph object of class "igraph" via circle diagram. For better visualisation, ordering of vertices is determined according to communities and degrees.

Usage

```
visNetCircle(
g,
com,
circles = c("single", "multiple"),
newpage = T,
ordering = NULL,
colormap = c("rainbow", "bwr", "jet", "gbr", "wyr", "br", "yr", "wb"),
vertex.label = V(g)$name,
vertex.size = log(igraph::degree(g)) + 2,
vertex.label.color = "black",
vertex.label.cex = 0.6,
vertex.label.dist = 0.75,
vertex.shape = "sphere",
edge.width = 1,
edge.lty = 1,
edge.color.within = "grey",
edge.color.crossing = "black",
mark.shape = 1,
mark.expand = 10,
...
)
```

Arguments

g	an object of class "igraph"
com	an object of class "communities" (see http://igraph.org/r/doc/communities.html)
circles	how circles are drawn in the plot. It can be either "single" for all communities being drawn in a single circle (by default) or "multiple" for communities being drawn in the different circles (i.e. one circle per community)
newpage	logical to indicate whether to open a new page. By default, it sets to true for opening a new page
ordering	a numeric vector about the ordering of vertices. It is optional. It is highly recommend to order vertices according to communities and degrees
colormap	short name for the colormap. It can be one of "jet" (jet colormap), "bwr" (blue-white-red colormap), "gbr" (green-black-red colormap), "wyr" (white-yellow-red colormap), "br" (black-red colormap), "yr" (yellow-red colormap), "wb" (white-black colormap), and "rainbow" (rainbow colormap, that is, red-yellow-green-cyan-blue-magenta). Alternatively, any hyphen-separated HTML color names, e.g. "blue-black-yellow", "royalblue-white-sandybrown", "darkgreen-white-darkviolet". A list of standard color names can be found in http://html-color-codes.info/color-names
vertex.label	the label of the vertices. The default vertex labels are the name attribute of the nodes
vertex.size	the size of each vertex. By default, it is decided according to node degrees
vertex.label.color	the color of vertex labels
vertex.label.cex	the font size of vertex labels
vertex.label.dist	the distance of the label from the center of the vertex. If it is 0 then the label is centered on the vertex. If it is 1 then the label is displayed beside the vertex.
vertex.shape	the shape of each vertex. It can be one of "circle", "square", "csquare", "rectangle", "crectangle", "vrectangle", "pie" (http://igraph.org/r/doc/vertex.shape.pie.html), "sphere", and "none". If it sets to NULL, these vertices with negative will be "csquare" and the rest "circle".
edge.width	line width for the edges (default 1)
edge.lty	line type for the edges (default 1)
edge.color.within	the color for edges within a community (default "grey")
edge.color.crossing	the color for edges between communities (default "black")
mark.shape	a numeric scalar or vector controlling the smoothness of the vertex group marking polygons. Its possible values are between -1 (fully polygons) and 1 (fully smoothness)
mark.expand	a numeric scalar or vector, the size of the border around the marked vertex groups
...	additional graphic parameters. See http://igraph.org/r/doc/plot.common.html for the complete list.

Value

invisible

Note

none

See Also

visNet

Examples

```
## Not run:
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/80)

# 2) produce the induced subgraph only based on the nodes in query
g <- dNetInduce(g, V(g), knn=0)

# 3) color nodes according to communities identified via a spin-glass model and simulated annealing
com <- spinglass.community(g, spins=4)
vgroups <- com$membership
palette.name <- visColormap(colormap="rainbow")
mcolors <- palette.name(length(com))
vcolors <- mcolors[vgroups]

# 4) size nodes according to degrees
vdegrees <- igraph::degree(g)

# 5) sort nodes: first by communities and then degrees
tmp <- data.frame(ind=1:vcount(g), vgroups, vdegrees)
ordering <- tmp[order(vgroups,vdegrees),]$ind

# 6) visualise graph using circle diagram
# 6a) drawn into a single circle
visNetCircle(g=g, colormap="bwr", com=com, ordering=ordering)

# 6b) drawn into multlpe circles (one circle per community)
visNetCircle(g=g, colormap="bwr", com=com, circles="multiple",
ordering=ordering)

# 7) as comparison, also visualise graph on 2-dimensional layout
mark.groups <- communities(com)
mark.col <- visColoralpha(mcolors, alpha=0.2)
mark.border <- visColoralpha(mcolors, alpha=0.2)
edge.color <- c("grey", "black")[crossing(com,g)+1]
visNet(g, colormap="bwr", glayout=layout.fruchterman.reingold,
vertex.color=vcolors,
vertex.frame.color=vcolors, vertex.shape="sphere",
mark.groups=mark.groups, mark.col=mark.col,
mark.border=mark.border, mark.shape=1, mark.expand=10,
```

```
edge.color=edge.color)

## End(Not run)
```

---

visNetMul *Function to visualise the same graph but with multiple graph node colorings according to input data matrix*

Description

visNetMul is supposed to visualise the same graph but with multiple colorings according to input data matrix

Usage

```
visNetMul(
g,
data,
height = 7,
margin = rep(0.1, 4),
border.color = "#EEEEEE",
colormap = c("bwr", "jet", "gbr", "wyr", "br", "yr", "rainbow", "wb"),
ncolors = 40,
zlim = NULL,
colorbar = T,
colorbar.fraction = 0.25,
newpage = T,
glayout = layout.fruchterman.reingold,
mtext.side = 3,
mtext.adj = 0,
mtext.cex = 1,
mtext.font = 2,
mtext.col = "black",
...
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
data	an input data matrix used to color-code vertices/nodes. One column corresponds to one graph node coloring. The input matrix must have row names, and these names should include all node names of input graph, i.e. V(g)$name, since there is a mapping operation. After mapping, the length of the patern vector should be the same as the number of nodes of input graph. The way of how to color-code is to map values in the pattern onto the whole colormap (see the next arguments: colormap, ncolors, zlim and colorbar)
height	a numeric value specifying the height of device

margin	margins as units of length 4 or 1
border.color	the border color of each figure
colormap	short name for the colormap. It can be one of "jet" (jet colormap), "bwr" (blue-white-red colormap), "gbr" (green-black-red colormap), "wyr" (white-yellow-red colormap), "br" (black-red colormap), "yr" (yellow-red colormap), "wb" (white-black colormap), and "rainbow" (rainbow colormap, that is, red-yellow-green-cyan-blue-magenta). Alternatively, any hyphen-separated HTML color names, e.g. "blue-black-yellow", "royalblue-white-sandybrown", "darkgreen-white-darkviolet". A list of standard color names can be found in http://html-color-codes.info/color-names
ncolors	the number of colors specified over the colormap
zlim	the minimum and maximum z/patttern values for which colors should be plotted, defaulting to the range of the finite values of z. Each of the given colors will be used to color an equispaced interval of this range. The midpoints of the intervals cover the range, so that values just outside the range will be plotted
colorbar	logical to indicate whether to append a colorbar. If pattern is null, it always sets to false
colorbar.fraction	the relative fraction of colorbar block against the figure block
newpage	logical to indicate whether to open a new page. By default, it sets to true for opening a new page
glayout	either a function or a numeric matrix configuring how the vertices will be placed on the plot. If layout is a function, this function will be called with the graph as the single parameter to determine the actual coordinates. This function can be one of "layout.auto", "layout.random", "layout.circle", "layout.sphere", "layout.fruchterman.reingold", "layout.kamada.kawai", "layout.spring", "layout.reingold.tilford", "layout.fruchterman.reingold.grid", "layout.lgl", "layout.graphopt", "layout.svd" and "layout.norm". A full explanation of these layouts can be found in http://igraph.org/r/doc/layout_nicely.html
mtext.side	on which side of the mtext plot (1=bottom, 2=left, 3=top, 4=right)
mtext.adj	the adjustment for mtext alignment (0 for left or bottom alignment, 1 for right or top alignment)
mtext.cex	the font size of mtext labels
mtext.font	the font weight of mtext labels
mtext.col	the color of mtext labels
...	additional graphic parameters. See http://igraph.org/r/doc/plot.common.html for the complete list.

Value

invisible

Note

none

See Also

visNet, visNetAnimate

Examples

```
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/80)

# 2) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)

# 3) visualise the module with vertices being color-coded by scores
nnodes <- vcount(subg)
nsamples <- 10
data <- matrix(runif(nnodes*nsamples), nrow=nnodes, ncol=nsamples)
rownames(data) <- V(subg)$name
visNetMul(g=subg, colormap="bwr", data=data,
glayout=layout.fruchterman.reingold)
```

---

visNetReorder *Function to visualise the multiple graph colorings reorded within a sheet-shape rectangle grid*

Description

visNetReorder is supposed to visualise the multiple graph colorings reorded within a sheet-shape rectangle grid

Usage

```
visNetReorder(
g,
data,
sReorder,
height = 7,
margin = rep(0.1, 4),
border.color = "#EEEEEE",
colormap = c("bwr", "jet", "gbr", "wyr", "br", "yr", "rainbow", "wb"),
ncolors = 40,
zlim = NULL,
colorbar = T,
colorbar.fraction = 0.5,
newpage = T,
glayout = layout.fruchterman.reingold,
mtext.side = 3,
mtext.adj = 0,
mtext.cex = 1,
mtext.font = 2,
```

```
mtext.col = "black",
...
)
```

Arguments

g	an object of class "igraph" or "graphNEL"
data	an input data matrix used to color-code vertices/nodes. One column corresponds to one graph node coloring. The input matrix must have row names, and these names should include all node names of input graph, i.e. V(g)$name, since there is a mapping operation. After mapping, the length of the pattern vector should be the same as the number of nodes of input graph. The way of how to color-code is to map values in the pattern onto the whole colormap (see the next arguments: colormap, ncolors, zlim and colorbar)
sReorder	an object of class "sReorder"
height	a numeric value specifying the height of device
margin	margins as units of length 4 or 1
border.color	the border color of each figure
colormap	short name for the colormap. It can be one of "jet" (jet colormap), "bwr" (blue-white-red colormap), "gbr" (green-black-red colormap), "wyr" (white-yellow-red colormap), "br" (black-red colormap), "yr" (yellow-red colormap), "wb" (white-black colormap), and "rainbow" (rainbow colormap, that is, red-yellow-green-cyan-blue-magenta). Alternatively, any hyphen-separated HTML color names, e.g. "blue-black-yellow", "royalblue-white-sandybrown", "darkgreen-white-darkviolet". A list of standard color names can be found in http://html-color-codes.info/color-names
ncolors	the number of colors specified over the colormap
zlim	the minimum and maximum z/patttern values for which colors should be plotted, defaulting to the range of the finite values of z. Each of the given colors will be used to color an equispaced interval of this range. The midpoints of the intervals cover the range, so that values just outside the range will be plotted
colorbar	logical to indicate whether to append a colorbar. If pattern is null, it always sets to false
colorbar.fraction	the relative fraction of colorbar block against the figure block
newpage	logical to indicate whether to open a new page. By default, it sets to true for opening a new page
glayout	either a function or a numeric matrix configuring how the vertices will be placed on the plot. If layout is a function, this function will be called with the graph as the single parameter to determine the actual coordinates. This function can be one of "layout.auto", "layout.random", "layout.circle", "layout.sphere", "layout.fruchterman.reingold", "layout.kamada.kawai", "layout.spring", "layout.reingold.tilford", "layout.fruchterman.reingold.grid", "layout.lgl", "layout.graphopt", "layout.svd" and "layout.norm". A full explanation of these layouts can be found in http://igraph.org/r/doc/layout_nicely.html

mtext.side	on which side of the mtext plot (1=bottom, 2=left, 3=top, 4=right)
mtext.adj	the adjustment for mtext alignment (0 for left or bottom alignment, 1 for right or top alignment)
mtext.cex	the font size of mtext labels
mtext.font	the font weight of mtext labels
mtext.col	the color of mtext labels
...	additional graphic parameters. See http://igraph.org/r/doc/plot.common.html for the complete list.

Value

invisible

Note

none

See Also

visNet, dNetReorder

Examples

```
# 1) generate a random graph according to the ER model
g <- erdos.renyi.game(100, 1/100)

# 2) produce the induced subgraph only based on the nodes in query
subg <- dNetInduce(g, V(g), knn=0)

# 3) reorder the module with vertices being color-coded by input data
nnodes <- vcount(subg)
nsamples <- 10
data <- matrix(runif(nnodes*nsamples), nrow=nnodes, ncol=nsamples)
rownames(data) <- V(subg)$name
sReorder <- dNetReorder(g=subg, data, feature="node",
node.normalise="none")

# 4) visualise the module with vertices being color-coded by input data
visNetReorder(g=subg, colormap="bwr", data=data, sReorder)
```

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